We are IntechOpen, the world’s leading publisher of Open Access books
Built by scientists, for scientists

6,600
Open access books available

177,000
International authors and editors

195M
Downloads

154
Countries delivered to

TOP 1%
Our authors are among the most cited scientists

12.2%
Contributors from top 500 universities

WEB OF SCIENCE™
Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com
Mechanism of In-Situ Catalytic Cracking of Biomass Tar over Biochar with Multiple Active Sites

Dongdong Feng, Yu Zhang, Yijun Zhao and Shaozeng Sun

Abstract

Biomass tar is the bottleneck in the development of efficient utilization of biomass syngas. The in-situ catalytic cracking biomass tar with multi-active biochar is investigated in a two-stage fluidized bed-fixed bed reactor. It indicates that adding H$_2$O or CO$_2$ is found to improve the homogeneous and heterogeneous cracking of biomass tar. Activation of biochar by H$_2$O or CO$_2$ impacted the morphology of biochar surface and distribution of metal species. H$_2$O or CO$_2$ affects the creation and regeneration of pore structures, influencing the biochar structure and dynamical distribution of alkali and alkaline earth metal species (AAEMs), which ensure enough surface active sites to maintain the catalytic activity of biochar. The tar cracking into low-quality tar or small-molecule gas may be catalyzed by K, while the combination of tar with biochar would be promoted by Ca. The volatilizations of K and Ca, due to their reaction with volatiles, are to a large extent in accordance with their valences and boiling points. The subsequent transformation from the small aromatic ring systems to the larger ones occurs due to the volatile-biochar interaction. During tar cracking over biochar, K and Ca act as the active sites on biochar surface to promote the increase of active intermediates (C▬O bonds and C▬O▬K/Ca).

Keywords: biochar, tar, catalytic cracking, AAEM species

1. Introduction

Tar is a generic term comprising all organic compounds present in syngas except for gaseous hydrocarbons. Tars can condense to more complex structures in pipes, filters, or heat exchangers of downstream equipment and processes, which may cause mechanical breakdown of the entire system [1]. For biomass gasification, the allowable limit for tar in the producer gas is less than 5 mg/Nm$^3$ for a direct-fired gas turbine [2], and for some fuel synthesis processes, the contents of tar and ammonia are required to be <0.1 mg/Nm$^3$ and <10 ppm, respectively [3–5], in order to protect the catalysts and downstream equipment and to improve the overall efficiency and economics. The tar mixture is classified into five classes by Padban [6]: undetectable, heterocyclic, light aromatic hydrocarbons (LAHs), light polyaromatic hydrocarbons (LPAHs), and heavy polyaromatic hydrocarbons (HPAHs). The removal of biomass tar is one of the main challenges for the biomass gasification industry [7, 8]. Catalytic cracking is a known method for the efficient removal of biomass tar [9–12]. Biochar, as a product of pyrolysis and gasification of biomass, is a relatively
cheap catalyst with high activity in tar heterogeneous cracking [13–20]. During tar catalytic cracking over biochars, even after the loss of catalytic activity through coking, the biochar samples can still be directly combusted, so as to recover the chemical energy of catalyst, thus avoiding any reprocessing as a result of deactivation.

In addition to the analysis of model tar compounds [21–23], studies of biomass tar over biochar mainly discuss the reforming of real tar from raw materials [24, 25]. However, the AAEM species (e.g., Na, K, Mg, and Ca) in raw biomass play an important role as the “cross points” during tar formation. The chemical bonds between AAEM species and the carbon matrix are repeatedly breaking and reforming. This process promotes the production of gaseous products from the fatty acid tar and a degree of small aromatic compounds. Simultaneously, larger aromatic ring compounds (≥5 aromatic ring system) are formed within the biochar structure [26, 27]. The presence of AAEM species can inhibit the release of volatile matter (especially for biomass tar)—even the strong interaction between volatile materials and biochar will affect tar composition, leading to the catalytic conversion of the real tar components before contacting with the catalyst, which misleads mechanistic studies of subsequent heterogeneous reforming over biochar catalyst.

The formation (e.g., 500–700°C) and thermal decomposition (e.g., 700–900°C) of tar during the gasification process are an extremely complex multistep reaction [28–32], which involves not only homogeneous conversion, but also heterogeneous reforming. H₂O and CO₂ are two important reforming agents [33] in the biomass gasification industry. Studying the influence of H₂O and CO₂ on tar homogeneous transformation and heterogeneous reformation is valuable to better understand the analysis of the tar complex gas-solid phase reaction. However, there is still less research on separate discussion between homogeneous conversion and heterogeneous reforming of biomass tar over biochar. Although there are reports detailing the influence of H₂O and CO₂ on tar during the gasification process [34–39], they were mainly focused on the single concentration of reforming agent (15 vol.% H₂O or pure CO₂ atmosphere). There has yet to be detailed a complete understanding of the influence of H₂O and CO₂ on the homogeneous conversion and heterogeneous reforming over biochar as a function of biomass tar evolution.

The effects of reforming agent concentration and reaction temperature on the tar homogeneous conversion and heterogeneous reforming over biochar were investigated in a two-stage fluidized bed/fixed bed reactor. The H-form biomass samples (with little AAEM species) were used to provide the real tar components, which effectively inhibited the tar-AAEM interactions in gas phase during H₂O/CO₂ homogeneous conversion and prevented any secondary catalytic effects of AAEM species from the volatilization of raw materials on the biochar catalyst surface. The analysis of biochar structures examined with Raman spectroscopy to comprehensively elucidate the changes of biochar catalyst structure after the H₂O and CO₂ heterogeneous reforming of biomass tar. In addition to the measurement of tar yields, GC/MS spectroscopy was used to characterize the detailed structural features of tar [40], so as to understand the molecular biomass tar transformation pathway and the coupling mechanism (e.g., collaboration and interaction effects) between the biochar structure and the AAEM species during tar reforming.

2. Experiment

2.1 Material preparation

Biomass (rice husks) obtained from the Wu Chang area in Harbin, Heilongjiang Province, China, was used in the experiments. The samples were dried overnight.
at 105°C, pulverized, and sieved to obtain a fraction with particle sizes between 0.15 and 0.25 mm. The proximate and ultimate analyses data [41] for the rice husk samples are listed in Table 1, which could be used to characterize the composition of biomass, grasping its reaction characteristics and application value (M: moisture, A: ash, V: volatile, FC: fixed carbon; C: carbon, H: hydrogen, O: oxygen, N: nitrogen, S: sulfur).

The H-form rice husk was used as the raw material to supply real biomass tar. The raw pyrolysis biochar was mixed with an aqueous solution of 0.2 M H$_2$SO$_4$ in an acid solution:sample mass ratio of 30:1 and stirred in an argon atmosphere for 24 h. The slurry was filtered and washed with deionized water until the filtrate pH was constant (pH ≈ 7). After drying, the acid-washed sample is termed as the H-form char.

### 2.2 Biochar catalyst preparation

Pyrolysis biochar was used as the catalyst for biomass tar reforming. The set-up to pyrolysis biochar comprises a quartz reactor and a standard muffle furnace, as shown in Figure 1.

The quartz tray (red tray) with 5.0 g raw rice husk was placed into the quartz reactor. Along with the reactor cover, the quartz reactor was placed into the muffle furnace. At room temperature, the air in the reactor was displaced by Ar at a rate of 2.0 L/min for 30 min. Pyrolysis was performed at a slow-heating rate of 10°C/min up to a final pyrolysis temperature of 700°C with 70 min. Thereafter, with the temperature of turn-off furnace back to room temperature, the door of the muffle furnace was opened, and the reaction quenched by removing the reactor from the furnace. Ar gas was passed continuously through the reactor to prevent oxidation during cooling. The pyrolyzed biochar was removed from the reactor and stored at 4°C. Ar gas is supplied through a gas pipe (400 mm, long) into the porous distributor (with a diameter of 120 mm) and fed from the bottom of the quartz reactor filling the entire reactor. The upper cover acts as a partial seal under the action of its own gravity; however, with increasing internal gas volume produced as a function

<table>
<thead>
<tr>
<th>Sample</th>
<th>Proximate analysis (wt.%)</th>
<th>Ultimate analysis (wt.%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$M_{ad}$</td>
<td>$A_{ad}$</td>
</tr>
<tr>
<td>Rice husk</td>
<td>6.86</td>
<td>17.00</td>
</tr>
</tbody>
</table>

Table 1. Proximate and ultimate analyses of rice husk samples.

![Image](image1.png)
of muffle furnace temperature, some gaps between the upper cover and the reactor allow the release of gases under internal positive pressure. The volatile matters formed during the volatilization of biomass were rapidly dispersed away from the reactor, carried by Ar gas, so as to ensure an inert atmosphere inside the reactor. The metal contents of the origin and H-form biochar are listed in Table 2.

2.3 Homogeneous/heterogeneous reforming of biomass tar

As shown in Figure 2, a two-stage fluidized bed/fixed bed reactor was used for the investigation of the homogeneous conversion and heterogeneous reforming of biomass tar over biochar. The inner diameter of the reactor was 37 mm. The reactor is divided into two layers by four quartz frits. The upper layer is fixed bed, while the lower is fluidized bed. The heights of upper and lower layers are 30 and

<table>
<thead>
<tr>
<th>Biochar</th>
<th>Primary metal contents (wt.%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Na</td>
</tr>
<tr>
<td>Origin biochar</td>
<td>0.03</td>
</tr>
<tr>
<td>H-form biochar</td>
<td>0.00</td>
</tr>
</tbody>
</table>

Table 2. Primary metal contents of pyrolysis rice husk biochar samples.

![Figure 2. Schematic diagram of a two-stage fluidized bed/fixed bed reactor for the homogeneous conversion and heterogeneous reforming of biomass tar.](image)
130 mm, respectively. For the homogeneous conversion of tar, the lower fluidized bed reactor was heated to 500°C, with the temperature increased to 500–900°C for the upper fixed bed reactor (without catalyst). The silica sand with the weight of 60 g was pre-loaded into the bottom stage of the quartz reactor followed by Ar purging (1.0 L/min carrier gas and 1.5 L/min fluidizing gas) before heating the desired temperature. Once stabilization of the temperatures was achieved, the H-form rice husk was injected into the fluidized bed through the water-cooled pipes at a feeding rate of 100 mg/min. Simultaneously, for the CO₂ or H₂O separate treatment, the atmosphere was switched to CO₂ (29 vol.% or H₂O (15 vol.%) through a dedicated gas injection system located in between the lower and upper reactors as shown in Figure 2. Pure Ar gas was also injected into the dedicated gas injection system, to balance the system, at rates of 1.03/0.92/0.82/0.75/0.68 L/min for reaction temperatures of 500/600/700/800/900°C, respectively, to maintain constant residence times for each reforming temperature. For 15 vol.% H₂O, steam injection was achieved by feeding a metered amount of water through a high-performance liquid chromatography (HPLC) pump into the heated zone of the reactor where the water was evaporated into steam. De-ionized H₂O was injected at rates of 0.34/0.30/0.28 ml/min along with 0.40/0.36/0.33 L/min of balanced Ar for the 700/800/900°C reaction temperatures, respectively. CO₂ was injected through the dedicated gas injection system at rates of 0.82/0.75/0.68 L/min to achieve 29 vol.% for 700/800/900°C reaction temperatures, respectively. The temperature was held for 10 min for each reaction. Reactions were terminated by switching the atmosphere to argon and removing the reactor out of the furnace.

For the heterogeneous reforming of biomass tar over biochar activated by H₂O or CO₂, the activation of biochar was carried out for 10 min in the fixed-bed zone in a 15 vol.% H₂O or a 29 vol.% CO₂ atmosphere with no supplemental H-form rice husk added to the fluidized-bed zone. This was followed by another 10 min at 800°C in an Ar atmosphere, to maintain the same total reaction time (20 min) as the tar-reforming conditions. Details of the five experimental conditions (A–E) are shown in Table 3. The experiments involved three pyrolysis experiments: tar reforming (A) in Ar with unactivated pyrolysis biochar; (B) in Ar over H₂O-activated biochar; and (C) in Ar over CO₂-activated biochar. In (B) and (C), a 10-min activation of the biochar was first carried out with the activated biochar then used for 10 min of tar reforming in an Ar atmosphere at 800°C, while H-form biomass was also fed to the reactor. In addition, two gasification experiments were carried out: tar reforming in (D) a 15 vol.% H₂O atmosphere over H₂O-activated biochar and in (E) a 29 vol.% CO₂ atmosphere over CO₂-activated biochar. In (D) and (E), both atmospheres were kept constant for 20 min even though the period was evenly divided into a biochar-activation stage, which was followed by tar reforming over biochar.

<table>
<thead>
<tr>
<th>Conditions</th>
<th>No.</th>
<th>Conditions of biomass tar reforming over biochar at 800°C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pyrolysis</td>
<td>A</td>
<td>Biomass tar reforming in Ar over pyrolysis biochar</td>
</tr>
<tr>
<td></td>
<td>B</td>
<td>Biomass tar reforming in Ar over H₂O-activated biochar</td>
</tr>
<tr>
<td></td>
<td>C</td>
<td>Biomass tar reforming in Ar over CO₂-activated biochar</td>
</tr>
<tr>
<td>Gasification</td>
<td>D</td>
<td>Biomass tar H₂O reforming over H₂O-activated biochar</td>
</tr>
<tr>
<td></td>
<td>E</td>
<td>Biomass tar CO₂ reforming over CO₂-activated biochar</td>
</tr>
</tbody>
</table>

Table 3. Experimental conditions investigated for tar reforming over biochar.
2.4 Sampling and analysis of biochar and tar

The biomass tar compounds were trapped in two gas bottles, connected in series, and filled with a mixture of HPLC-grade chloroform and methanol (4:1, v/v), as shown in Figure 2. The bottles were placed in a brine ice bath (≤0°C). After the reaction, the total solution was transferred to a 200 mL volumetric flask and made up to volume with a mixture of chloroform and methanol (4:1, v/v).

The tar yield was determined by evaporating the solvents and water at 35°C for 4 h. The tar is thus experimentally defined as the material soluble in the chloroform/methanol (4:1, v/v) solvent mixture not being evaporated (with the solvents) at 35°C within 4 h [24, 25, 42]. The residues in the solvents themselves (i.e., blank) and the biomass moisture content were considered in the tar yield calculation. The equation used for tar concentration in the solution is shown as follows Eq. (1):

\[
C = \frac{C_2 - C_1}{1 - C_2}
\]

where \(C\) is the concentration of tar; \(C_1\) is the concentration of the mixed solution residue (blank experiment); and \(C_2\) is the concentration of residue in tar solution.

The equation used to determine tar yield is shown as follows Eq. (2):

\[
\text{Tar yield} = \frac{C \times M_{\text{tar solution}}}{M_{\text{biomass}}}
\]

where \(C\) is the concentration of tar in the solution; \(M_{\text{tar solution}}\) is the total mass of tar collected in the solution; and \(M_{\text{biomass}}\) is the feed quality of the H-form rice husk into the reactor.

The samples were analyzed using an Agilent Gas Chromatography Mass Spectrometer (GC-MS) instrument (6890 series GC with a 5973 MS detector) with a capillary column (DB-5 ms; length 30 m, internal diameter 0.25 mm, film thickness 0.5 μm). The sample solution (5 μL) was injected into the injection port, set at 260°C, with a split ratio of 80:1. The column was operated in constant-flow mode using 2.0 mL/min of helium as the carrier gas. The column temperature was initially maintained at 35°C for 3 min, then increased to 260°C at a heating rate of 10°C/min, and then maintained at 260°C for 5 min. Mass spectra were acquired after a 4-min solvent delay [21]. The chromatogram peaks were identified by comparison with the standard spectra of compounds in the National Institute of Standards and Technology library (NIST) and/or from the retention times/spectra of known injected species.

The total amount of metal species in the biochar samples was quantified by employing a previously established procedure [43]. Using a microwave system (Ethos 1, Milestone, Sorisole, Italy), the sample (0.1 g) was digested in a 1:3:8 (v/v/v) mixture of 40% HF, 30% H₂O₂, and 65% HNO₂ at 200°C for 60 min. The metal species content was then quantified by inductively coupled plasma-atomic emission spectroscopy (ICP-AES). Three measurements were conducted with the average values and then taken as the results.

The biochar’s particle morphology and surface composition were measured by an EVO18 scanning electron microscope coupled to an energy dispersive X-ray spectrometer (SEM-EDX, Carl Zeiss, Germany).

Biochar samples were set for at least 24 h to displace the reaction gas within the pore structure with air. N₂-adsorption isotherms were then obtained at −196°C (ASAP 2020M, Micromeritics Instrument Crop, USA) and analyzed by the BET model to determine the sample’s surface area and pore volume.

XPS analysis was used to evaluate the characteristics of surface elements in biochar. This was performed using a K-Alpha spectrometer (Thermo Fisher Scientific).
equipped with monochromatic Al Kα X-rays at 1486.6 eV. To exclude effects on the binding energies caused by changing the sample during measurements, the data were corrected by a linear shift with the maximum peak of the C1s binding energy of the adventitious carbon corresponding to 284.6 eV. The surface's elemental condition was analyzed using the number of escaped electrons from the char surface at a depth of 1–10 nm, according to the findings in our previous investigations [44, 45].

3. Results and discussion

3.1 Homogeneous conversion of biomass tar

The homogeneous conversion of tar mainly refers to the initial pyrolysis tar experiences during a series of decomposition and polymerization processes under ambient conditions (heat and atmosphere). Tar yield during the Ar, H₂O, and CO₂ homogeneous conversion experiments performed at 500–900°C can be seen in Figure 3. In the presence of an Ar-only atmosphere, the tar yield decreased gradually as a function of increasing temperature from 500 (26.18%) to 900°C (6.38%). Temperature in the Ar-only experiments has a greater influence between 500 and 700°C. Further increasing the temperature to 700–900°C results in increased biomass decomposition, thus lowering tar yields. Thermal decomposition is considered to be the main factor in the conversion of tar [46, 47]. As shown in Figure 3, in 15 vol.% H₂O and 29 vol.% CO₂, the effects of H₂O and CO₂ on the homogeneous transformation of biomass tar over biochar are significant. At the same temperature, the tar reforming effect of 15 vol.% H₂O is significantly higher than that of 29 vol.% CO₂. In 15 vol.% H₂O, the tar yield decreased from 6.95% at 700°C to 3.56% at 900°C. In 29 vol.% CO₂, the tar yield decreased from 7.99% at 700°C to 5.01% at 900°C. For the higher temperatures, 700–900°C are required for H₂O and CO₂ to influence tar homogeneous transformations, while for the lower temperatures, 500 and 600°C are not in the gasification thermal range.

As shown in Figure 4, it can be seen that at lower temperatures (500–600°C), the majority of the biomass tar still comprises components based on the primary biomass tar containing oxygen and substituent compounds, such as levoglucosan and dimethoxymethane. However, when subjecting the biomass to higher temperatures (700–900°C), most of the primary pyrolyzed tar gradually transforms [48]. The tar composition seems to be mainly composed of aromatic compounds having

![Figure 3. Tar yield during homogeneous conversion at 500–900°C.](image-url)
good thermal stability, such as toluene, indene, and naphthalene, among others. Increasing the temperature resulted in either a gradual reduction or a complete removal of tars containing branched or heteroatom compounds, and polycyclic aromatic hydrocarbons (PAHs) were gradually formed. For the biomass tar homogeneous conversion, the aromatic ring structure has higher thermal stability than that of the non-aromatic structures. Specific tar components decompose into small molecular gases and C$_1$–C$_5$ hydrocarbons, while there is evidence for the promotion of aromatic rings as a function of increasing temperature. H-abstraction, C$_2$H$_2$-addition (HACA), and cyclodehydrogenation are the mechanisms typically responsible for such processes [49, 50]. Performing the reactions at the mid-temperature range (700–800°C) results in aromatic conversion with oxygen and substituents. Thermal decomposition [51] and additional reactions convert short-chain hydrocarbons (C$_1$–C$_5$) into compounds containing unsaturated double and triple bonds that gradually increase in concentration by the acetylene addition reaction. The aromatic components can also be polymerized by dehydrogenation. Further increasing the temperature to 900°C results in the relative content of PAHs, such as naphthalene, phenanthrene, and anthracene, to increase the above conversion pathway yielding highly stable aromatic hydrocarbons.

GC-MS analysis during H$_2$O and CO$_2$ homogeneous conversion at 700–900°C can be seen in Figure 5. At 700–900°C, H$_2$O and CO$_2$ have a degree of influence on the conversion of tar. The degree of tar homogeneous conversion in the presence of either a H$_2$O or CO$_2$ atmosphere was significantly higher than that of the thermal decomposition in Ar. PAH concentration was low. The results show that H$_2$O and CO$_2$ have obvious effects on the transformation of aromatics, especially PAHs [52]. The free radical theory is used to explain the homogeneous transformation of tar. The formation of aromatic radicals in the polymerization of aromatic hydrocarbons is considered to be the key to the reaction. The continuous polymerization process is considered to be the main pathway [53–56]. Thermal decomposition is a method of generating free radicals through thermal breaking of bonds. The free radicals generated by the original tar form different final products by reacting with different free radicals produced as a function of the atmosphere. The presence of H$_2$O and CO$_2$ promoted the formation of free radicals with H/O/OH moieties. The influence of temperature is mainly reflected in the promotion of the decomposition reaction caused by free radicals [57]. CO$_2$ is a pure oxygen donor. Figure 6 shows that the active oxygen atoms used for oxidative decomposition of hydrocarbons and intermediate
products are mainly produced by the reaction $\text{CO}_2 + e \rightarrow \text{CO} + \text{O} + e$. Active OH free radicals can be formed by replacing the hydrogen atoms in the hydrocarbons with oxygen atoms. Increasing the content of CO$_2$ is helpful to inhibit the cyclization of aromatics. The addition of CO$_2$ promotes the formation of free radicals such as O, which can further react with hydrocarbon groups. The oxidation reaction of active oxygen atoms with hydrocarbons forms CO, H$_2$O, and other products. The oxidative cracking process of tar is initiated, and the polymerization process of aromatic
hydrocarbons is also inhibited. \( \text{H}_2 \text{O} \) not only promotes tar cracking conversion but also inhibits the polymerization reaction. This is related to the higher activity of free radical formation being a more active reformer in the conversion of tar. \( \text{H}_2 \text{O} \) and \( \text{CO}_2 \) have similar oxidation capacities. The difference between the two is mainly reflected in the product—\( \text{H}_2 \text{O} \) produces higher numbers of \( \text{H} \) free radicals than \( \text{CO}_2 \) \[58\]. \( \text{O} \) and \( \text{OH} \) free radicals can be formed by ionization of \( \text{H}_2 \text{O} \) (\( \text{H}_2 \text{O} \rightarrow \text{H} + \text{OH} \)) in the presence of steam. The fracture of \( \text{OH} \) can form new \( \text{H} \) and \( \text{O} \) free radicals. The \( \text{H}_2 \text{O}/\text{OH} \) atoms in the gas phase exist in radical form. According to the free radical mechanism, the primary constituents of the biomass are broken into activated tar fragments at high temperatures. A large number of \( \text{H}/\text{OH} \) free radicals will combine with activated tar fragments before tar polymerization.

As shown in Figure 6, the conversion of the tar homogeneous transformation process is considered to be a two-stage process. The first stage involves the decomposition and transformation of the active heteroatom-containing groups in the tar, along with the decomposition of dealkylated side chains, hydrocarbon molecular cyclization, and aromatization reactions. The products include low-chain aliphatic hydrocarbons, oxygen-containing small molecular gases, and single-ring aromatic hydrocarbons. The second stage is the reforming of tar components; the dehydrogenation of cyclization products; the addition of acetylene; and the growth, recombination, and isomerization of aromatics. The two processes constitute the basis of the biomass tar homogeneous reaction. In the presence of different reforming agents (\( \text{H}_2 \text{O} \) or \( \text{CO}_2 \)), the atmosphere can promote or inhibit tar pyrolysis conversion, thus influencing the composition of the final tar. The addition of \( \text{H}_2 \text{O} \) and \( \text{CO}_2 \) can promote the generation of active free radicals such as \( \text{O} \), \( \text{OH} \), \( \text{H} \), and so on. These free radicals can react with the active free tar fragments generated from the first stage of thermal decomposition demonstrating the importance of the \( \text{H}_2 \text{O} \) and \( \text{CO}_2 \) reforming agents in the homogeneous conversion of biomass tar.

### 3.2 Heterogeneous reforming of biomass tar over biochar

#### 3.2.1 Biomass tar reforming

As shown in Figure 7, the highest proportion of bio-tar was reformed (including homogeneous and heterogeneous phases) in the 15 vol.% \( \text{H}_2 \text{O} \) atmosphere over \( \text{H}_2 \text{O} \)-activated biochar (D). The proportion of tar reformed in the 29 vol.% \( \text{CO}_2 \) atmosphere over \( \text{CO}_2 \)-activated biochar (E) was also considerably higher than results for reforming in an \( \text{Ar} \) atmosphere (A, B, and C). This illustrates that the presence of a gasification agent (\( \text{H}_2 \text{O}/\text{CO}_2 \)) greatly promotes in-situ reforming of nascent bio-tar over biochar. Under pyrolysis conditions, the homogeneous transformation of biomass tar was mainly based on secondary reactions (i.e., tar thermal cracking at 800°C), yielding a conversion efficiency of 70.86%. The ability of \( \text{H}_2 \text{O}/\text{CO}_2 \) activation to improve biochar reactivity was also clearly observed. In the \( \text{Ar} \) atmosphere, the highest proportion of tar was reformed over the \( \text{H}_2 \text{O} \)-activated biochar (B, 20.08%), followed by that over the \( \text{CO}_2 \)-activated biochar (C, 19.01%), while the lowest conversion was for the (unactivated) pyrolysis biochar (A, 17.41%). El-Rub and Kamel [59] and Chen et al. [60] studied biochar’s catalytic activity for tar reforming using a fixed char bed. They concluded that in an inert atmosphere, the tar molecules were mainly adsorbed on biochar active sites and converted into larger polyaromatic molecules and coke via a series of dehydrogenation, cyclization, and condensation reactions. Differences between the unactivated and \( \text{H}_2 \text{O}/\text{CO}_2 \)-activated biochars may be attributed to differences in inherent catalytic AAEM species (such as K and Ca) and the biochars’ physiochemical structures [61], as discussed later.
The presence of a gasification agent further improved the homogeneous reforming of the bio-tar (72.62 and 71.57% with H$_2$O and CO$_2$ present, respectively). The homogeneous transformation of biomass tar in the gas phase was in a certain extent in H$_2$O/CO$_2$ gasification condition, which was in broad agreement with the results obtained by Wang et al. [17] and Min et al. [24]. Besides, in the gasification conditions, H$_2$O and CO$_2$ further improved the biochar’s catalytic reactivity for the heterogeneous reforming process with the proportion reformed increasing from 20.08% (B) to 26.85% (D) with H$_2$O and from 19.01% (C) to 22.17% (E) with CO$_2$. According to Min et al. [24], the reforming of tar molecules over biochar may be activated in two ways. First, in the gas phase, nascent tar (volatiles) contains abundant free radicals that would react with extant tar molecules to form activated tar fragments. Second, tar molecules and biochar may be activated by H$_2$O/CO$_2$ during the chemisorption of tar on the biochar surface with reactions between tar and H$_2$O/CO$_2$ adsorbed on the biochar’s active sites then leading to further reforming reactions. Overall, gasification agents (H$_2$O and CO$_2$) improved and maintained the system’s ability to carry out homogeneous and heterogeneous reforming of biomass tar.

3.2.2 H$_2$O/CO$_2$ activation of biochar

The metal contents of the biochar samples are shown in Table 4. Apart from K, there was little difference (±0.02 wt.%) observed for the metals between the biochar samples. During the H$_2$O/CO$_2$ activation of biochar, K appears to have been released from the biochar, decreasing from 1.12 wt.% in the pyrolysis biochar to 1.06 wt.% when activated by H$_2$O and 1.09 wt.% in the CO$_2$-activated biochar.

<table>
<thead>
<tr>
<th>Biochar samples</th>
<th>Metal species content (wt.% in biochar)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>K</td>
</tr>
<tr>
<td>H$_2$O-activated biochar</td>
<td>1.06</td>
</tr>
<tr>
<td>CO$_2$-activated biochar</td>
<td>1.09</td>
</tr>
</tbody>
</table>

Table 4. Biochar samples’ metal-content analysis.
Because of their valence states [43], other metal species like Ca bond to the biochar more strongly (i.e., at two or more sites) than K (which only bonds at one site).

3.2.3 SEM-EDX analysis of biochar

As shown in Figure 8(a), the unactivated biochar particles’ surfaces showed more, larger (40 × 60 μm) hill-like structures than the surfaces of the activated biochars. H₂O and CO₂ activate the biochar via C + H₂O → CO + H₂ and C + CO₂ → 2CO, respectively. However, its larger size meant activation by CO₂ was limited to the surface of biochar, resulting in small structures (15 × 15 μm), which can be seen in Figure 8(c). However, as illustrated by the structure shown in Figure 8(b), H₂O, as well as H/O/OH radicals, was able to alter the surface morphology (creating structures of 20 × 20 μm) and infiltrate into the particle’s carbon matrix to produce new larger pore structures from the inside out. According to Wu et al. [62], interactions between radicals and metal species take place on the surface of internal pores or inside the char matrix. Table 4 shows little change in the biochar’s internal metal (K, Ca, Mg, Fe, and Al) contents before and after H₂O/CO₂ activation. Thus, the effect of H₂O/CO₂ may be more focused on changing the distribution of metal species within the biochar samples. As we reported previously [43], the effect of K in biochar on tar reforming is stronger than that of Ca and other species. Thus, the surface content and distribution of K were studied, as shown in Figure 8(a)–(c). The surface content of K significantly increased from 0.18% in the unactivated biochar to 0.35% in the H₂O-activated biochar and 0.21% in CO₂-activated biochar. In addition, an obvious enrichment occurred on the surface of H₂O-activated biochar. Klinghoffer et al. [63] reported that during thermal treatment the metal species migrated to the biochar surface, some of which formed clusters that then acted as an active site for catalytic reactions. During H₂O/CO₂ activation, an increase in surface O content occurred alongside the migration of AAEM species from the interior of the particles to the surface, forming metal-carbon complexes. The redox properties of these metal-carbon complexes may have had implications for the biochar’s catalytic properties. Also, highly dispersed metal species in a highly porous carbon matrix could have effectively acted as active adsorption sites that

![Figure 8](image_url)

Figure 8. SEM-EDX analysis of biochar samples. (a) Pyrolysis biochar, (b) H₂O activated biochar, and (c) CO₂ activated biochar.
Mechanism of In-Situ Catalytic Cracking of Biomass Tar over Biochar with Multiple Active Sites

DOI: http://dx.doi.org/10.5772/intechopen.91380

promote volatile hydrocarbon condensation reactions to form coke [25], which were caused by hydrodeasphalting (HDA) reactions. Thus, the H₂O/CO₂ activation of biochar impacted the biochar surface's morphology and metal content, both of which influenced the reforming of biomass tar over biochar.

3.2.4 BET analysis of biochar during H₂O/CO₂ activation and tar reforming

The biochar samples’ N₂-absorption/desorption isotherms at 77 K during H₂O/CO₂ activation and biomass tar reforming are shown in Figure 9. Compared with those of the original pyrolysis biochar, the pore systems of H₂O/CO₂-activated biochar samples and that from tar H₂O reforming over H₂O-activated biochar were better developed. Conversely, the other conditions exhibited pore structures that were somewhat blocked, especially for reforming over pyrolysis biochar in Ar and for the CO₂-activated biochar.

To further investigate the microphysical structures of the biochar samples, their BET surface properties were evaluated and are presented in Table 4. The unactivated pyrolysis biochar presented a BET surface area of 195.35 m²/g and a pore volume of 0.0999 cm³/g. Activation by H₂O and CO₂ increased the BET surface area to 307.45 and 237.71 m²/g, respectively. The biochar's porous structure enabled efficient tar adsorption, resulting in a good residence time of the tar reacting with the catalyst [46]. Table 5 also shows that the concentration of micropores (<2 nm), mesopores (2–50 nm), and macropores (>50 nm) varied between the samples. Thus, the ratio of micropores (<2 nm) to mesopores and macropores (>2 nm) (S_Mic./S_Ext.) was employed. The H₂O-activated biochar showed a lower value of this ratio (2.28) than that of the CO₂-activated biochar (4.57) indicating that activation/gasification under a CO₂ atmosphere produced a higher relative micropore content, whereas under an H₂O atmosphere mesopores were favored. This may be explained by considering that H₂O removes carbon atoms from the particle's interior, enlarging open micropores and opening closed micropores, promoting the formation of mesopores. Meanwhile, CO₂ causes changes in the biochar surface that create more micropores. According to Klinghoffer et al. [63], the higher biochar surface area was the main reason for better catalyst performance, but pore size distribution also affected its activity, and evidence of diffusion limitations in microporous biochar was observed. Elsewhere, it has been confirmed that mesopores significantly

Figure 9. N₂-absorption/desorption isotherms at 77 K for biochar obtained from different conditions: (A) in Ar over pyrolysis biochar; (B) in Ar over H₂O-activated biochar; (C) in Ar over CO₂-activated biochar; (D) in 15 vol.% H₂O over H₂O-activated biochar; and (E) in 29 vol.% CO₂ over CO₂-activated biochar.
enhance catalytic activity by allowing the penetration of macromolecules, facilitating their adsorption on the catalyst surface [64–66]. Thus, biochar used for catalytic tar reforming should ideally possess a high surface area and high mesoporosity (i.e., a small $S_{\text{Mic.}}/S_{\text{Ext.}}$ value).

After tar reforming in the Ar atmosphere, the biochar samples' BET surface area and pore volume markedly decreased. This was especially the case for tar reforming in Ar over the pyrolysis biochar and the CO$_2$-activated biochar where the BET surface areas fell to 3.78 and 4.07 m$^2$/g, respectively. According to the findings of Hosokai et al. [19], the decrease in surface area was attributed to tar forming coke deposits on the biochar’s surface. In the Ar atmosphere, tar was mainly decomposed via coking $[C_mH_n$ (aromatic compounds) $= C_mH_x$(coke) + $(n - x)/2 H_2]$. Thus, the biochar's activity could have fallen with a decrease in the biochar’s surface area and/or pore volume caused by coke deposition. This implies that when some tar molecules reacted with the biochar they were absorbed in a way that yielded a condensed-phase product (coke) that remained on the biochar surface.

However, with the gasification agents, especially H$_2$O, the relatively high BET surface area and pore volume of biochar were maintained following the tar-reforming reactions (see Table 5). This indicated that the tar was not reformed directly to give gaseous products but instead involved the intermediate formation of coke, which was subsequently gasified by H$_2$O/CO$_2$. El-Rub and Kamel [59] suggested that tars can be adsorbed onto the active sites of biochar particles. Adsorbed tar and coke molecules can be catalytically reformed to give CO and H$_2$ by steam and dry CO$_2$ thermochemical reactions, regenerating the pore structure. Meanwhile, free radicals that enter polymerization reactions and coke on biochar surfaces can be formed from tar decomposition. For the H$_2$O-activated biochar, the BET surface area only decreased a little following reforming (to 268.52 m$^2$/g) and the $S_{\text{Mic.}}/S_{\text{Ext.}}$ value remained 2.30. Given that these conditions also reformed the greatest portion of the tar, the existence of gasification agents, especially H$_2$O, appeared to stimulate and maintain catalytic activity by continually creating and regenerating pore structures in the biochar.

<table>
<thead>
<tr>
<th>Conditions</th>
<th>BET surface area (m$^2$/g)</th>
<th>Pore volume (cm$^3$/g)</th>
<th>Micro pore &lt; 2 nm (m$^2$/g)</th>
<th>External pore &gt; 2 nm (m$^2$/g)</th>
<th>$S_{\text{Mic.}}/S_{\text{Ext.}}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pyrolysis biochar</td>
<td>195.35</td>
<td>0.0999</td>
<td>170.46</td>
<td>24.89</td>
<td>6.85</td>
</tr>
<tr>
<td>H$_2$O-activated biochar</td>
<td>307.45</td>
<td>0.1745</td>
<td>213.83</td>
<td>93.62</td>
<td>2.28</td>
</tr>
<tr>
<td>CO$_2$-activated biochar</td>
<td>237.71</td>
<td>0.1330</td>
<td>195.02</td>
<td>42.69</td>
<td>4.57</td>
</tr>
<tr>
<td>(A) Pyrolysis and Ar reforming</td>
<td>3.78</td>
<td>0.0070</td>
<td>3.14</td>
<td>0.65</td>
<td>4.86</td>
</tr>
<tr>
<td>(B) H$_2$O activation and Ar reforming</td>
<td>117.53</td>
<td>0.0693</td>
<td>78.28</td>
<td>39.25</td>
<td>1.99</td>
</tr>
<tr>
<td>(C) CO$_2$ activation and Ar reforming</td>
<td>4.07</td>
<td>0.0074</td>
<td>2.12</td>
<td>1.95</td>
<td>1.09</td>
</tr>
<tr>
<td>(D) H$_2$O activation and H$_2$O reforming</td>
<td>268.52</td>
<td>0.1512</td>
<td>187.08</td>
<td>81.44</td>
<td>2.30</td>
</tr>
<tr>
<td>(E) CO$_2$ activation and CO$_2$ reforming</td>
<td>54.31</td>
<td>0.0346</td>
<td>42.98</td>
<td>11.33</td>
<td>3.79</td>
</tr>
</tbody>
</table>

Table 5. BET properties of biochar samples during activation and tar reforming.
3.2.5 XPS analysis of biochar during biomass tar reforming

The elemental contents (C, O, K, and Ca) at the surface of the biochar samples are shown in Figure 10. Samples taken the following tar reforming in (A) Ar over pyrolysis biochar, (B) Ar over H₂O-activated biochar, and (C) Ar over CO₂-activated biochar showed that the H₂O/CO₂ activation of biochar played an important role in maintaining the biochar's active sites, such as surface O-containing functional groups and AAEM species (especially K and Ca) and improved its tar-reforming performance. According to Du et al. [67], XPS revealed the evolution of AAEM species and char structures, and concentrations of AAEM species agreed well with surface atomic O concentrations. Similar results were obtained in Figure 10, where Ar reforming over H₂O-activated biochar yielded a biochar with a higher surface content of O (16.25 atomic%), K (0.80 atomic%), and Ca (0.45 atomic%) than the samples from Ar reforming with the CO₂-activated biochar and the pyrolysis biochar. Abundant O-containing groups on the biochar surface can form acidic centers that can combine with biomass tar precursors, which have negatively charged π-electron systems and activate thermal cracking reactions [61]. For tar reforming in Ar, more carboxylic (O=C=O)/carbonyl (C=O) groups and fewer aromatic (C=C/C=O) groups were formed on the H₂O/CO₂-activated biochar surface. Franz et al. [68] investigated the effects of O-containing groups, particularly carboxylic and carbonyl groups, on the adsorption of dissolved aromatics on ash-free activated carbon. They found that the adsorption mechanism was influenced by the surface functional group's properties, especially its ability to hydrogen-bond, and through its activating/deactivating influence on the tar's aromatic ring. As shown in Figure 5 for conditions (D) and (E), the existence of the gasification agents during tar reforming over H₂O/CO₂-activated biochar helped to limit coke formation on the biochar surface, likely by continually creating and regenerating surface active sites. This finding was consistent with that of a previous investigation [24].

The surface AAEM content remained high. For example, 2.12 atomic% K in H₂O and 1.83 atomic% in CO₂. This was similar for the surface O content (34.01 atomic% in H₂O and 32.07 atomic% in CO₂). A biochar with a higher O content appeared to favor the retention of AAEM species, with O likely serving as a link between the AAEM species and the char matrix [69]. In addition, the results of Wu et al. [70] suggest that adding H₂O was likely to have eliminated more tar, while the presence of CO₂ induced the formation of OH, H, and O radicals, which increase hydrocarbon conversion.

Figure 10.
XPS analysis of biochar samples from different conditions: (A) in Ar over pyrolysis biochar; (B) in Ar over H₂O-activated biochar; (C) in Ar over CO₂-activated biochar; (D) in 15 vol.% H₂O over H₂O-activated biochar; and (E) in 29 vol.% CO₂ over CO₂-activated biochar.
3.2.6 GC-MS analysis of biomass tar reforming over biochar

The biomass tar reforming in the gas phase (without involving biochar) in Ar at 800°C was mainly composed of aromatic tar compounds, owing to the secondary thermal cracking of in-situ biomass tar [52], compared with tar from H-form rice husk pyrolysis in fluidized bed at 500°C. Thermal cracking cannot completely convert tars [71]. Regarding the biochar catalyzed reactions, defined as the net tar loss owing to exposure to the biochar (i.e., the amount remaining after subtracting the amount of tar destroyed by vapor-phase cracking upstream and downstream of the biochar bed from the total change in tar amount during thermal treatment) [16], no new tar compounds were observed. According to Yao et al. [72], absent biochar, the gasification agent has a larger effect on the reforming of large aromatic ring systems (e.g., ≥2 fused benzene rings) than on smaller and isolated aromatics. However, here, biomass tars with a single aromatic ring or more than one ring structure were catalytically reformed over the various biochars. The conversion rates of tar compounds for the tar reformed without biochar in Ar at 800°C can be seen in Figure 11(a) and (b). These figures also show that H$_2$O/CO$_2$ notably enhanced in-situ reforming of both large and small aromatic ring systems in biomass tar. For the experiments in Ar (conditions (A), (B), and (C)), individual tar conversion rates were improved by activation by H$_2$O/CO$_2$. For example, phenylethyne conversion increased from 42.27% over pyrolysis biochar to 77.43% over H$_2$O-activated biochar and to 49.93% over CO$_2$-activated biochar. However, the magnitude of the improvement was limited because of coke formation on the active sites, which deactivated the biochar. Thus, continuously supplying gasification agents (H$_2$O and CO$_2$ in conditions D and E, respectively) made more complete biomass tar conversion possible. For example, reforming in 15 vol.% H$_2$O over H$_2$O-activated biochar saw conversion rates of tars with both single aromatic ring structures (e.g., phenylethyne and benzofuran) and multiring structures (e.g., 1-methy-naphthalene, 2-methy-naphthalene, and phenanthrene) reach almost 100%. Although tar conversion was not completed in the 29 vol.% CO$_2$ atmosphere, it was also notably higher than for reforming in Ar over CO$_2$-activated biochar. As mentioned above, the gasification agent directly affected gas-phase tar reforming reactions [72], and it is likely that H$_2$O/CO$_2$ indirectly affected tar destruction by influencing the biochar structure and distribution of AAEM catalysts during the reaction by helping to ensure enough active sites on the biochar surface to maintain its catalytic activity.

Figure 11. Biomass tar conversion rates (based on tar observed following the treatment without biochar in Ar at 800°C) for different reforming conditions: (A) in Ar over pyrolysis biochar; (B) in Ar over H$_2$O-activated biochar; (C) in Ar over CO$_2$-activated biochar; (D) in 15 vol.% H$_2$O over H$_2$O-activated biochar; and (E) in 29 vol.% CO$_2$ over CO$_2$-activated biochar.
The heterogeneous reforming mechanism of the biomass tar over biochar and in the presence of the H$_2$O and CO$_2$ reforming agents at 800°C is shown in Figure 12. H$_2$O and CO$_2$ dissociate in space to form a large number of H/O/OH radicals, which play an important role in the tar-biochar reforming reaction. Biomass tar, through the biochar layer, is adsorbed onto the acid-base active sites (oxygen-containing functional groups and AAEM catalysts). The attraction effect of the carbon-rich biochar matrix invokes an electron pair shift in the tar molecules (relatively small mass), which promotes the tar molecules to break at high temperatures. According to the free radical theory [73], the tar adsorbed on the catalyst surface will catalytically crack to form the corresponding free radicals. The chemical reaction between these free radicals may permit new products. H$_2$O and CO$_2$ act as the reforming agents in the biochar carbon matrix, resulting in the fragmentation of the smaller aromatic rings. The empty active sites, formed by bond cleavage, were gradually occupied by H/O/OH radicals, forming active groups such as O-containing functional groups. In the presence of H$_2$O and CO$_2$, a significant amount of H/O/OH radicals in the vicinity ingress into the biochar carbon structure. The catalytic elements, such as AAEM species migrate at different rates and transformation from the carbon matrix onto the gas-solid interface or the gas phase undergoes, as shown in Figure 12. As the AAEM species are bonded with the C element on the biochar surface by the O element [21], H$_2$O and CO$_2$ react with these C elements on the biochar surface resulting in AAEM-O bond cleavage followed by precipitation. The valence state of Ca results in a stronger bonding interaction with the biochar when compared with K. Additionally, Ca migration and precipitation are more difficult than K. When tar adsorbs then cleaves the AAEM-O bond and functional group bond on the biochar surface, an aromatic fragmented radical is formed when other free radicals are encountered. At the same time, active AAEM species in the vicinity will continue to occupy active sites on the tar fragment groups, thereby inhibiting their secondary polymerization. At the same time, the H/O/OH radicals are exchanged to the AAEM species, which increases the possibility for the reforming of tar macromolecules. After the reaction, gas and light tars (C$_n$H$_y$/CO/H$_2$) were formed, thus realizing the H$_2$O or CO$_2$ heterogeneous reforming of biomass tar over the biochar catalyst.

Figure 12.
Heterogeneous reforming mechanism of biomass tar over biochar in the presence of H$_2$O and CO$_2$ at 800°C [52].
4. Conclusions

The tar yield decreases as a function of increasing temperature from 26.18% at 500°C to 6.38% at 900°C. H$_2$O and CO$_2$ influence significantly the tar homogeneous transformations at 700–900°C, while the tar reforming effect of 15 vol.% H$_2$O is significantly higher than that of 29 vol.% CO$_2$. H$_2$O and CO$_2$ have obvious effects on the transformation of PAHs. H$_2$O and CO$_2$ not only directly affect the tar transformation on biochar but also indirectly influence the reforming of tar through changing the structure of biochar catalyst. The formation of additional oxygen-containing functional groups is strengthened with the concentration of H$_2$O and CO$_2$ increasing. During tar heterogeneous reforming over biochar, the transformation of small aromatic ring systems (3–5 fused rings) to larger aromatic ring systems (≥6 fused rings) in the biochar structure is promoted by the increasing concentration of H$_2$O and CO$_2$. The activation by H$_2$O/CO$_2$ of biochar impacted the biochar surface's morphology and distribution of metal species. Activation/gasification under a CO$_2$ in an Ar atmosphere produced more micropores, while adoption under a H$_2$O in an Ar atmosphere favored the formation of mesopores. With the existence of gasification agents, especially for H$_2$O, the simultaneous creation of pore structures is necessary to maintain biochar's catalytic activity during tar reforming. H$_2$O/CO$_2$ also indirectly affects tar destruction by influencing the biochar structure and distribution of AAEM catalysts, while the reaction is occurring to ensure enough active sites on the biochar surface to maintain its catalytic activity. The activation and/or activity-maintaining effects of H$_2$O/CO$_2$ can notably enhance the in-situ reforming of both large and small aromatic ring systems present in biomass tar.

Acknowledgements

This work is supported by the National Natural Science Foundation of China (51906052), the National Postdoctoral Program for Innovative Talents of China (BX20180086), China Postdoctoral Science Foundation Funded Project (2018M641826), Heilongjiang Provincial Postdoctoral Science Foundation, Foundation of State Key Laboratory of High-efficiency Utilization of Coal and Green Chemical Engineering (2019-KF-14), and Fundamental Research Funds for the Central Universities (Grant no. HIT. NSRIF. 2020052).

Conflict of interest

The authors declare no conflict of interest.
References


[17] Wang D, Yuan W, Ji W. Char and char-supported nickel catalysts for secondary syngas cleanup and
Mechanism of In-Situ Catalytic Cracking of Biomass Tar over Biochar with Multiple Active Sites
DOI: http://dx.doi.org/10.5772/intechopen.91380


[34] Takahashi H, Iwatsuki M, Essaki K, Tsutsumi A, Chiba T. Rapid conversion...


[50] Dong G, Hüttenger K. Consideration of reaction mechanisms leading to
pyrolytic carbon of different textures. 
Carbon. 2002;40:2515-2528


[56] Poutsma ML. A Review of Thermolysis Studies of Model Compounds Relevant to Processing of Coal. TN, USA: Oak Ridge National Lab; 1987

[57] Futamura S, Annadurai G. Plasma reforming of aliphatic hydrocarbons with CO2. ITIA. 2005;41:1515-1521


