We are IntechOpen, the world’s leading publisher of Open Access books
Built by scientists, for scientists

4,100
Open access books available

116,000
International authors and editors

125M
Downloads

154
Countries delivered to

TOP 1%
Our authors are among the most cited scientists

12.2%
Contributors from top 500 universities

WEB OF SCIENCE™
Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com
Chapter

Photodynamic Inactivation of Escherichia coli with Cationic Porphyrin Sensitizers

Jin Matsumoto, Tomoko Matsumoto, Kazuya Yasuda and Masahide Yasuda

Abstract

The activity of singlet-oxygen sensitizers for photodynamic inactivation (PDI) of microorganisms and photodynamic therapy of tumor cells has been evaluated using Escherichia coli, Saccharomyces cerevisiae, and human cancer cell lines. In this chapter, drug resistance of E. coli was examined based on the PDI activity of a variety of RPy-P-porphyrin sensitizers with different number of ionic valence and different hydrophobic characters. The PDI activities toward E. coli were evaluated using the minimum effective concentrations ([P]) of the porphyrin sensitizers. It was found that the [P] value for E. coli was larger than that for S. cerevisiae. E. coli has drug-resistance toward hydrophobic and mono-cationic porphyrins. However, E. coli has weak drug-resistance toward the porphyrins with both polycationic character and hydrophobicity. Since the outer membrane mainly consists of lipopolysaccharides and phospholipids that are negatively charged, cationic porphyrins are able to adsorb to the outer leaflet. Then the cationic porphyrins with hydrophobic character can interact with not only the outer leaflet but also inner leaflet of the outer membrane and the plasma membrane. Thus, porphyrins may be incorporated inside E. coli cells via the self-promoted uptake pathway. Moreover, polycationic porphyrins can interact with DNA and proteins by strong binding affinities.

Keywords: PDT sensitizer, singlet oxygen, porphyrins, PDI activity, Escherichia coli, Saccharomyces cerevisiae

1. Introduction

Singlet-oxygen (¹O₂) sensitizers for photodynamic inactivation (PDI) of microorganisms and photodynamic therapy of tumor cells have been developed using Escherichia coli, Saccharomyces cerevisiae, and human cancer cell lines (e.g., HeLa cell) as model cells [1–4]. As E. coli is a Gram-negative bacterium, the cell wall consists of an inner membrane, cytoplasmic membrane, a periplasmic space with a peptidoglycan layer, and an outer membrane [5]. Since the E. coli cell wall has a low permeability, there are only a few ¹O₂-sensitizers that can permeate the cell wall and inactivate E. coli efficiently at low concentrations.

PDI refers to the use of a visible-light source, oxidizing agents (e.g., O₂), and photosensitizers. Photosensitizers absorb light energy that causes an energy transfer
to O₂, which leads to the formation of reactive oxygen such as O₂⁻, thereby inactivating cells and bacteria. Preliminary studies on the photodynamic action for biological systems started in the 1930s by PDI of phages using methylene blue [6, 7]. PDI of bacteria has received considerable attention as a methodology leading to the medical application of infection therapy beyond antimicrobial resistance. Among the large variety of photosensitizers developed for PDI over the last 60 years, porphyrins and metalloporphyrins became attractive sensitizers owing to their strong absorption band in the visible-light region [8–11].

In the case of porphyrin sensitizers, their solubilities in water are an important characteristic for handling them as aqueous solutions, since porphyrin derivatives, in general, are poorly soluble in water. The most popular method to improve the solubility in water is the introduction of ionic groups to the porphyrin ring. Especially, the introduction of an alkylpyridinium (RPy) group into porphyrins is a useful method to make porphyrins water-soluble [12, 13]. A typical RPy-bonded porphyrin is represented by meso-tetra[4-(1-methyl-pyridinium)] porphyrin (TMP). The first application of TMP to PDI was reported by Ben Amor et al. in 1998 [14]. For the last two decades, a variety of RPy-bonded porphyrins have been prepared and studied for PDI [15–21].

We have interested in axially RPy-bonded tricationic P-porphyrins and their PDI activity [22–26]. It is advantageous that the water solubilization is easily achieved through the modification of the axial ligands of P-porphyrins. It is expected that polycationic porphyrins have strong binding affinities to DNA [27–32]. In this chapter, drug resistance of E. coli was discussed based on PDI activity of a variety of P- and Sb-porphyrin sensitizers with different number of ionic valence and different hydrophobic character. The typical structure of the porphyrin sensitizer is shown in Figure 1, and they are named P-type porphyrin.

2. Materials and methods

2.1 Axially RPy-bonded tricationic P-porphyrins: (RPy3)₂P(Tpp)³⁺

The preparation of tricationic bis[3-(1-alkyl-4-pyridinium)propoxo]tetraphenylporphyrinatophosphorus(V) complex, (RPy3)₂P(Tpp)³⁺ (Tpp = tetr phenylporphyrinato group), was performed as follows [22]. Dichloro(tetraphenylporphyrinato)phosphorus chloride ([Cl₂P(Tpp)]Cl [33], 300 mg) was reacted with 3-(4-pyridyl)-1-propanol (5.0 mL) in MeCN (30 mL) at reflux temperature.
Photodynamic Inactivation of Escherichia coli with Cationic Porphyrin Sensitizers

DOI: http://dx.doi.org/10.5772/intechopen.82645

for about 24 h until the Soret band shifted from 435 to 428 nm. Bis[3-(4-pyridyl) propoxo]tetraphenylporphyrinatoantimony(V) chloride, (Py3)2Sb(Tpp)1+, was produced in 47% yield. The (Py3)2P(Tpp)1+ (50 mg) was reacted with alkyl halides (1.0 mL) in MeCN (25 mL) at reflux temperature for about 24 h to give (RPy3)2P(Tpp)3+, the yields of (RPy3)2P(Tpp)3+ are listed in Table 1.

2.2 Axially RPy-bonded polycationic Sb-porphyrins

Axially RPy-bonded polycationic Sb-porphyrins were prepared using dibromo(tetraphenylporphyrinato)antimony bromide ([Br2Sb(Tpp)]Br) as the starting material [34]. The partial methanolysis of [Br2Sb(Tpp)]Br (1.077 g) was performed in MeOH-MeCN (1:1, 160 mL) in the presence of pyridine (0.75 mL) at 80°C until the Soret band shifted from 427 to 423 nm. Bromo(methoxo)-(tetraphenylporphyrinato)antimony bromide ([MeO(Br)Sb(Tpp)]Br, 520 mg) was formed in 61% yield [35]. An MeCN (20 mL) solution of [Br2Sb(Tpp)]Br (150 mg) and [MeO(Br)Sb(Tpp)]Br (180 mg) was heated with 3-(4-pyridyl)-1-propanol (3.7 mL) at refluxing temperature for about 24 h until the Soret band

<table>
<thead>
<tr>
<th>Sensitizers</th>
<th>n^b</th>
<th>Z^a</th>
<th>Metal</th>
<th>Yield/%</th>
<th>ε10^4 M−1 cm−1^c</th>
<th>CW/mM^d</th>
</tr>
</thead>
<tbody>
<tr>
<td>(MePy3)2P(Tpp)</td>
<td>1</td>
<td>+3</td>
<td>P</td>
<td>95</td>
<td>26.9</td>
<td>1.38</td>
</tr>
<tr>
<td>(BuPy3)2P(Tpp)</td>
<td>4</td>
<td>+3</td>
<td>P</td>
<td>93</td>
<td>23.1</td>
<td>1.18</td>
</tr>
<tr>
<td>(PentPy3)2P(Tpp)</td>
<td>5</td>
<td>+3</td>
<td>P</td>
<td>32</td>
<td>27.2</td>
<td>1.32</td>
</tr>
<tr>
<td>(HexPy3)2P(Tpp)</td>
<td>6</td>
<td>+3</td>
<td>P</td>
<td>47</td>
<td>31.3</td>
<td>1.45</td>
</tr>
<tr>
<td>(HeptPy3)2P(Tpp)</td>
<td>7</td>
<td>+3</td>
<td>P</td>
<td>32</td>
<td>26.7</td>
<td>1.26</td>
</tr>
<tr>
<td>(OctPy3)2P(Tpp)</td>
<td>8</td>
<td>+3</td>
<td>P</td>
<td>48</td>
<td>18.7</td>
<td>0.97</td>
</tr>
<tr>
<td>(HexPy3)Sb(Tpp)</td>
<td>6</td>
<td>+3</td>
<td>Sb</td>
<td>35</td>
<td>16.3</td>
<td>4.18</td>
</tr>
<tr>
<td>(MePy3)Sb(Tpp)</td>
<td>1</td>
<td>+2</td>
<td>Sb</td>
<td>42</td>
<td>12.7</td>
<td>4.45</td>
</tr>
<tr>
<td>(HexPy3)Sb(Tpp)</td>
<td>6</td>
<td>+2</td>
<td>Sb</td>
<td>25</td>
<td>15.1</td>
<td>4.48</td>
</tr>
<tr>
<td>(MePy5)2P(Tpp)</td>
<td>1</td>
<td>+3</td>
<td>P</td>
<td>73</td>
<td>28.2</td>
<td>1.36</td>
</tr>
<tr>
<td>(EtPy5)2P(Tpp)</td>
<td>2</td>
<td>+3</td>
<td>P</td>
<td>58</td>
<td>29.6</td>
<td>1.40</td>
</tr>
<tr>
<td>(BuPy5)2P(Tpp)</td>
<td>4</td>
<td>+3</td>
<td>P</td>
<td>44</td>
<td>25.3</td>
<td>1.29</td>
</tr>
<tr>
<td>(HexPy5)2P(Tpp)</td>
<td>6</td>
<td>+3</td>
<td>P</td>
<td>44</td>
<td>24.7</td>
<td>1.22</td>
</tr>
<tr>
<td>(4EtPy5)2P(Tpp)</td>
<td>2</td>
<td>+3</td>
<td>P</td>
<td>72</td>
<td>12.7</td>
<td>0.57</td>
</tr>
<tr>
<td>(Me)2P(PyHex)</td>
<td>6</td>
<td>+2</td>
<td>P</td>
<td>57</td>
<td>22.6</td>
<td>1.31</td>
</tr>
<tr>
<td>(Me2)P(PyHex)</td>
<td>6</td>
<td>+2</td>
<td>P</td>
<td>78</td>
<td>14.1</td>
<td>0.89</td>
</tr>
<tr>
<td>(Bu2)P(PyMe)</td>
<td>1</td>
<td>+2</td>
<td>P</td>
<td>94</td>
<td>18.1</td>
<td>1.01</td>
</tr>
<tr>
<td>(Bu2)P(PyMe)</td>
<td>1</td>
<td>+2</td>
<td>P</td>
<td>32</td>
<td>21.7</td>
<td>1.21</td>
</tr>
<tr>
<td>(Hex2)P(PyMe)</td>
<td>1</td>
<td>+2</td>
<td>P</td>
<td>45</td>
<td>28.6</td>
<td>1.63</td>
</tr>
</tbody>
</table>

^aZ = charge of the complex.

^b n = carbon number of the alkyl chain on the Ap.

^c Molar absorption coefficient for the Soret and the Q bands in MeOH solution.

^d CW = water solubility in mM.

^e Broadening of UV spectra occurred.

Table 1.

PDI of E. coli with cationic porphyrins.
shifted to 418 nm, respectively. Thus, bis[3-(4-pyridyl)propoxo]tetraphenylporphyrinatoantimony (V) bromide ((Py3)_2Sb(Tpp)^+, 83 mg) and 3-(4-pyridyl)propoxo(methoxo)tetraphenylporphyrinatoantimony (V) bromide (Py3Sb(Tpp)^+, 90 mg) were obtained in 50% and 43% yields, respectively. (Py3)_2Sb(Tpp)^+ (50 mg) was reacted with 1-bromohexane (0.5 mL) in MeCN (13 mL) at reflux temperature for about 24 h to give bis[3-(1-hexyl-4-pyridinio)-1-propoxo]-5,10,15,20-tetraphenylporphyrinatoantimony (V) tribromide ((HexPy3)_2Sb(Tpp)^3+, 20 mg, 35%). The reaction of (Py3Sb(Tpp)^+, 50 mg) with Mel and 1-bromohexane (0.5 mL in MeCN (13 mL) at reflux temperature for about 24 h gave α-(methoxo)-β-[3(1-methyl-4-pyridinio)-1-propoxo]-5,10,15,20-tetraphenylporphyrinatoantimony (V) dibromide (MePy3Sb(Tpp)^2+, 25 mg, 42%) and α-(methoxo)-β-[3(1-hexyl-4-pyridinio)-1-propoxo]-5,10,15,20-tetraphenylporphyrinatoantimony (V) dibromide (HexPy3Sb(Tpp)^2+, 20 mg, 25%), respectively [24].

2.3 Axially RPy-bonded tricationic P-porphyrins: (RPy5)_2P(Tpp)^3+

Bis[5-(3-alkyl-1-pyridinio)-3-oxapentyloxo]tetraphenylporphyrinato-phosphorus(V) dibromide, chloride ((RPy5)_2P(Tpp)^3+) was prepared from dihydroxo(tetraphenylporphyrinato)phosphorus chloride ([HO2P(Tpp)]Cl), which was prepared by hydrolysis of [Cl2P(Tpp)]Cl (300 mg) by refluxing in a mixed solvent of MeCN (160 mL) with pyridine (60 mL) and H2O (60 mL) [22]. Alkylation of [HO2P(Tpp)]Cl (80 mg) with di(2-bromoethyl) ether (1 mL) was performed in the presence of K2CO3 (19 mg) and 18-crown-6 ether (4.2 mg) in MeCN (5 mL) at 50°C to give bis(5-bromo-3-oxa-pentyloxo)tetraphenylporphyrinatophosphorus(V) chloride ((Br5)_2P(Tpp)^+). The (Br5)_2P(Tpp)^+ (50 mg) was reacted with 3-alkylpyridine (1.0 mL) in MeCN (10 mL) under heating at 100°C for 20–68 h for the preparations of (RPy5)_2P(Tpp)^3+ [22]. Similarly, bis[5-(4-ethyl-1-pyridinio)-3-oxapentyloxo]tetraphenylporphyrinatophosphorus(V) dibromide, chloride, (4EtPy5)_2P(Tpp)^3+ was prepared via the reaction of (Br5)_2P(Tpp)^+ (63 mg) with 4-ethylpyridine (1.0 mL) in dry MeCN (10 mL) at 100°C for 20 h.

2.4 RPy-bonded dicationic P-porphyrins at meso position: (R’m)_2P(RPy-Tpp)^2+

At first, 5,10,15-triphenyl-20-(4-pyridinyl)porphyrin (PyTpp) was prepared by reaction of pyrrole (1.55 mL), benzaldehyde (1.83 mL), and 4-formylpyridine (0.56 mL) in propanoic acid (100 mL) in an oil bath heated at 140°C for 1 h to give PyTpp (533 mg, 14%) [24]. PyTpp (101 mg) was reacted with phosphoryl chloride (POCl3, 2.0 mL) in pyridine (10 mL) in a pressure bottle heated at 180°C for 1 day to give dichloro[triphenyl(4-pyridinyl)porphyrinato]phosphorus chloride ([Cl2P(PyTpp)]Cl, 99.0 mg) in 81% yield. Substitution of the axial chloro ligand with a methoxo group was performed by refluxing [Cl2P(PyTpp)]Cl (82.7 mg) in MeOH (20 mL) at 100°C for 20 h. Dimethoxo[5-(1-hexyl-4-pyridinio)-10,15,20-triphenyl-porphyrinato]phosphorus (V) dichloride (Me2P(HexPyTpp)^2+) was prepared by reaction of (Me2P(PyTpp)]Cl (62.0 mg) with 1-iodohexane (2 mL) in DMF (5 mL) in the presence of K2CO3 (19 mg) at 100°C for 2 h. (Me2P(HexPyTpp)^2+ was purified through anion exchange with chloride ions, as follows. An aqueous solution (10 mL) of AgBF4 (115 mg) was added to a MeCN-MeOH (1:1 v/v, 20 mL) solution of the porphyrins. After stirring for 24 h at room temperature, the solution was washed with water (100 mL) and an aqueous NaCl solution (100 mL) three times and subjected to precipitation with hexane (200 mL) [24].
[Cl\(2P(PyTpp)\)]Cl (78–100 mg) was reacted with ethylene glycol derivatives (H(OCH\(2\)CH\(2\))\(m\)OR', R' = Me, n-Bu, n-Hex, 5–7 mL) in MeCN (10 mL) in the presence of pyridine (0.75 mL) for 24 h to give bis(2-alkyloxyethoxo)-5-(4-pyridinyl)-10,15,20-triphenylporphyrinatophosphorus (V) chloride (\([(R')\(m\)2P(PyTpp)]Cl\) in 66–88%. Bis(2-methoxyethoxo)-5-(1-hexyl-4-pyridinyl)-10,15,20-triphenylporphyrinatophosphorus (V) bromide, chloride ((Me\(1\)2P(HexPyTpp))\(2+\) was prepared by reaction of 
\([(Me\(1\)2P(PyTpp)]Cl (51 mg) with 1-iodohexane (2 mL) in DMF (5 mL) in the presence of K\(2\)CO\(3\) (19 mg) in an oil bath heated at 100°C for 2 h. After anion-exchange, dichloride salt of 
(Me\(1\)2P(HexPyTpp))\(2+\) (27 mg, 78%) was obtained. Also, other meso-RPy-bonded
dicationic P-porphyrins (61–90 mg) were reacted with MeI (1.2 mL) in DMF (7.5 mL) in the presence of K\(2\)CO\(3\) (43 mg) by heating at 100°C for 24 h to give an 
N-methyl-substituted complex. After anion exchange, (Me\(1\)2P(HexPyTpp))\(2+\) (35 mg, 94%), (Bu\(2\)2P(MePyTpp))\(2+\) (13.7 mg, 32%), and (Hex\(2\)2P(MePyTpp))\(2+\) (28.0 mg, 45%) were formed [24].

2.5 Preparation of E. coli suspension

E. coli K-12 (IFO 3301) was cultured aerobically at 30°C for 8 h in a LB medium (pH 6.5) consisting of bactotryptone (10 g L\(^{-1}\)), yeast extract (5 g L\(^{-1}\)), and NaCl (10 g L\(^{-1}\)). After centrifugation of the cultured broth at 12,000 rpm for 10 min, the harvested cells were washed with physiological saline (NaCl, 7 g L\(^{-1}\)) and then 
suspended in physiological saline, resulting in a cell suspension of E. coli. The cell 
concentrations were determined using a calibration curve and turbidity quantified 
by the absorbance measured at 600 nm on an UV–Vis spectrometer [24].

2.6 PDI of E. coli

PDI of E. coli was performed as follows. A phosphate buffer (0.1 M, pH 7.6) was 
prepared by dissolving Na\(_2\)HPO\(_4\) (2.469 g) and NaH\(_2\)PO\(_4\) (0.312 g) in 100 mL of water. The suspension of E. coli cells (1 \(\times\) 10\(^{8}\) cells mL\(^{-1}\), 1.0 mL), an aqueous solution of the studied sensitizers (25–100 \(\mu\)M, 0.1 mL), and the phosphate buffer (0.1 M, pH 7.6, 8.9 mL) were introduced into L-type glass tubes, resulting in a buffer solution (10 mL) containing E. coli (1 \(\times\) 10\(^{8}\) cells mL\(^{-1}\)) and the studied sensitizers (0.25–1.0 \(\mu\)M). Under dark conditions, the L-type glass tubes were set on a reciprocal shaker and shaken at 160 rpm at room temperature for 2 h [24]. And then the L-type glass tubes were 
irradiated using a fluorescent lamp (Panasonic FL-15ECW, Japan; wave length = 400– 
723 nm; the maximum intensity: 545 nm; 10.5 W cm\(^{-2}\)) on a reciprocal shaker at room 
temperature. A portion of the reaction mixture (0.1 mL) was taken up to 2 h at 20-min 
intervals and plated on LB plates. The LB plates were incubated for 30 h at 30°C.

The amount of the living cells (\(B\)) was defined as the average number of E. coli 
colonies that appeared after an incubation period of 30 h in three replicate plates. The \(B\) values for the PDI sensitizers were recorded at each irradiation time.

2.7 Fluorescence imaging

Incorporation of porphyrin sensitizers inside cells can be examined by fluores-
cence microscopy images of E. coli on a confocal laser scanning microscope (CLSM) under laser excitation at 543 nm. The aqueous solution containing the porphyrin 
sensitizers and E. coli was incubated for 3 h at 25°C. The concentrated solution was 
sandwiched between a cover slip and an agar pad on a bottom cover slip to maintain 
its position within the same focal plane [36].
3. Results

3.1 Properties of RPy-bonded P-porphyrins

Figure 2 shows the structures of the prepared porphyrins, which were water soluble due to cationic complexes. The water solubility ($C_{W}$) is listed in Table 1. In addition, Table 1 lists the absorption coefficient ($\epsilon$) of Soret band around 431 nm and Q-band at 562 nm in MeOH. These porphyrins could absorb strongly visible
light. Moreover, they could generate $^1$O$_2$ efficiently, since the quantum yields for the formation of $^1$O$_2$ were found to be 0.88 for (HexPy3)$_2$P(Tpp)$_{3+}$ and 0.87 for (Bu2)$_2$P(MePyTpp)$_{2+}$ [23].

3.2 Results of PDI of E. coli

Results of PDI of E. coli are summarized in Table 2. As seen from Table 2, Meso-RPy-substituted P-porphyrins ((R'm)$_2$P(RPyTpp)$_{2+}$) have cytotoxicity, since E. coli was inactivated under dark conditions.

Based on Table 2, the survival ratios were calculated as $100B/B_0$ where $B_0$ is the initial amount of bacteria. From the time-course plots of survival ratios ($100B/B_0$), the half-life ($T_{1/2}$ in min), i.e., the time required to reduce $B$ from $B_0$ to 0.5$B_0$, was measured. A typical example of time-course plots is the case of PDI of E. coli by (HexPy3)$_2$P(Tpp)$_{3+}$ as shown in Figure 3. In this case, the $T_{1/2}$ value of

<table>
<thead>
<tr>
<th>Sensitizers</th>
<th>[P]/μM</th>
<th>Amount of bacteria ([B])/CFU mL$^{-1}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>(MePy5)$_2$P(tpp)</td>
<td>2.0</td>
<td>512 ± 22, 450 ± 14, 383 ± 13, 344 ± 20</td>
</tr>
<tr>
<td>(BuPy5)$_2$P(tpp)</td>
<td>0.25</td>
<td>145 ± 11, 123 ± 76, 92 ± 7.5, 63 ± 4.6</td>
</tr>
<tr>
<td>(HexPy5)$_2$P(tpp)</td>
<td>0.25</td>
<td>213 ± 10, 213 ± 9.5, 176 ± 16, 166 ± 6.8</td>
</tr>
<tr>
<td>(EtPy5)$_2$P(tpp)</td>
<td>0.25</td>
<td>139 ± 14, 85 ± 13, 88 ± 16, 62 ± 6.0</td>
</tr>
<tr>
<td>(Me)$_2$P(PyHex)</td>
<td>0.5</td>
<td>89 ± 2.7, 57 ± 2.9, 42 ± 7.2, 43 ± 2.7</td>
</tr>
<tr>
<td>(Bu1)$_2$P(PyMe)</td>
<td>0.5</td>
<td>34 ± 5.0, 25 ± 3.5, 28 ± 6.1, 31 ± 3.5</td>
</tr>
<tr>
<td>(Bu2)$_2$P(PyMe)</td>
<td>2.0</td>
<td>126 ± 14, 56 ± 3.8, 21 ± 4.9, 8.7 ± 2.1</td>
</tr>
<tr>
<td>(Hex2)$_2$P(PyMe)</td>
<td>1.0</td>
<td>63 ± 5.9, 50 ± 7.5, 56 ± 2.1, 45 ± 8.1</td>
</tr>
</tbody>
</table>

*PDI of E. coli was performed in a phosphate buffer solution (10 mL, pH 7.6) containing E. coli (ca. 2 × 10$^8$ cell mL$^{-1}$) and porphyrin sensitizers under the irradiation of a fluorescent lamp. CFU = colony formation unit.

**[P] was adjusted to attain the value of $T_{1/2}$ between 20 and 120 min.

*Irradiation time (t) in min.

*Under dark conditions.

Table 2. 
PDI of E. coli with cationic porphyrins under visible light irradiation.
The Universe of Escherichia coli

3.3 PDI activity of the porphyrin sensitizers toward E. coli

As shown in Table 3, the $A_F$ values were dependent on the number of carbon atoms ($n$) in the alkyl group on the RPy group in (RPy3)$_2$M(Tpp)$_{3+}$ ($M = P, Sb$), RPy3Sb(Tpp)$_{2+}$, and (RPy5)$_2$P(Tpp)$_{3+}$. Figure 4A shows the dependence of the $A_F$ values on $n$ in the case of a series of (RPy3)$_2$M(Tpp)$_{3+}$ ($M = P, Sb$) and RPy3Sb(Tpp)$_{2+}$. The maximum value of $A_F$ appeared at $n = 7$ whose $[P]$ value was 0.40 $\mu $M. Moderately long alkyl chain made the sensitizer more active toward E. coli [24]. In the case of a series of (RPy5)$_2$P(Tpp)$_{3+}$ (Figure 4B), the maximum value of $A_F$ appeared at $n = 2$ whose $[P]$ value for E. coli was 0.25 $\mu $M [25]. Therefore, the $A_F$ and $[P]$ values of 3-ethyl analog were compared with those of 4-ethyl isomer. It was found that the $A_F$ value of 4-ethyl isomer was lower than that of 3-ethyl isomer. In the case of the 4-ethyl analog, broadening of Soret and Q bands occurred due to aggregation of porphyrin chromophores. It is suggested that aggregation caused to lower the $A_F$ value of 4-ethyl isomer (4EtPy5)$_2$P(Tpp)$_{3+}$).

Figure 5 shows the fluorescence images of E. coli in the presence of depicting the emission from (MePy3)$_2$P(Tpp)$_{3+}$ and (HexPy3)$_2$P(Tpp)$_{3+}$ inside E. coli. The images show that (HexPy3)$_2$P(Tpp)$_{3+}$ was accumulated inside E. coli, whereas (MePy3)$_2$P(Tpp)$_{3+}$ was not. (HexPy3)$_2$P(Tpp)$_{3+}$, which had a large affinity to E. coli, had the high PDI activity. The RPy group with a long alkyl chain made the sensitizer reactive toward E. coli.
3.4 Comparison of the PDI activity in E. coli with the PDI activity in Saccharomyces cerevisiae

For comparison of the PDI activity in E. coli and other microorganisms, PDI of S. cerevisiae was performed using (RPy3)2P(Tpp)3. It could photoinactivate S. cerevisiae in lower concentration compared with the case of E. coli [23]. For example, the [P] values of (MePy3)2P(Tpp)3 for S. cerevisiae were 0.05 μM, while that for E. coli was 2.0 μM. Moreover, PDI of S. cerevisiae was performed using other porphyrins (Type E, Figure 6), which were monocationic and highly hydrophobic. The PDI of S. cerevisiae occurred efficiently by Type E porphyrins [37]. The [P] values for the PDI of S. cerevisiae were optimized to be 0.005 μM. Thus, S. cerevisiae has low drug resistance for hydrophobic sensitzers rather than polycationic sensitizers, since the [P] value of tricationic porphyrins was larger than that of monocationic porphyrins (Type E). On the contrary, no PDI of E. coli by Type E porphyrins occurred at all. This result shows that a more positive character is required for an efficient PDI of E. coli.

<table>
<thead>
<tr>
<th>Sensitizer</th>
<th>Z</th>
<th>Metal</th>
<th>n</th>
<th>[P]/μM</th>
<th>T1/2/min</th>
<th>A5/μM−1 h−1</th>
</tr>
</thead>
<tbody>
<tr>
<td>(MePy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>1</td>
<td>2.0</td>
<td>66</td>
<td>0.5</td>
</tr>
<tr>
<td>(BuPy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>4</td>
<td>2.0</td>
<td>27</td>
<td>1.1</td>
</tr>
<tr>
<td>(PentPy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>5</td>
<td>0.5</td>
<td>29</td>
<td>4.1</td>
</tr>
<tr>
<td>(HexPy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>6</td>
<td>0.5</td>
<td>31</td>
<td>3.8</td>
</tr>
<tr>
<td>(HeptPy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>7</td>
<td>0.4</td>
<td>24</td>
<td>6.3</td>
</tr>
<tr>
<td>(OctPy3)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>8</td>
<td>0.5</td>
<td>63</td>
<td>1.9</td>
</tr>
<tr>
<td>(HexPy3)2Sb(Tpp)</td>
<td>+2</td>
<td>Sb</td>
<td>6</td>
<td>1.0</td>
<td>36</td>
<td>1.7</td>
</tr>
<tr>
<td>(MePy3)2Sb(Tpp)</td>
<td>+2</td>
<td>Sb</td>
<td>1</td>
<td>1.0</td>
<td>106</td>
<td>0.6</td>
</tr>
<tr>
<td>(HexPy3)2Sb(Tpp)</td>
<td>+2</td>
<td>Sb</td>
<td>6</td>
<td>1.0</td>
<td>68</td>
<td>0.9</td>
</tr>
<tr>
<td>(MePy5)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>1</td>
<td>1.0</td>
<td>40</td>
<td>1.5</td>
</tr>
<tr>
<td>(EtPy5)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>2</td>
<td>0.25</td>
<td>32</td>
<td>7.5</td>
</tr>
<tr>
<td>(ButPy5)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>4</td>
<td>0.25</td>
<td>53</td>
<td>4.5</td>
</tr>
<tr>
<td>(HexPy5)2P(Tpp)</td>
<td>+3</td>
<td>P</td>
<td>6</td>
<td>0.25</td>
<td>120</td>
<td>2.0</td>
</tr>
<tr>
<td>(4EtPy5)2P(Tpp)</td>
<td>+2</td>
<td>P</td>
<td>2</td>
<td>0.5</td>
<td>50</td>
<td>2.4</td>
</tr>
<tr>
<td>(Me2)2P(PyHex)</td>
<td>+2</td>
<td>P</td>
<td>6</td>
<td>2.0</td>
<td>45</td>
<td>0.7</td>
</tr>
<tr>
<td>(Me2)2P(PyHex)</td>
<td>+2</td>
<td>P</td>
<td>6</td>
<td>0.5</td>
<td>37</td>
<td>3.2</td>
</tr>
<tr>
<td>(Bu1)2P(PyMe)</td>
<td>+2</td>
<td>P</td>
<td>1</td>
<td>0.5</td>
<td>55</td>
<td>2.2</td>
</tr>
<tr>
<td>(Bu2)2P(PyMe)</td>
<td>+2</td>
<td>P</td>
<td>1</td>
<td>2.0</td>
<td>23</td>
<td>1.3</td>
</tr>
<tr>
<td>(Hex2)2P(PyMe)</td>
<td>+2</td>
<td>P</td>
<td>1</td>
<td>1.0</td>
<td>116</td>
<td>0.5</td>
</tr>
</tbody>
</table>

The PDI did not occur under dark conditions except for meso-RPy-substituted P-porphyrins, which were cytotoxic under dark conditions

Z = charge of the complex.

n = carbon number of the alkyl chain on the AP.

[P] = minimum concentrations of the porphyrins adjusted to attain the value of T1/2 between 20 and 120 min.

T1/2 = half-life in min.

AF = PDI activity in μM−1 h−1: AF = 60/(P × T1/2).

Table 3. The [P], T1/2, and AF values in the PDI of E. coli by cationic porphyrins.

3.4 Comparison of the PDI activity in E. coli with the PDI activity in Saccharomyces cerevisiae

For comparison of the PDI activity in E. coli and other microorganisms, PDI of S. cerevisiae was performed using (RPy3)2P(Tpp)3. It could photoinactivate S. cerevisiae in lower concentration compared with the case of E. coli [23]. For example, the [P] values of (MePy3)2P(Tpp)3 for S. cerevisiae were 0.05 μM, while that for E. coli was 2.0 μM. Moreover, PDI of S. cerevisiae was performed using other porphyrins (Type E, Figure 6), which were monocationic and highly hydrophobic. The PDI of S. cerevisiae occurred efficiently by Type E porphyrins [37]. The [P] values for the PDI of S. cerevisiae were optimized to be 0.005 μM. Thus, S. cerevisiae has low drug resistance for hydrophobic sensitizers rather than polycationic sensitizers, since the [P] value of tricationic porphyrins was larger than that of monocationic porphyrins (Type E). On the contrary, no PDI of E. coli by Type E porphyrins occurred at all. This result shows that a more positive character is required for an efficient PDI of E. coli.
Figure 4. Relationship between the $A_F$ values and number of carbon atoms ($n$) in the alkyl group on the alkylpyridinium (RPy) in PDI of E. coli using (A) P-porphyrins ((RPy)$_3$P(Tpp)$_{3+}$, ○) and Sb-porphyrins ((RPy)$_3$Sb(Tpp)$_{3+}$ and RPySb(Tpp)$_{2+}$, △) and (B) 3-alkyl-substituted P-porphyrins ((RPy)$_5$P(Tpp)$_{3+}$, ◻) and their 4-ethyl-analog ((4EtRPy)$_5$P(Tpp)$_{3+}$, ◼).

Figure 5. The incorporation of porphyrins inside bacteria through self-promoted mechanism. (i) Cationic porphyrin adsorbs to the anionic outer membrane; (ii) amphiphilic porphyrin interacts with hydrophobic parts of outer and inner membranes; (iii) porphyrin is incorporated inside the cell.
Photodynamic Inactivation of Escherichia coli with Cationic Porphyrin Sensitizers
DOI: http://dx.doi.org/10.5772/intechopen.82645

Figure 6. Fluorescence images of E. coli obtained with a CLSM under laser-excitation at 543 nm. Fluorescence coming from inside the cells was observed with the addition of (HexPy3)₂P(Tpp)³⁺ (D), but not observed with the addition of (MePy3)₂P(Tpp)³⁺ (A). Transmission images of E. coli containing (HexPy3)₂P(Tpp)³⁺ (E) and (MePy3)₂P(Tpp)³⁺ (B). The image of C is obtained by overlapping images in A and B, and the image in F is obtained by overlapping images in D and E.

4. Discussion

The mechanism behind the PDI activity in E. coli is still not completely understood. However, it is known that the first contact of porphyrin photosensitizers occurs at the outer membrane. The outer leaflet of the outer membrane mainly consists of lipopolysaccharides and phospholipids, which are negatively charged and are stabilized with divalent cations such as Ca²⁺ and Mg²⁺ [38]. Therefore, electrostatic interaction between cationic photosensitizers and the outer leaflet instead of these divalent cations promotes destabilization of the outer membrane [39]. In the case of the cationic porphyrins with hydrophobic character, or the amphiphilic one, they can also interact with not only the outer leaflet but also the inner leaflet of the outer membrane and the plasma membrane (Figure 7). Thus, the amphiphilic porphyrins may be incorporated inside E. coli cells via the self-promoted uptake pathway [37]. The porphyrin

![Type E porphyrins](image)

Figure 7. P₆-porphyrins (Type E) substituted with alkylethylene glycol ligands.
sensitizers passed through the cell wall may reach biogenic proteins, lipids, and DNA. Under irradiation, reactive oxygen such as $^{1}\text{O}_2$ was generated near to these molecules to induce cell death. Although E-type porphyrins generate $^{1}\text{O}_2$ efficiently under visible light irradiation, the lifetime of $^{1}\text{O}_2$ in aqueous medium is very short (~3 μs) [40]. Thus, for efficient PDI, $^{1}\text{O}_2$ should be generated as close as possible to the target molecules. The P type porphyrins with amphiphilic characters, which can be incorporated inside E. coli, will be advantageous to PDI via $^{1}\text{O}_2$ generation.

5. Conclusion
PDI of E. coli K-12 (IFO 3301) was examined using 19 kinds of cationic porphyrin sensitizers. In conclusion, (1) E. coli has high drug-resistance toward the hydrophobic and monocationic porphyrins such as Type E. (2) However, E. coli has low drug-resistance toward polycationic porphyrins such as Type P. (3) Especially, E. coli has low drug-resistance toward polycationic porphyrins with moderately long alkyl chain, for example, (HeptPy3)$_2$P(Tpp)$^{3+}$ and (EtPy5)$_2$P(Tpp)$^{3+}$. Alkyl chains might result in moderate hydrophobicity to take advantage of interaction between hydrophobic parts of cell membranes. (4) Polycationic porphyrins can interact with the anionic outer membrane at the first step and DNA and proteins inside the cells with strong binding affinities.

Acknowledgements
We thank Mr. Tomohiko Shinbara, Mr. Hiroki Kanemaru, Mr. Yusaku Suemoto, Mr. Kyosuke Takemori, Mr. Masato Shigehara, Mr. Kou Suzuki, Ms. Akari Miyamoto, and Hidekazu Uezono for their efforts on PDI of E. coli at University of Miyazaki.

Conflict of interest
The authors declare that they have no competing interests.

Abbreviations

$A_F$ PDI activity (in μM$^{-1}$ h$^{-1}$): $A_F = 60/([P] \times T_{1/2})$
$B$ mount of bacteria
$B_0$ initial amount of bacteria
$CFU$ colony formation unit
$C_W$ water solubility
$\varepsilon$ molar absorption coefficient
$LB$ Luria-Bertani medium
$m$ number of ethylene glycol unit
$n$ carbon number of the alkyl chain on the Ap
$[P]$ minimum effective concentrations of sensitizer
PDI photodynamic inactivation
$\text{RPy}$ N-alkylpyridinium group
$t$ irradiation time
$T_{1/2}$ half-life time required to reduce $B$ from $B_0$ to 0.5$B_0$
$Z$ valence number of the porphyrin complex
Abbreviations of substances

(\text{BrS}_5)\text{P(Tpp)}^+ \quad \text{bis}(5\text{-bromo-3-oxapentyloxo})\text{tetraphenylporphyrinato-phosphorus chloride}

(\text{PyS}_3)\text{P(Tpp)}^+ \quad \text{bis}[3\text{-}(4\text{-pyridyl})\text{propoxo}]\text{tetraphenylporphyrinato-phosphorus chloride}

(\text{PyS}_3)\text{Sb(Tpp)}^+ \quad \text{bis}[3\text{-}(4\text{-pyridyl})\text{propoxo}]\text{tetraphenylporphyrinato-antimony bromide}

\text{PySb(Tpp)}^+ \quad 3\text{-}(4\text{-Pyridyl})\text{propoxo}(\text{methoxo})\text{tetraphenylporphyrinato-antimony bromide}

\text{PyTpp} \quad \text{triphenyl}(4\text{-pyridinyl})\text{porphyrin}

(\text{RPyS}_3)\text{P(Tpp)}^{3+} \quad \text{bis}[3\text{-}(1\text{-alkyl-4-pyridinio})\text{propoxo}]\text{tetraphenylporphyrinato-phosphorus chloride, dihalide}

(\text{RPyS}_3)\text{Sb(Tpp)}^{3+} \quad \text{bis}[3\text{-}(1\text{-alkyl-4-pyridinio})\text{propoxo}]\text{tetraphenylporphyrinato-antimony trihalide}

(\text{RPyS}_5)\text{P(Tpp)}^{3+} \quad \text{bis}[5\text{-}(3\text{-alkyl-1-pyridinio})\text{oxapentyloxo}]\text{tetraphenylporphyrinato-antimony tribromide}

\text{RPySb(Tpp)}^{2+} \quad \alpha\text{-}(\text{methoxo})\text{-}[3\text{-}(1\text{-hexyl-4-pyridinio})\text{-1-propoxo}]\text{5,10,15,20-tetraphenylporphyrinato-antimony (V) dibromide}

(\text{R'b})\text{P(RPyTpp)}^{2+} \quad \text{bis}(2\text{-alkoxyethoxy})\text{-5-(1-alkyl-4-pyridinio)}\text{-10,15,20-triphenylporphyrinato-antimony (V) dichloride}

\text{meso-TMP} \quad \text{meso-tetra}[4\text{-}(1\text{-methylpyridinium})]\text{porphyrin}

Author details

Jin Matsumoto\textsuperscript{1}, Tomoko Matsumoto\textsuperscript{2}, Kazuya Yasuda\textsuperscript{3} and Masahide Yasuda\textsuperscript{1}*

1 Department of Applied Chemistry, Faculty of Engineering, University of Miyazaki, Miyazaki, Japan

2 Center for Collaborative Research and Community Cooperation, University of Miyazaki, Miyazaki, Japan

3 Department of Pharmacy, University of Miyazaki Hospital, Miyazaki, Japan

*Address all correspondence to: yasuda@cc.miyazaki-u.ac.jp
The Universe of Escherichia coli

References


Photodynamic Inactivation of Escherichia coli with Cationic Porphyrin Sensitizers
DOI: http://dx.doi.org/10.5772/intechopen.82645


