

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,400

Open access books available

117,000

International authors and editors

130M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Searching for Metabolic Pathways of Anaerobic Digestion: A Useful List of the Key Enzymes

*Anna Sikora, Anna Detman, Damian Mielecki,
Aleksandra Chojnacka and Mieczysław Błaszczak*

Abstract

The general scheme of anaerobic digestion is well known. It is a complex process promoted by the interaction of many groups of microorganisms and has four major steps: hydrolysis, acidogenesis, acetogenesis, and methanogenesis. The aim of the study was to prepare a systematized list of the selected enzymes responsible for the key pathways of anaerobic digestion based on the Kyoto Encyclopedia of Genes and Genomes database resource. The list contains (i) key groups of hydrolases involved in the process of degradation of organic matter; (ii) the enzymes catalyzing reactions leading to pyruvate formation; (iii) the enzymes of metabolic pathways of further pyruvate transformations; (iv) the enzymes of glycerol transformations; (v) the enzymes involved in transformation of gaseous or nongaseous products of acidic fermentations resulting from nonsyntrophic nutritional interactions between microbes; (vi) the enzymes of amino acid fermentations; (vii) the enzymes involved in acetogenesis; and (viii) the enzymes of the recognized pathways of methanogenesis. Searching for the presence and activity of the enzymes as well as linking structure and function of microbial communities allows to develop a fundamental understanding of the processes, leading to methane production. In this contribution, the present study is believed to be a piece to the enzymatic road map of anaerobic digestion research.

Keywords: anaerobic digestion, enzymes, hydrolysis, acidogenesis, acetogenesis, methanogenesis, syntrophy, metabolic pathways

1. Introduction

Anaerobic digestion (AD), whose final products are methane and carbon dioxide, is a common process in natural anoxic environments such as water sediments, wetlands, or marshlands. The environments have to be rich in organic matter and poor with other electron acceptors such as nitrate, compounds containing oxidized forms of metals, and sulfate. AD is also common in landfills and wastewater treatment plants and was used by man to produce biogas from waste biomass as an alternative energy source.

AD is a complex process that requires the metabolic interaction of many groups of microorganisms responsible for four closely related major steps. The first one is hydrolysis of complex organic polymers (e.g., polysaccharides, lipids, proteins) to

monomers (sugars, fatty acids, amino acids). The second step is acidogenesis that results in formation of hydrogen and carbon dioxide as well as nongaseous fermentation products, that is, low-molecular-weight organic acids and alcohols. These products are further oxidized to hydrogen, carbon dioxide, and acetate in acetogenic step that involves mainly syntrophic degradation of nongaseous fermentation products. The fourth step is methanogenesis. Three groups of substrates for methane production and three types of methanogenic pathways are known: splitting of acetate (acetoclastic/acetotrophic methanogenesis); reduction of CO₂ with H₂ or formate and rarely ethanol or secondary alcohols as electron donors (hydrogenotrophic methanogenesis); and reduction of methyl groups of methylated compounds such as methanol, methylated amines, or methylated sulfides (hydrogen-dependent and hydrogen-independent methylotrophic methanogenesis). The two last steps, acetogenesis and methanogenesis, are closely related and involve syntrophic associations between hydrogen-producing acetogenic bacteria and hydrogenotrophic methanogens (**Figure 1**) [1–5].

Recently, there has been a rapid development in culture-independent techniques (meta-omics approaches such as metagenomics, metatranscriptomics, metaproteomics, metabolomics) for exploring microbial communities, which have led to a new insight into their structure and function in both natural environments and anaerobic digesters. The current trends involve the combined use of meta-omic approaches and detailed reactor performance data as well as isotope labeling techniques that allow us to develop a fundamental understanding of the processes occurring in AD. Those activities are aimed to improve

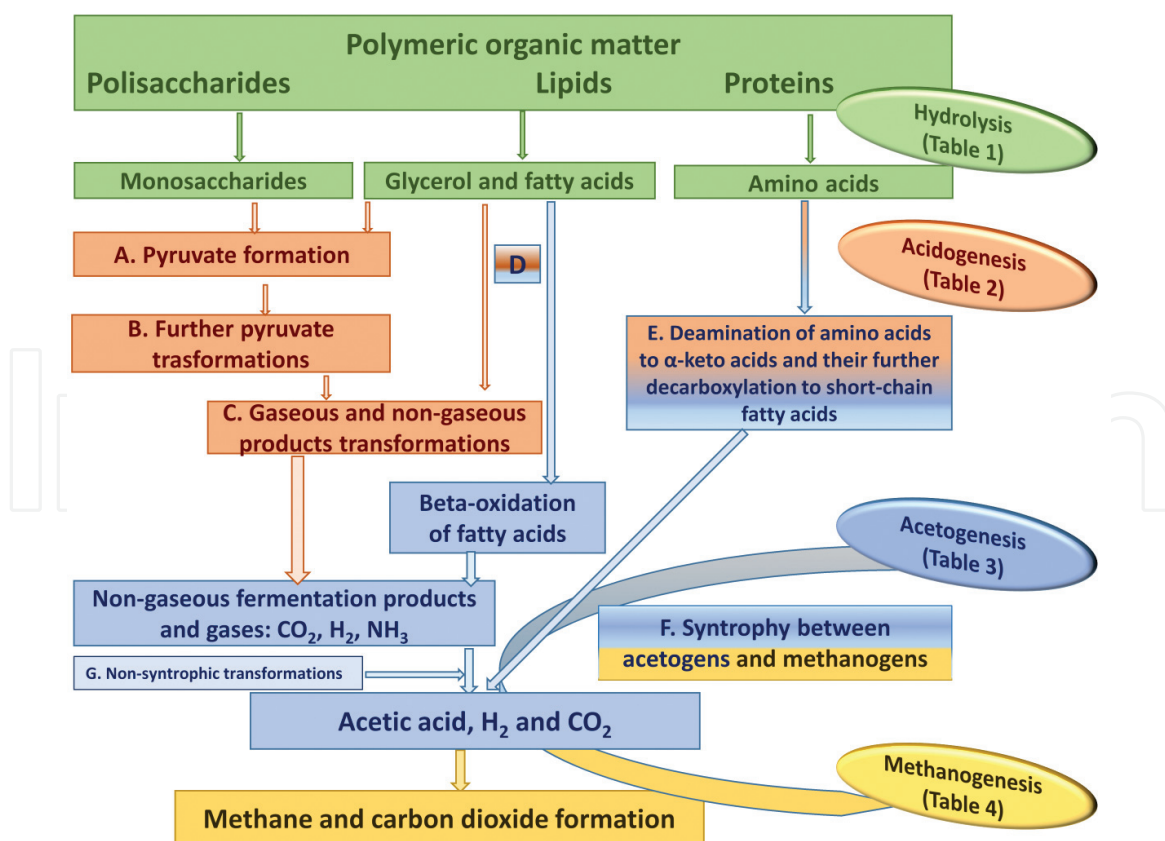


Figure 1. A scheme of anaerobic digestion of organic matter. Enzymes catalysing specific reactions of AD are presented in Tables 1–4. Thus in Figure 1 there are the links to Tables 1–4. Furthermore, background colours in the Figure correspond to the background colours of the title rows in the Tables 1–4: hydrolysis is indicated in green, acidogenesis in orange, acetogenesis in blue and methanogenesis in yellow. A, B, C, D, E refer to the title rows in Table 2; F, G refer to the title rows in Table 3.

biogas production and increase the share of renewable energy in total energy consumption [6–9].

Analysis of many studies on metagenomes of microbial communities from anaerobic digesters shows that (i) contribution of methanogens in the methane-yielding microbial communities is relatively small, below 20%; (ii) the most abundant phyla of bacteria are usually *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, and *Actinobacteria*; (iii) methanogenic archaea are dominated by acetotrophs or hydrogenotrophs with a certain contribution of methylotrophs; (iv) substrate, operational conditions such as temperature, pH, ammonia concentration, etc. shape the structure, percentage distribution of specific taxons, and functioning of the community of microorganisms; (v) it is important to describe interactions within microbial communities and assign functions in AD steps to specific groups of microbes; and (vi) the majority of sequences are not classified at the genus level confirming that most of the microorganisms are still unrecognized [6, 10–15].

In this contribution, the purpose of the study was to prepare a list of the selected enzymes and their catalyzed reactions, being a specific enzymatic road map of AD metabolic pathways, useful in molecular studies. The available metabolic pathway databases such as KEGG PATHWAY Database [16–18], MetaCyc Metabolic Pathway Database, BioCyc Database Collection [19], and BRENDA—The Comprehensive Enzyme Information System [20] were used to select metabolic pathways dedicated only to AD from hydrolysis to methanogenic steps exerted by microbes.

2. Selected enzymes of anaerobic digestion

Figure 1 shows a scheme of AD and **Tables 1–4** present a summary of the selected enzymes and enzymatic reactions involved in decomposition of organic matter to methane and carbon dioxide. **Tables 1–4** are an extension of **Figure 1**, and in **Figure 1**, there are the links to **Tables 1–4**.

The key groups of hydrolases involved in the process of degradation of organic matter are esterases, glycosidases, and peptidases, which catalyze the cleavage of ester bonds, glycoside bonds, and peptide bonds, respectively (**Table 1**). **Table 1** also includes other classes of hydrolases such as acting on carbon-nitrogen bonds, other than peptide bonds.

In the acidogenic stage of AD, the key step is pyruvate formation from carbohydrates (**Table 2**, Part A) or other compounds and further pyruvate transformations toward short-chain fatty acids and ethanol (**Table 2**, Part B). The Part C of the **Table 2** also considers transformation of gaseous and nongaseous products of acidic fermentations, resulting from nonsyntrophic nutritional interaction between bacteria. The Parts D and E present the enzymes of glycerol and amino acid transformations, respectively. The latter requires syntrophic cooperation between microorganisms.

The enzymes catalyzing oxidation of nongaseous products of acidogenesis mainly butyrate, propionate, acetate, lactate, ethanol including the enzymes of reverse electron transfer (process responsible for energy conservation in syntrophically growing acetogens) are shown in **Table 3**.

The enzymes of the three recognized pathways of methanogenesis such as acetotrophic, hydrogenotrophic, and methylotrophic are listed in **Table 4**.

The data were prepared on the basis of detailed analysis of AD research. The enzyme nomenclature comes from the Kyoto Encyclopedia of Genes and Genomes (KEGG) database resource.

| Hydrolytic enzyme | Reaction/process | EC number |
|--|---|--------------|
| Esterases | Acting on ester bonds | EC 3.1 |
| Glycosidases | Acting on glycoside bonds | EC 3.2 |
| Acting on cellulose | | |
| Cellulase; endo-1,4-beta-D-glucanase | Endohydrolysis of (1 → 4)-beta-D-glucosidic linkages in cellulose, lichenin, and cereal beta-D-glucans | EC 3.2.1.4 |
| Cellulose 1,4-beta-cellobiosidase (nonreducing end) | Hydrolysis of (1 → 4)-beta-D-glucosidic linkages in cellulose and cellotetraose, releasing cellobiose from the nonreducing ends of the chains | EC 3.2.1.91 |
| Beta-glucosidase | Hydrolysis of terminal, nonreducing beta-D-glucosyl residues with release of beta-D-glucose | EC 3.2.1.21 |
| Acting on hemicellulose | | |
| Endo-1,4-beta-xylanase | Endohydrolysis of (1 → 4)-beta-D-xylosidic linkages in xylans | EC 3.2.1.8 |
| Xylan 1,4-beta-xylosidase | Hydrolysis of (1 → 4)-beta-D-xylans, to remove successive D-xylose residues from the nonreducing termini | EC 3.2.1.37 |
| Mannan endo-1,4-beta-mannosidase | Random hydrolysis of (1 → 4)-beta-D-mannosidic linkages in mannans, galactomannans, and glucomannans | EC 3.2.1.78 |
| Beta-mannosidase | Hydrolysis of terminal, nonreducing beta-D-mannose residues in beta-D-mannosides | EC 3.2.1.25 |
| Alpha-galactosidase | Hydrolysis of terminal, nonreducing alpha-D-galactose residues in alpha-D-galactosides, including galactose oligosaccharides, galactomannans, and galactolipids | EC 3.2.1.22 |
| Alpha-glucuronidase | An alpha-D-glucuronoside + H ₂ O → an alcohol + D-glucuronate | EC 3.2.1.139 |
| Peptidases | Acting on peptide bonds | EC 3.4 |
| <i>Other hydrolases</i> | | |
| Hydrolases acting on carbon-nitrogen bonds, other than peptide bonds | | EC 3.5 |
| Hydrolases acting on ether bonds | | EC 3.3 |
| Hydrolases acting on carbon-carbon bonds | | EC 3.7 |
| Hydrolases acting on halide bonds | | EC 3.8 |
| Hydrolases acting on phosphorus-nitrogen bonds | | EC 3.9 |
| Hydrolases acting on sulfur-nitrogen bonds | | EC 3.10 |
| Hydrolases acting on carbon-phosphorus bonds | | EC 3.11 |
| Hydrolases acting on sulfur-sulfur bonds | | EC 3.12 |
| Hydrolases acting on carbon-sulfur bonds | | EC 3.13 |
| Hydrolases acting on acid anhydrides | | EC 3.6 |

Table 1.
The selected enzymes of hydrolytic step of anaerobic digestion [21, 22].

| Enzyme | Reaction/process | EC number |
|--|---|--------------|
| A. Pyruvate formation from carbohydrates [23] | | |
| <i>Glycolysis (the Embden-Meyerhof-Parnas pathway)</i> | | |
| Hexose kinase | D-Glucose + ATP \leftrightarrow D-glucose-6-phosphate + ADP | EC 2.7.1.1 |
| Phosphoglucose isomerase | D-Glucose 6-phosphate \leftrightarrow D-fructose 6-phosphate | EC 5.3.1.9 |
| Phosphofructose kinase | ATP + D-fructose 6-phosphate \leftrightarrow ADP + D-fructose 1,6-bisphosphate | EC 2.7.1.11 |
| Fructose-bisphosphate aldolase | Fructose-1,6-bisphosphate \leftrightarrow dihydroxyacetone phosphate + glyceraldehyde-3-phosphate | EC 4.1.2.13 |
| Triose phosphate isomerase | Glyceraldehyde 3-phosphate \leftrightarrow dihydroxyacetone phosphate | EC 5.3.1.1 |
| Glyceraldehyde-3-phosphate dehydrogenase | D-Glyceraldehyde 3-phosphate + phosphate + NAD ⁺ \leftrightarrow 1,3-bisphosphoglycerate + NADH + H ⁺ | EC 1.2.1.12 |
| Phosphoglycerate kinase | 1,3-Bisphosphoglycerate + ADP \leftrightarrow 3-phosphoglycerate + ATP | EC 2.7.2.3 |
| Phosphoglycerate mutase | 3-Phosphoglycerate \leftrightarrow 2-phosphoglycerate | EC 5.4.2.1 |
| Enolase | 2-Phospho-D-glycerate \leftrightarrow phosphoenolpyruvate + H ₂ O | EC 4.2.1.11 |
| Pyruvate kinase | Phosphoenolpyruvate + ADP \leftrightarrow pyruvate + ATP | EC 2.7.1.40 |
| 2-Keto-3-deoxy-6-phosphogluconate (the Entner-Doudoroff pathway) | | |
| Glucose-6-phosphate dehydrogenase | D-glucose 6-phosphate + NADP ⁺ \leftrightarrow 6-phospho-D-glucono-1,5-lactone + NADPH + H ⁺ | EC 1.1.1.49 |
| Phosphogluconate dehydrogenase | 6-Phospho-D-gluconate + NAD(P) ⁺ \leftrightarrow 6-phospho-2-dehydro-D-gluconate + NAD(P)H + H ⁺ | EC 1.1.1.43 |
| 2-Keto-3-deoxy-6-phosphogluconate aldolase | 2-Dehydro-3-deoxy-6-phospho-D-gluconate \leftrightarrow pyruvate + D-glyceraldehyde 3-phosphate | EC 4.1.2.14 |
| B. Further transformations of pyruvate—glycolytic fermentations [23–27] | | |
| Lactate dehydrogenase | Pyruvate + NADH \leftrightarrow lactate + NAD ⁺ | EC 1.1.1.27 |
| Pyruvate:ferredoxin oxidoreductase, PFOR | Pyruvate + CoA + oxidized Fd \leftrightarrow acetyl-CoA + reduced Fd + CO ₂ + H ⁺ | EC 1.2.7.1 |
| NADH:ferredoxin oxidoreductase, NFOR | Oxidized Fd + NADH \leftrightarrow reduced Fd + NAD ⁺ + H ⁺ | EC 1.18.1.3 |
| Ferredoxin hydrogenase | 2 reduced ferredoxin + 2 H ⁺ \leftrightarrow H ₂ + 2 oxidized ferredoxin | EC 1.12.7.2 |
| Phosphotransacetylase | CoA + acetyl phosphate \leftrightarrow acetyl-CoA + phosphate | EC 2.3.1.8 |
| Acetate kinase | ATP + acetate \leftrightarrow ADP + acetyl phosphate | EC 2.7.2.1 |
| NAD ⁺ -dependent ethanol dehydrogenase | Acetaldehyde + NADH + H ⁺ \leftrightarrow ethanol + NAD ⁺ An aldehyde + NADH + H ⁺ \leftrightarrow a primary alcohol + NAD ⁺ | EC 1.1.1.1 |
| Acetaldehyde dehydrogenase | Acetaldehyde + CoA + NAD ⁺ \leftrightarrow acetyl-CoA + NADH + H ⁺ | EC 1.2.1.10 |
| Acetyl-CoA acetyltransferase | 2-acetyl-CoA \leftrightarrow CoA + acetoacetyl-CoA | EC 2.3.1.9 |
| 3-Hydroxybutyryl-CoA dehydrogenase | 3-Acetoacetyl-CoA + NADPH + H ⁺ \leftrightarrow 3-hydroxybutanoyl-CoA + NADP ⁺ | EC 1.1.1.157 |
| Crotonase 3-OH-butyryl-CoA dehydratase | 3-Hydroxybutanoyl-CoA \leftrightarrow crotonoyl-CoA + H ₂ O | EC 4.2.1.55 |

| Enzyme | Reaction/process | EC number |
|--|--|--------------|
| 2NADH+ oxidized Fd + crotonyl-CoA → 2 NAD+ reduced Fd + butyryl-CoA catalyzed by butyryl CoA dehydrogenase/electron-transfer flavoprotein complex | | |
| Butyryl-CoA dehydrogenase | A short-chain acyl-CoA + electron-transfer flavoprotein ↔ a short-chain trans-2,3-dehydroacyl-CoA + reduced electron-transfer flavoprotein | EC 1.3.8.1 |
| Butyryl-CoA dehydrogenase/Etf complex | Butanoyl-CoA + 2 NAD ⁺ + 2 reduced Fd ↔ Crotonoyl-CoA + 2 NADH + 2 oxidized Fd | EC 1.3.1.109 |
| Phosphotransbutyrylase | Butanoyl-CoA + phosphate ↔ CoA + butanoyl phosphate | EC 2.3.1.19 |
| Butyrate kinase | Butanoyl phosphate + ADP ↔ butanoate + ATP | EC 2.7.2.7 |
| PFL—pyruvate formate lyase | Pyruvate + CoA ↔ acetyl-CoA + formate | EC 2.3.1.54 |
| FHL—formate hydrogen lyase | Formate → H ₂ + CO ₂ | EC 1.17.99.7 |
| Pyruvate carboxylase | ATP + pyruvate + HCO ₃ ⁻ ↔ ADP + phosphate + oxaloacetate | EC 6.4.1.1 |
| Malate dehydrogenase | Malate + NAD ⁺ ↔ oxaloacetate + NADH + H ⁺ | EC 1.1.1.37 |
| Fumarate hydratase | Malate ↔ fumarate + H ₂ O | EC 4.2.1.2 |
| Fumarate reductase | Fumarate + a quinol ↔ succinate + a quinone | EC 1.3.5.4 |
| | Fumarate + NADH ↔ succinate + NAD ⁺ | EC 1.3.1.6 |
| Succinyl-CoA synthetase | GTP + succinate + CoA = GDP + phosphate + succinyl-CoA | EC 6.2.1.4 |
| Methylmalonyl CoA mutase | Succinyl-CoA ↔ (R)-methylmalonyl-CoA | EC 5.4.99.2 |
| Methylmalonyl CoA epimerase | (R)-methylmalonyl-CoA ↔ (S)-methylmalonyl-CoA | EC 5.1.99.1 |
| Methylmalonyl-CoA decarboxylase | (S)-methylmalonyl-CoA ↔ propanoyl-CoA + CO ₂ | EC 4.1.1.41 |
| Propionate-CoA transferase | Acetate + propanoyl-CoA ↔ acetyl-CoA + propanoate | EC 2.8.3.1 |
| C. Transformation of gaseous and nongaseous products of acidic fermentations (the selected examples) | | |
| <i>Transformation of lactate and acetate to butyrate, hydrogen, and carbon dioxide ([28] and cited therein)</i> | | |
| Lactate dehydrogenases | (S)-lactate + NAD ⁺ ↔ pyruvate + NADH + H ⁺ | EC 1.1.1.27 |
| | (R)-lactate + NAD ⁺ ↔ pyruvate + NADH + H ⁺ | EC 1.1.1.28 |
| | Lactate + 2 NAD ⁺ + 2 reduced Fd ↔ pyruvate + 2 NADH + 2 oxidized Fd | EC 1.3.1.110 |
| See Table 3 | | |
| Pyruvate is oxidized to acetyl coenzyme A, which is further routed to acetate and butyrate with hydrogen release. See Part B: Further transformations of pyruvate—glycolytic fermentations | | |
| <i>Transformation of ethanol and acetate to butyrate and hydrogen in Clostridium kluveri [29]</i> | | |
| Acetate kinase | See Part B. Further transformations of pyruvate—glycolytic fermentations | EC 2.7.2.1 |
| Acetyl-CoA acetyltransferase | | EC 2.3.1.9 |
| 3-Hydroxybutyryl-CoA dehydrogenase | | EC 1.1.1.157 |
| 3-Hydroxyacyl-CoA dehydratase | | EC 4.2.1.55 |
| Butyryl-CoA dehydrogenase/Etf complex | | EC 1.3.1.109 |

| Enzyme | Reaction/process | EC number |
|---|--|--------------|
| Acetate CoA-transferase | Acyl-CoA + acetate ↔ a fatty acid anion + acetyl-CoA | EC 2.8.3.8 |
| Reductive carbon monoxide dehydrogenase/acetyl-CoA synthase pathway (reductive CODH/ACS) [30] | | |
| NADP-dependent formate dehydrogenase | CO ₂ + NADPH ↔ formate + NADP ⁺ | EC 1.17.1.10 |
| Formyltetrahydrofolate synthetase | ATP + formate + tetrahydrofolate ↔ ADP + phosphate + 10-formyltetrahydrofolate | EC 6.3.4.3 |
| Methenyltetrahydrofolate cyclohydrolase | 10-Formyltetrahydrofolate ↔ 5,10-methenyltetrahydrofolate + H ₂ O | EC 3.5.4.9 |
| NADP-dependent methylenetetrahydrofolate dehydrogenase | 5,10-Methenyltetrahydrofolate + NADPH + H ⁺ ↔ 5,10-Methylenetetrahydrofolate + NADP ⁺ | EC 1.5.1.5 |
| Ferredoxin-dependent methylenetetrahydrofolate reductase | 5,10-Methylenetetrahydrofolate + 2 reduced Fd + 2 H ⁺ ↔ 5-methyltetrahydrofolate + 2 oxidized Fd | EC 1.5.7.1 |
| 5,10-Methylenetetrahydrofolate reductase | 5,10-Methylenetetrahydrofolate + NAD(P)H + H ⁺ ↔ 5-methyltetrahydrofolate + NAD(P) ⁺ | EC 1.5.1.20 |
| 5-Methyltetrahydrofolate: corrinoid/iron-sulfur protein Co-methyltransferase | [Co(I) corrinoid Fe-S protein] + 5-methyltetrahydrofolate ↔ [methyl-Co(III) corrinoid Fe-S protein] + tetrahydrofolate | EC 2.1.1.258 |
| Carbon monoxide dehydrogenase | CO ₂ + 2 reduced Fd + 2 H ⁺ ↔ CO + H ₂ O + 2 oxidized Fd | EC 1.2.7.4 |
| CO-methylating acetyl-CoA synthase | CO + CoA + [methyl-Co(III) corrinoid Fe-S protein] ↔ acetyl-CoA + [Co(I) corrinoid Fe-S protein] | EC 2.3.1.169 |
| D. Glycerol transformations [31, 32] | | |
| <i>Oxidative pathway</i> | | |
| Glycerol dehydrogenase | Glycerol + NAD ⁺ ↔ glycerone (dihydroxyacetone) + NADH + H ⁺ | EC 1.1.1.6 |
| Dihydroxyacetone kinase | ATP + glycerone ↔ ADP + glycerone phosphate | EC 2.7.1.29 |
| For further reactions, see Part A: Pyruvate formation | | |
| <i>Reductive pathway</i> | | |
| Glycerol dehydratase | Glycerol ↔ 3-hydroxypropionaldehyde + H ₂ O | EC 4.2.1.30 |
| 1,3-Propanediol dehydrogenase | 3-Hydroxypropionaldehyde + NADH + H ⁺ ↔ 1,3-propanediol + NAD ⁺ | EC 1.1.1.202 |
| E. Amino acids fermentations [33–37] | | |
| <p>Syntrophy with H₂-scavenging microorganism: amino acid degradation involves NAD(P)- or FAD-dependent deamination of amino acids to the corresponding α-keto acids by amino acid dehydrogenases (EC 1.4.1.X): RCH(NH₄⁺)COO⁻ + H₂O → RCOCOO⁻ + NH₄⁺ + H₂ and further conversion of α-keto acids via oxidative decarboxylation to fatty acids: RCOCOO⁻ + H₂O → RCOO⁻ + CO₂ + H₂ [33]</p> | | |
| <p>Without syntrophy with H₂-scavenging microorganism: Stickland Reaction—coupled oxidation-reduction reactions between suitable amino acids (coupled deamination of amino acids); one member of the pair is oxidized (dehydrogenated) and the other is reduced (hydrogenated) [34], for example,</p> <p>Alanine and glycine: alanine + 2 glycine + 3H₂O → 3 acetate⁻ + 3NH₄⁺ + HCO₃⁻ + H⁺ Valine and glycine: valine + 2 glycine + 3H₂O → isobutyrate⁻ + 2 acetate⁻ + 3NH₄⁺ + HCO₃⁻ + H⁺ Leucine and glycine: leucine + 2 glycine + 3H₂O → isovalerate⁻ + 2 acetate⁻ + 3NH₄⁺ + HCO₃⁻ + H⁺</p> | | |

| Enzyme | Reaction/process | EC number |
|---|--|-------------|
| Examples of amino acid dehydrogenases catalyzing deamination of amino acids to the corresponding α-keto acids [33] | | |
| Aspartate dehydrogenase | $L\text{-aspartate} + H_2O + NAD(P)^+ \leftrightarrow \text{oxaloacetate} + NH_3 + NAD(P)H + H^+$ | EC 1.4.1.21 |
| Valine dehydrogenase | $L\text{-valine} + H_2O + NADP^+ \leftrightarrow 3\text{-methyl-2-oxobutanoate} + NH_3 + NADPH + H^+$ | EC 1.4.1.8 |
| Alanine dehydrogenase | $L\text{-alanine} + H_2O + NAD^+ \leftrightarrow \text{pyruvate} + NH_3 + NADH + H^+$ | EC 1.4.1.1 |
| Leucine dehydrogenase | $L\text{-leucine} + H_2O + NAD^+ \leftrightarrow 4\text{-methyl-2-oxopentanoate} + NH_3 + NADH + H^+$ | EC 1.4.1.9 |
| Key enzymes of Stickland reaction [34–36] | | |
| Glycine reductase GR pathway (<i>grd</i> operon) | | |
| Glycine reductase | $\text{Glycine} + \text{phosphate} + \text{reduced thioethoxin} + H^+ \leftrightarrow \text{acetyl phosphate} + NH_3 + \text{oxidized thioethoxin} + H_2O$ | EC 1.21.4.2 |
| Acetate kinase | $\text{Acetyl phosphate} + ADP \leftrightarrow \text{acetate} + ATP$ | EC 2.7.2.1 |
| Proline reductase PR pathway (<i>prd</i> operon) | | |
| D-proline reductase (dithiol) | $D\text{-proline} + \text{dihydroliipoate} \leftrightarrow 5\text{-aminopentanoate (5-aminovalerate)} + \text{lipoate}$ | EC 1.21.4.1 |
| Others examples [33] | | |
| Serine dehydratase | $L\text{-serine} \leftrightarrow \text{pyruvate} + NH_3$ (overall reaction) (1a) $L\text{-serine} \leftrightarrow 2\text{-aminoprop-2-enoate} + H_2O$ (1b) $2\text{-Aminoprop-2-enoate} \leftrightarrow 2\text{-iminopropanoate}$ (spontaneous) (1c) $2\text{-Iminopropanoate} + H_2O \leftrightarrow \text{pyruvate} + NH_3$ (spontaneous) | EC 4.3.1.17 |
| Threonine dehydratase | $L\text{-threonine} \leftrightarrow 2\text{-oxobutanoate} + NH_3$ (overall reaction) (1a) $L\text{-threonine} \leftrightarrow 2\text{-aminobut-2-enoate} + H_2O$; (1b) $2\text{-Aminobut-2-enoate} \leftrightarrow 2\text{-iminobutanoate}$ (spontaneous) (1c) $2\text{-Iminobutanoate} + H_2O \leftrightarrow 2\text{-oxobutanoate} + NH_3$ (spontaneous) | EC 4.3.1.19 |
| Detailed pathways of glutamate fermentation via 3-methylaspartate [37] | | |
| Glutamate mutase (methylaspartate mutase) | $L\text{-glutamate} \leftrightarrow L\text{-threo-3-methylaspartate}$ | EC 5.4.99.1 |
| Methyl aspartase | $L\text{-threo-3-methylaspartate} \leftrightarrow \text{mesaconate (2-methylfumarate)} + NH_3$ | EC 4.3.1.2 |
| Mesaconase (2-methylmalate dehydratase) | $2\text{-Methylfumarate} + H_2O \leftrightarrow (S)\text{-2-methylmalate}$ | 4.2.1.34 |
| Citramalate lyase | $(2S)\text{-2-hydroxy-2-methylbutanedioate} \leftrightarrow \text{acetate} + \text{pyruvate}$ $(S)\text{-2-methylmalate} = 2\text{-hydroxy-2-methylbutanedioate}$ | 4.1.3.22 |
| For further transformations of pyruvate to acetate and butyrate, see Part B. | | |
| For further transformations of pyruvate to propionate, see Part B. | | |
| Detailed pathway of glutamate fermentation via 2-hydroxyglutarate [37] | | |
| Glutamate dehydrogenase | $L\text{-glutamate} + H_2O + NAD^+ \leftrightarrow 2\text{-oxoglutarate} + NH_3 + NADH + H^+$ | 1.4.1.2 |

| Enzyme | Reaction/process | EC number |
|--|---|--------------|
| 2-Hydroxyglutarate dehydrogenase | (S)-2-hydroxyglutarate + acceptor ↔ 2-oxoglutarate + reduced acceptor | 1.1.99.2 |
| Glutaconate (2-hydroxyglutarate) CoA-transferase | Acetyl-CoA + (E)-glutaconate ↔ acetate + glutaconyl-1-CoA | 2.8.3.12 |
| 2-Hydroxyglutaryl-CoA dehydratase | (R)-2-hydroxyglutaryl-CoA ↔ (E)-glutaconyl-CoA + H ₂ O | EC 4.2.1.167 |
| Glutaconyl-CoA decarboxylase | 4-Carboxybut-2-enoyl-CoA ↔ but-2-enoyl-CoA + CO ₂ | 4.1.1.70 |

Table 2.
 The selected enzymes of acidogenic step of anaerobic digestion. A, B, C, D, and E refer to the processes indicated in Figure 1.

| Enzyme | Reaction/process | EC number |
|---|---|-------------------------------|
| F. Acetogenesis dependent on syntrophic relations between microorganisms | | |
| Acetate oxidation by, for example, <i>Clostridium ultunense</i>—oxidative carbon monoxide dehydrogenase/acetyl-CoA synthase pathway (oxidative CODH/ACS): | | |
| Acetate ⁻ + 4H ₂ O → 2 HCO ₃ ⁻ + 4H ₂ + H ⁺ , ΔG ^{0'} = + 104.6 kJ/mol, with the H ₂ consuming methanogen, ΔG ^{0'} = -31.0 kJ/mol [38] | | |
| | NADP-dependent formate dehydrogenase | See Table 2, Part C |
| | Formyltetrahydrofolate synthetase | |
| | Methenyltetrahydrofolate cyclohydrolase | |
| | NADP-dependent methylenetetrahydrofolate dehydrogenase | |
| | Ferredoxin-dependent methylenetetrahydrofolate reductase | |
| | 5,10-Methylenetetrahydrofolate reductase | |
| | 5-Methyltetrahydrofolate:corrinoid/iron-sulfur protein Co-methyltransferase | |
| | Carbon monoxide dehydrogenase | |
| | CO-methylating acetyl-CoA synthase | |
| Reverse electron transfer during acetate oxidation has yet to be confirmed. Direct interspecies electron transfer (DIET) is not excluded (Westerholm et al., 2016) | | |
| Acetate oxidation by <i>Geobacter sulfurreducens</i>: | | |
| Acetate oxidation coupled to reduction of fumarate to succinate (ΔG ^{0'} = -249 kJ per mol acetate), acetate metabolism proceeds via reactions of the citric acid cycle [39] | | |
| | Acetate kinase | See Table 2, Part B |
| | Phosphotransacetylase | |
| <i>Citric acid cycle</i> | | |
| Citrate synthase | Acetyl-CoA + H ₂ O + oxaloacetate ↔ citrate + CoA | EC 2.3.3.1 |
| Aconitase | Citrate ↔ isocitrate (overall reaction) | EC 4.2.1.3 |
| Isocitrate dehydrogenase (NADP ⁺ -dependent) | Isocitrate + NADP ⁺ ↔ 2-oxoglutarate + CO ₂ + NADPH + H ⁺ | EC1.1.1.42 |
| 2-Oxoglutarate:ferredoxin oxidoreductase | 2-Oxoglutarate + CoA + 2 oxidized Fd = succinyl-CoA + CO ₂ + 2 reduced Fd + 2 H ⁺ | EC 1.2.7.3 |
| Succinyl-CoA:acetate CoA-transferase | Succinyl-CoA + acetate ↔ acetyl-CoA + succinate | EC 2.8.3.18 |

| Enzyme | Reaction/process | EC number |
|---|--|-----------------------------|
| Succinate dehydrogenase | succinate + a quinone ↔ fumarate + a quinol | EC 1.3.5.1 |
| Fumarate hydratase | (S)-malate ↔ fumarate + H ₂ O | EC 4.2.1.2 |
| Malate dehydrogenase | (S)-malate + NAD ⁺ ↔ oxaloacetate + NADH + H ⁺ | EC 1.1.1.37 |
| Butyrate oxidation by <i>Syntrophomonas wolfei</i>: | | |
| Butyrate ⁻ + 2H ₂ O → 2 acetate ⁻ + 2H ⁺ + 2H ₂ , ΔG ^{0'} = + 48.3 kJ/mol, with the H ₂ consuming methanogen, ΔG ^{0'} = -17.3 kJ/mol [4] | | |
| CoA transferase | Butyrate + acetyl-CoA ↔ butyryl-CoA + acetate | EC 2.8.3.9 |
| Butyryl-CoA dehydrogenase | | See Table 2 , Part B |
| Crotonase-3-OH-butyryl-CoA dehydratase | | |
| 3-Acetyl-CoA acetyltransferase | | |
| Hydroxybutyryl-CoA dehydrogenase | | |
| Phosphotransacetylase | | |
| Acetate kinase | | |
| <p>Butyrate oxidation coupled with a reverse electron transfer that involves electron transfer flavoprotein EtfAB, membrane-anchored electron carrier DUF224 protein, the menaquinone pool in the membrane, a membrane-bound cytochrome, NADH:hydrogenase/formate-dehydrogenase complex (NDH/HYD1/FDH-1 complex), Rnf (proton-translocating ferredoxin:NAD⁺ oxidoreductase) [40]</p> | | |
| Propionate oxidation by <i>Syntrophobacter wolinii</i>: | | |
| Propionate ⁻ + 3H ₂ O → acetate ⁻ + HCO ₃ ⁻ + H ⁺ + 3H ₂ , ΔG ^{0'} = + 76.0 kJ/mol, with the H ₂ consuming methanogen, ΔG ^{0'} = -22.4 kJ/mol [4] | | |
| Pyruvate carboxylase | | See Table 2 , Part B |
| Malate dehydrogenase | | |
| Fumarate hydratase | | |
| Fumarate reductase | | |
| Succinate dehydrogenase | Succinate + a quinone ↔ fumarate + a quinol | EC 1.3.5.1 |
| Succinyl-CoA synthetase | | See Table 2 , Part B |
| Methylmalonyl CoA mutase | | |
| Methylmalonyl CoA epimerase | | |
| Methylmalonyl-CoA decarboxylase | | |
| Propionate-CoA transferase | | |
| <p>Propionate oxidation coupled with a reverse electron transfer that involves menaquinone, proteins encoded by cytochrome c homologous genes, cytochrome b:quinone oxidoreductases, formate dehydrogenases, hydrogenases including confurcating [FeFe]-hydrogenases [41]</p> | | |
| Six syntrophy-specific functional domains found in the genomes of the butyrate- or propionate-oxidizing syntrophs [42] | | InterPro number |
| Extra-cytoplasmic formate dehydrogenase (FDH) alpha subunit, EC 1.17.1.9 | | IPR006443 |
| FdhE-like protein—tightly connected with FDH | | IPR024064 |

| Enzyme | Reaction/process | EC number |
|---|--|--------------------------------|
| FDH accessory protein—tightly connected with FDH | | IPR006452 |
| CapA—a membrane-bound complex, a protein involved in capsule or biofilm formation that may facilitate syntrophic growth (<i>also present in acetate-oxidizers</i>) | | IPR019079 |
| FtsW, RodA, SpoVE—membrane-integrated proteins involved in membrane integration, cell division, sporulation, and shape determination | | IPR018365 |
| Ribonuclease P involved in tRNA maturation | | IPR020539 |
| Functional domains involved in electron transfer identified by [42] | InterPro number | |
| Cytoplasmic FDH | IPR027467, IPR006655, IPR006478, IPR019575, IPR001949 | |
| Extracytoplasmic FDH | IPR006443 | |
| Formate transporter | IPR000292, IPR024002 | |
| Fe-Fe hydrogenase | IPR004108, IPR009016, IPR003149, IPR013352 | |
| NiFe hydrogenase | IPR001501, IPR018194 | |
| Rnf complex: 2 reduced Fd + NAD ⁺ + H ⁺ + Na ⁺ ↔ 2 oxidized Fd + NADH + Na ⁺ (EC 1.18.1.8) | IPR007202, IPR010207, IPR026902, IPR010208, IPR004338, IPR011303, IPR007329 | |
| Ech complex: 2 reduced Fd + NADP ⁺ + H ⁺ ↔ 2 oxidized Fd + NADPH (EC 1.18.1.2) | IPR001750, IPR001516, IPR001694, IPR006137, IPR001268, IPR012179, IPR001135 | |
| Etf alpha, Etf beta, Bcd (Butyryl-CoA dehydrogenase): see Table 2 , Part B (EC 1.3.1.109) | IPR014731, IPR012255, IPR006089, IPR009075, IPR006092, IPR006091, IPR013786, IPR009100 | |
| Cytochromes: c cIII b561 b5 | IPR023155, IPR024673 IPR020942, IPR002322 IPR016174, IPR000516 IPR001199 | |
| DUF224 protein complex | IPR003816, IPR004017, IPR023234 | |
| Lactate oxidation by <i>Desulfovibrio vulgaris</i>: | | |
| $\text{Lactate}^- + \text{H}_2\text{O} \rightarrow \text{acetate}^- + \text{CO}_2 + 4 \text{H}_2$, $\Delta G^{0'} = -8.8 \text{ kJ/mol}$ with the H ₂ consuming methanogen, $\Delta G^{0'} = -74.2 \text{ kJ/mol}$ [43] | | |
| Lactate dehydrogenase | | See Table 2 , Part B |
| Pyruvate:ferredoxin oxidoreductase | | |
| Phosphate acetyltransferase | | |
| Acetate kinase | | |
| Alcohol dehydrogenase | | |
| Lactate oxidation coupled with a reverse electron transfer that involves the membrane-bound Qmo complex, cytochromes, hydrogenases (Coo, Hyn, Hyd, Hys), formate dehydrogenases, menaquinone, membrane-bound Qrc complex [43, 44] | | |

| Enzyme | Reaction/process | EC number |
|---|---|-----------------------------|
| Ethanol oxidation by <i>Pelobacter carbinolicus</i> | | |
| Ethanol + H ₂ O → acetate ⁻ + H ⁺ + 2H ₂ , ΔG ^{0'} = + 9.6 kJ/mol with the H ₂ consuming methanogen, ΔG ^{0'} = - 56 kJ/mol [4] | | |
| | NAD ⁺ -dependent ethanol dehydrogenase | See Table 2 , Part B |
| | Acetaldehyde dehydrogenase (acetylating) | |
| Nonacetylating acetaldehyde dehydrogenase | An aldehyde + NAD ⁺ + H ₂ O ↔ a carboxylate + NADH + H ⁺ | EC 1.2.1.3 |
| | Phosphotransacetylase | See Table 2 , Part B |
| | Acetate kinase | |
| Ethanol oxidation coupled with a reverse electron transfer that involves membrane-bound ion-translocating ferredoxin:NAD ⁺ oxidoreductase, formate dehydrogenases, and confurcating hydrogenases [1, 45] | | |
| G. Acetogenesis independent on syntrophic relations between microorganisms | | |
| Ethanol oxidation by <i>Acetobacterium woodii</i>: 2 ethanol + 2 CO₂ → 3 acetate—75.4 kJ/mol [46] | | |
| Bifunctional acetaldehyde-CoA/alcohol dehydrogenase | Ethanol + NAD ⁺ → acetaldehyde + NADH + H ⁺ acetaldehyde + NAD ⁺ + CoA → acetyl-CoA + 2 NADH + H ⁺ Ethanol is oxidized to acetyl-CoA in a two-step reaction by a bifunctional acetylating ethanol/aldehyde dehydrogenase | [EC:1.2.1.10 1.1.1.1] |
| | Acetyl-CoA is transformed to acetate with the release of ATP | See Table 2 , Part B |
| | Reduction of ferredoxin by NADH by reverse electron flow in a reaction catalyzed by Rnf complex | See Part F |
| | Carbon dioxide is reduced to acetate via the Wood-Ljungdahl pathway | See Table 2 , Part C |
| Lactate oxidation by <i>Acetobacterium woodii</i>: 2 lactate → 3 acetate—61 kJ/mol [47] | | |
| Lactate dehydrogenase | Lactate + 2 NAD ⁺ + 2 reduced Fd ↔ pyruvate + 2 NADH + 2 oxidized Fd The enzyme uses flavin-based electron confurcation to drive endergonic lactate oxidation with NAD ⁺ as oxidant at the expense of simultaneous exergonic electron flow from reduced ferredoxin to NAD ⁺ | EC 1.3.1.110 |
| | Pyruvate is transformed to acetyl-CoA and further to acetate with the release of ATP | See Table 2 , Part B |
| | Reduction of ferredoxin by NADH by reverse electron flow in a reaction catalyzed by Rnf complex | See Part F |
| | Carbon dioxide is reduced to acetate via the Wood-Ljungdahl pathway | See Table 2 , Part C |

Table 3.
The selected enzymes of acetogenic step of anaerobic digestion. F and G refer to the processes indicated in Figure 1.

| Enzyme | Reaction/process | EC number |
|---|--|--|
| MFR—methanofuran, H-S-CoM—coenzyme M, H-S-CoB—coenzyme B, H ₄ MPT—tetrahydromethanopterin, F ₄₂₀ —5′deazaflavin, H ₄ SPT—tetrahydrosarcinapterin | | |
| Hydrogenotrophic pathway | | |
| Formylmethanofuran dehydrogenase | CO ₂ + MFR + 2 reduced Fd + 2H ⁺ ↔ formyl-MFR + H ₂ O + 2 oxidized Fd | EC 1.2.7.12 |
| Formylmethanofuran-H ₄ MPT formyltransferase | Formyl-MFR + H ₄ MPT ↔ MFR + formyl-H ₄ MPT | EC 2.3.1.101 |
| Methenyl-H ₄ MPT cyclohydrolase | Formyl-H ₄ MPT + H ⁺ ↔ methenyl-H ₄ MPT + H ₂ O | EC 3.5.4.27 |
| F ₄₂₀ -dependent methylene-H ₄ MPT dehydrogenase | Methenyl-H ₄ MPT + reduced F ₄₂₀ ↔ methylene-H ₄ MPT + oxidized F ₄₂₀ | EC 1.5.98.1 |
| H ₂ -forming methylene-H ₄ MPT dehydrogenase | Methenyl-H ₄ MPT + H ₂ ↔ methylene-H ₄ MPT + H ⁺ | EC 1.12.98.2 |
| F ₄₂₀ -dependent methylene-H ₄ MPT reductase | Methylene-H ₄ MPT + reduced F ₄₂₀ ↔ CH ₃ -H ₄ MPT + oxidized F ₄₂₀ | EC 1.5.98.2 |
| Methyl-H ₄ MPT:coenzyme M methyltransferase | Coenzyme M + methyl-H ₄ MPT + 2 Na ⁺ /in ↔ 2-methyl-coenzyme M + 2 Na ⁺ /out + H ₄ MPT | EC 2.1.1.86 |
| Methyl-CoM reductase | CH ₃ -S-CoM + H-S-CoB ↔ CoM-S-S-CoB + CH ₄ | EC 2.8.4.1 |
| Heterodisulfide reductase | CoM-S-S-CoB + dihydromethanophenazine ↔ CoB + CoM + methanophenazine | EC 1.8.98.1 |
| Acetotrophic pathway | | |
| Acetate kinase-phosphotransacetylase system in <i>Methanosarcina</i> ; acetate thiokinase in <i>Methanosaeta</i> | Acetate + CoA ↔ acetyl-CoA + H ₂ O | EC 2.7.2.1 EC 2.3.1.8 EC 6.2.1.1 |
| CO-methylating acetyl-CoA synthase | Acetyl-CoA + a [Co(I) corrinoid Fe-S protein] ↔ CO + CoA + [methyl-Co(III) corrinoid Fe-S protein] | EC 2.3.1.169 |
| 5-Methyltetrahydrosarcinapterin: corrinoid/iron-sulfur protein Co-methyltransferase | [Methyl-Co(III) corrinoid Fe-S protein] + tetrahydrosarcinapterin ↔ a [Co(I) corrinoid Fe-S protein] + 5-methyltetrahydrosarcinapterin | EC 2.1.1.245 |
| Anaerobic carbon monoxide dehydrogenase | CO + H ₂ O + 2 oxidized Fd ↔ CO ₂ + 2 reduced Fd + 2 H ⁺ | EC 1.2.7.4 |
| Methyl H ₄ SPT: coenzyme M methyltransferase | CH ₃ H ₄ SPT + H-S-CoM ↔ CH ₃ -S-CoM + H ₄ SPT | EC 2.1.1.- |
| Methyl-CoM reductase | CH ₃ -S-CoM + H-S-CoB ↔ CoM-S-S-CoB + CH ₄ | EC 2.8.4.1 |
| Heterodisulfide reductase | CoM-S-S-CoB + dihydromethanophenazine ↔ CoB + CoM + methanophenazine | EC 1.8.98.1 |
| Methylotrophic pathway | | |
| Methanol:corrinoid protein Co-methyltransferase | Methanol + Co(I) corrinoid protein ↔ Methyl-Co(III) corrinoid protein + H ₂ O | EC 2.1.1.90 |
| [Methyl-Co(III) corrinoid protein]: coenzyme M methyltransferase | Coenzyme M + Methyl-Co(III) corrinoid protein ↔ 2-(methylthio)ethanesulfonate + Co(I) corrinoid protein | EC 2.1.1.246 |

| Enzyme | Reaction/process | EC number |
|--|---|--------------|
| Methylamine:corrinoic protein Co-methyltransferase | Methylamine + [Co(I) methylamine-specific corrinoic protein] ↔ a [methyl-Co(III) methylamine-specific corrinoic protein] + NH ₃ | EC 2.1.1.248 |
| Dimethylamine:corrinoic protein Co-methyltransferase | Dimethylamine + [Co(I) dimethylamine-specific corrinoic protein] ↔ a [methyl-Co(III) dimethylamine-specific corrinoic protein] + methylamine | EC 2.1.1.249 |
| Trimethylamine:corrinoic protein Co-methyltransferase | Trimethylamine + a [Co(I) trimethylamine-specific corrinoic protein] ↔ a [methyl-Co(III) trimethylamine-specific corrinoic protein] + dimethylamine | EC 2.1.1.249 |
| [Methyl-Co(III) methylamine-specific corrinoic protein]:coenzyme M methyltransferase | [Methyl-Co(III) methylamine-specific corrinoic protein] + CoM ↔ methyl-CoM + a [Co(I) methylamine-specific corrinoic protein] | EC 2.1.1.247 |
| Methyl-CoM reductase | CH ₃ -S-CoM + H-S-CoB ↔ CoM-S-S-CoB + CH ₄ | EC 2.8.4.1 |
| Heterodisulfide reductase | CoM-S-S-CoB + dihydromethanophenazine ↔ CoB + CoM + methanophenazine | EC 1.8.98.1 |

Table 4.
The selected enzymes of methanogenic step of anaerobic digestion [48, 49].

3. Conclusion

Biomass conversion to methane and carbon dioxide is the effect of complex interactions between microorganisms. These processes occur due to the microbial enzymatic machinery involved in specific metabolic pathways. Meta-omic analyses of microbial communities involved in AD reveal (i) dependence of microbial communities on the type of feedstock and operational conditions and (ii) describe interactions within microbial communities and ecophysiological functions of the specific taxa. Searching for the gene presence, gene expression, and protein expression, as well as linking structure and function of microbial communities, allows to develop a fundamental understanding of AD. This chapter is believed to contribute to the studies on the enzymatic road map of anaerobic digestion. However, it is only the tip of the iceberg of processes occurring in the microbial cells/microbial communities.

Acknowledgements

We acknowledge the support of The National Science Centre, Poland, through grant UMO-2015/17/B/NZ9/01718 and The National Centre for Research and Development, Poland, through grant BIOSTRATEG2/297310/13/NCBiR/2016.

Conflict of interest

The authors declare that there are no conflicts of interest.

IntechOpen

Author details

Anna Sikora^{1*}, Anna Detman¹, Damian Mielecki¹, Aleksandra Chojnacka¹
and Mieczysław Błaszczak²

¹ Institute of Biochemistry and Biophysics—Polish Academy of Sciences, Warsaw, Poland

² Faculty of Agriculture and Biology, Warsaw University of Life Sciences, Warsaw, Poland

*Address all correspondence to: annaw@ibb.waw.pl

IntechOpen

© 2018 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Sieber JR, McInerney MJ, Gunsalus RP. Genomic insights into syntrophy: The paradigm for anaerobic metabolic cooperation. *Annual Review Microbiology*. 2012;**66**:429-452. DOI: 10.1146/annurev-micro-090110-102844
- [2] Mao CL, Feng YZ, Wang XJ, Ren GX. Review on research achievements of biogas from anaerobic digestion. *Renewable and Sustainable Energy Reviews*. 2015;**45**:540-555. DOI: 10.1016/j.rser.2015.02.032
- [3] Sikora A, Detman A, Chojnacka A, Błaszczak MK. Anaerobic digestion: I. A common process ensuring energy flow and the circulation of matter in ecosystems. II. A tool for the production of gaseous biofuels. In: Jozala AF, editor. *Fermentation Processes*. Rijeka: InTech; 2017. pp. 271-301. DOI: 10.5772/64645
- [4] Kamagata Y. Syntrophy in anaerobic digestion. In: Fang HP, Zhang T, editors. *Anaerobic Biotechnology: Environmental Protection and Resource Recovery*. London: Imperial College Press, World Scientific; 2015. pp. 13-32. DOI: 10.1142/p1034/suppl_file/p1034_chap02
- [5] Stams AJM, Plugge CM. Electron transfer in syntrophic communities of anaerobic bacteria and archaea. *Nature Reviews*. 2009;**7**:568-577. DOI: 10.1038/nrmicro2166
- [6] Kleinstüber S. Metagenomics of methanogenic communities in anaerobic digesters. Biogenesis of hydrocarbons. In: Stams AJM, Sousa DZ, editors. *Biogenesis of Hydrocarbons, Handbook of Hydrocarbon and Lipid Microbiology*. Springer Nature Switzerland: Springer International Publishing AG; 2018. pp. 1-23. DOI: 10.1007/978-3-319-53114-4_16-1
- [7] Vanwonterghem I, Jensen PD, Ho DP, Batstone DJ, Tyson GW. Linking microbial community structure, interactions and function in anaerobic digesters using new molecular techniques. *Current Opinion Biotechnology*. 2014;**27**:55-64. DOI: 10.1016/j.copbio.2013.11.004
- [8] Koch C, Müller S, Harms H, Harnisch F. Microbiomes in bioenergy production: From analysis to management. *Current Opinion Biotechnology*. 2014;**27**:65-72. DOI: 10.1016/j.copbio.2013.11.006
- [9] Abram F. Systems-based approaches to unravel multi-species microbial community functioning. *Computational and Structural Biotechnology Journal*. 2015;**13**:24-32. DOI: 10.1016/j.csbj.2014.11.009
- [10] Cai M, Wilkins D, Chen J, Ng S-K, Lu H, Jia Y, et al. Metagenomic reconstruction of key anaerobic digestion pathways in municipal sludge and industrial wastewater biogas-producing systems. *Frontiers in Microbiology*. 2016;**7**:778. DOI: 10.3389/fmicb.2016.00778
- [11] Granada CE, Hasan C, Marder M, Konrad O, Vargas LK, Passaglia LMP, et al. Biogas from slaughterhouse wastewater anaerobic digestion is driven by the archaeal family *Methanobacteriaceae* and bacterial families *Porphyromonadaceae* and *Tissierellaceae*. *Renewable Energy*. 2018; **118**:840-846. DOI: 10.1016/j.renene.2017.11.077
- [12] Delforno TP, Lacerda GV Jr, Sierra-Garcia IN, Okada DY, Macedo TZ, Varesche MBA, et al. Metagenomic analysis of the microbiome in three different bioreactor configurations applied to commercial laundry wastewater treatment. *Science of the*

- Total Environment. 2017;**587-588**: 389-398. DOI: 10.1016/j.scitotenv.2017.02.170
- [13] Campanaro S, Treu L, Kougiaris PG, De Francisci D, Valle G, Angelidaki I. Metagenomic analysis and functional characterization of the biogas microbiome using high throughput shotgun sequencing and a novel binning strategy. *Biotechnology for Biofuels*. 2016;**9**:26. DOI: 10.1186/s13068-016-0441-1
- [14] Luo G, Fotidis IA, Angelidaki I. Comparative analysis of taxonomic, functional, and metabolic patterns of microbiomes from 14 full-scale biogas reactors by metagenomic sequencing and radioisotopic analysis. *Biotechnology for Biofuels*. 2016;**9**:51. DOI: 10.1186/s13068-016-0465-6
- [15] Guo J, Peng Y, Ni B-J, Han X, Fan L, Yuan Z. Dissecting microbial community structure and methane-producing pathways of a full-scale anaerobic reactor digesting activated sludge from wastewater treatment by metagenomics sequencing. *Microbial Cell Factories*. 2015;**14**:33. DOI: 10.1186/s12934-015-0218-4
- [16] Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: New perspectives on genomes, pathways, diseases and drugs. *Nucleic Acids Research*. 2017;**45**:D353-D361. DOI: 10.1093/nar/gkw1092
- [17] Kanehisa M, Sato Y, Kawashima M, Furumichi M, Tanabe M. KEGG as a reference resource for gene and protein annotation. *Nucleic Acids Research*. 2016;**44**:D457-D462. DOI: 10.1093/nar/gkv1070
- [18] Kanehisa M, KEGG GS. Kyoto Encyclopedia of genes and genomes. *Nucleic Acids Research*. 2000;**28**:27-30. DOI: 10.1093/nar/27.1.29
- [19] Caspi R, Billington R, Fulcher CA, Keseler IM, Kothari A, et al. The MetaCyc database of metabolic pathways and enzymes. *Nucleic Acids Research*. 2018;**46**(Database issue): D633-D639 <http://doi.org/10.1093/nar/gkx935>
- [20] BRENDA—The Comprehensive Enzyme Information System. Available from: <https://www.brenda-enzymes.org/>. [Accessed: July 24, 2018]
- [21] The Enzyme List Class 3—Hydrolases. Nomenclature Committee of the International Union of Biochemistry and Molecular Biology (NC-IUBMB) Generated from the ExplorEnz database; 2010
- [22] Shrestha S, Fonolla X, Khanal SK, Raskina L. Biological strategies for enhanced hydrolysis of lignocellulosic biomass during anaerobic digestion: Current status and future perspectives. *Bioresource Technology*. 2017;**245**: 1245-1257. DOI: 10.1016/j.biortech.2017.08.089
- [23] Berg JM, Tymoczko JL, Gatto GJ, Stryer L. *Biochemistry*. 8th ed. New York: W.H. Freeman & Company; 2015
- [24] Angenent LT, Karim K, Al-Dahhan MH, Wrenn BA, Domiguez-Espinosa R. Production of bioenergy and biochemicals from industrial and agricultural wastewater. *Trends in Biotechnology*. 2004;**22**: 477-485. DOI: 10.1016/j.tibtech.2004.07.001
- [25] Hallenbeck PC. Fundamentals of the fermentative production of hydrogen. *Water Science & Technology*. 2005;**52**: 21-29. PMID: 16180405
- [26] Kraemer JT, Bagley DM. Improving the yield from fermentative hydrogen production. *Biotechnology Letters*. 2007;**29**:685-695. DOI: 10.1007/s10529-006-9299-9

- [27] Lee D-J, Show K-Y, Su A. Dark fermentation on biohydrogen production: Pure cultures. *Bioresource Technology*. 2011;**102**(18):8393-8402
- [28] Sikora A, Błaszczuk M, Jurkowski M, Zielenkiewicz U. Lactic acid bacteria in hydrogen producing consortia: On purpose or by coincidence. In: Kongo M, editor. *Lactic Acid Bacteria—R & D for Food, Health and Livestock Purposes*. Rijeka, InTech; 2013. pp. 487-514. DOI: 5772/50364
- [29] Li F, Hinderberger J, Seedorf H, Zhang J, Buckel W, Thauer RK. Coupled ferredoxin and crotonyl coenzyme A (CoA) reduction with NADH catalyzed by the butyryl-CoA dehydrogenase/Etf complex from *Clostridium klyuveri*. *Journal of Bacteriology*. 2008;**190**: 843-850. DOI: 10.1128/JB.01417-07
- [30] Diekert G, Wohlfarth G. Metabolism of homoacetogens. *Antonie Van Leeuwenhoek*. 1994;**66**:209-221. DOI: 10.1007/BF00871640
- [31] Viana MB, Freitas AV, Leitão RC, Pinto GAS, Santaella ST. Anaerobic digestion of crude glycerol: A review. *Environmental Technology Reviews*. 2012;**1**:81-92. DOI: 10.1080/09593330.2012.692723
- [32] Ganzle MG. Lactic metabolism revisited: Metabolism of lactic acid bacteria in food fermentations and food spoilage. *Current Opinion in Food Science*. 2015;**2**:106-117. DOI: 10.1016/j.cofs.2015.03.001
- [33] Schink B, Stams AJM. Syntrophism among prokaryotes. In: Dworkin M, editor. *The Prokaryotes*. 3rd ed. New York: Springer; 2006. pp. 309-335. DOI: 10.1007/978-3-642-30123-0_59
- [34] Nisman B. The Stickland reaction. *Bacteriology Reviews*. 1954;**18**:16-42
- [35] Bouillaut L, Self WT, Sonensheina AL. Proline-dependent regulation of *Clostridium difficile* Stickland metabolism. *Journal of Bacteriology*. 2013;**195**:844-854. DOI: 10.1128/JB.01492-12
- [36] Fonknechten N, Chaussonnerie S, Tricot S, Lajus A, Andreesen JR, et al. *Clostridium sticklandii*, a specialist in amino acid degradation: Revisiting its metabolism through its genome sequence. *BMC Genomics*. 2010;**11**:555. DOI: 10.1186/1471-2164-11-555
- [37] Buckel W. Unusual enzymes involved in five pathways of glutamate fermentation. *Applied Microbiology and Biotechnology*. 2001;**57**:263-273. DOI: 10.1007/s002530100773
- [38] Hattori S. Syntrophic acetate-oxidizing microbes in methanogenic environments. *Microbes and Environments*. 2008;**23**:118-127. DOI: 10.1264/jsme2.23.118
- [39] Galushko AS, Schink B. Oxidation of acetate through reactions of the citric acid cycle by *Geobacter sulfurreducens* in pure culture and in syntrophic coculture. *Archives of Microbiology*. 2000;**174**:314-321. DOI: 10.1007/s002030000208
- [40] Schmidt A, Müller N, Schink B, Schleheck D. A proteomic view at the biochemistry of syntrophic butyrate oxidation in *Syntrophomonas wolfei*. *PLoS One*. 2013;**8**(2):e56905. DOI: 10.1371/journal.pone.0056905
- [41] Muller N, Worm P, Schink B, Stams AJM, Plugge CM. Syntrophic butyrate and propionate oxidation processes: From genomes to reaction mechanisms. *Environmental Microbiology Reports*. 2010;**2**:489-499. DOI: 10.1111/j.1758-2229.2010.00147.x
- [42] Worm P, Koehorst JJ, Visser M, Sedano-Núñez VT, Schaap PJ, et al. A genomic view on syntrophic versus non-syntrophic lifestyle in anaerobic fatty

acid degrading communities.
Biochimica et Biophysica Acta
Bioenergetics. 2014;**1837**:2004-2016.
DOI: 10.1016/j.bbabi.2014.06.005

Nature Reviews Microbiology. 2008;**6**:
579-591. DOI: 10.1038/nrmicro1931

[43] Walker CB, He Z, Yang ZK,
Ringbauer JA, He Q, et al. The electron
transfer system of syntrophically grown
Desulfovibrio vulgaris. Journal of
Bacteriology. 2009;**191**:5793-5801. DOI:
10.1128/JB.00356-09

[44] Meyer B, Kuehl J,
Deutschbauer AM, Price MN, Arkin AP,
et al. Variation among *Desulfovibrio*
species in electron transfer systems used
for syntrophic growth. Journal of
Bacteriology. 2013;**195**(5):990-1004.
DOI: 10.1128/JB.01959-12

[45] Schmidt A, Frensch M, Schleheck D,
Schink B, Muller N. Degradation of
acetaldehyde and its precursors by
Pelobacter carbinolicus and *P.*
acetylenicus. PLoS One. 2014;**9**(9, 12):
e115902. DOI: 10.1371/journal.
pone.011590

[46] Bertsch J, Siemund AL, Kremp F,
Muller V. A novel route for ethanol
oxidation in the acetogenic bacterium
Acetobacterium woodii: The
acetaldehyde/ethanol dehydrogenase
pathway. Environmental Microbiology.
2016;**18**:2913-2922. DOI: 10.1111/
1462-2920.13082

[47] Weghoff MC, Bertsch J, Muller V. A
novel mode of lactate metabolism in
strictly anaerobic bacteria.
Environmental Microbiology. 2015;**17**:
670-6777. DOI: 10.1111/1462-2920.12493

[48] Thauer RK. Biochemistry of
methanogenesis: A tribute to Marjory
Stephenson. Microbiology. 1998;**144**:
2377-2406. DOI: 10.1099/00221287-144-
9-2377

[49] Thauer RK, Kaster AK, Seedorf H,
Buckel W, Hedderich R. Methanogenic
Archaea: Ecologically relevant
differences in energy conservation.