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Chemistry and Biodiversity of *Rhizophora*-Derived Endophytic Fungi

Jing Zhou and Jing Xu

Abstract

*Rhizophora* are salt-tolerant mangrove flora located in tropical and subtropical intertidal coastal regions. This review summarizes frequently occurring fungal endophytes in *Rhizophora*. In total, 41 families and 64 genera belonging to 23 taxonomic orders of Ascomycota have been reported. Among those discussed here, *Pestalotiopsis*, *Penicillium*, and *Mucor* are the most abundant fungal genera, and they are widely studied. In previous studies, 195 metabolites were encountered in *Rhizophora*-derived endophytic fungi, and their structures are reported within a biogenetic context. Bioassays showed antitumor, antimicrobial, as well as anti-H1N1 activities to be the most notable bioactivities of the secondary metabolites discussed.

Keywords: *Rhizophora*-derived endophytic fungi, biodiversity, secondary metabolites, biological activities

1. Introduction

Endophytic fungi, a polyphyletic group of highly diverse, primarily ascomycetous fungi that spend all or at least for a part of their life cycle inter- or intracellularly colonizing healthy tissues of plants without causing visible disease symptoms [1]. They are found in almost all vascular plants and grass plants [2]. It is worth noting that of the nearly 300,000 plant species that exist on Earth, any given plant is colonized by several to few hundreds of endophytic fungal species. Only a few of these plants have ever been completely studied relative to their endophytic biology [3]. Until recently, extensive work has been conducted on traditionally investigated terrestrial endophytic fungi with biological significance, and these studies mostly concentrated on the tropical and rainforest regions of the world. However, systematic
and comparative approaches to identifying endophytic fungi and their specific location in the plants they colonize, especially in ecological niches such as mangrove endosymbionts growing in high salinity, high temperature, extreme tides, oxygen pressure, high humidity, and light and air limitations, have received considerable attention in recent decades [4, 5]. Hence, it is now generally accepted that the highly complex mangrove ecosystems could act as an effective selector for metabolic pathway evolution via the generation of structurally unprecedented and biologically interesting metabolites of pharmaceutical importance. Such metabolites are believed to be involved in ecological adaptability, defense, communication, and predation [6]. In this review, we summarize the biodiversity of *Rhizophora* endophytic fungi. Additionally, the metabolites encountered in *Rhizophora*-derived endophytic fungi and their structures are reported within a biogenetic context. Special emphasis is placed on the prospect of discovering unique functional metabolites.

2. Endophytic fungi from *Rhizophora*

Mangroves are composed of a large group of salt-tolerant plant communities growing in tropical and subtropical intertidal estuarine zones, which are distributed approximately in the area between 30° N and 30° S latitude [7]. Asia and Australia have the greatest diversity and distribution of mangrove species. Among the 18 million hectares of mangrove forests, more than 40% are found along the Asian coasts, including the South China Sea Coast [10]. The most established mangroves can be found in Bangladesh, Brazil, Indonesia, India, and Thailand [8, 9]. According to the statistical data of the International Society of Mangrove Ecosystem, there are 84 mangrove species globally, belonging to 16 families and 24 genera. Among them, 70 species are true mangroves, pertaining to 16 genera and 11 families. Another 14 species are considered semimangroves, belonging to 8 genera and 5 families [10]. China has 26 species, and 24 of them are distributed in Hainan [11, 12].

*Rhizophora* is one of the most conspicuous genera of the most widespread mangrove family, the Rhizophoraceae. The genus is relatively old among cosmopolitan mangrove genera, and it has notable discontinued species distributions [13]. In total, eight species comprise the *Rhizophora*, including *R. stylosa*, *R. apiculata*, *R. mucronata*, *R. mangle*, *R. harrisonii*, *R. racemosa*, *R. annamalayana*, and *R. samoensis* (Table 1). *R. stylosa*, *R. mucronata*, and *R. apiculata* are mainly distributed in islands and coastal areas bordering the Pacific Ocean and the Indian Ocean, while *R. mangle*, *R. annamalayana*, *R. samoensis*, *R. harrisonii*, and *R. racemosa* are mainly distributed from the eastern Pacific through the American islands to the Atlantic Ocean (Figure 1).

Fungi colonized in mangrove forests, which comprise the second largest ecological group of the marine fungi, have specially adapted their own morphological structures and physiological mechanisms to promote the survival of host plants in harsh environmental conditions through long-term endophyte-host interactions [52]. Most mangrove endophytic fungi are facultative halophiles and euryhaline in nature. Since they do not require added salt for growth, they are able to grow at high salt concentrations and show a balanced symbiotic continuum of mutualism with host mangroves [5]. For instance, the halotolerant *Rhizophora stylosa*
endophytic Pestalotiopsis sp. is isolated and capable of producing lignin-degrading enzymes. This species secretes over 400 salt-adapted lignocellulolytic enzymes, which enhance the salt adaptation of mangrove hosts [18].

To date, the species of mangrove endophytic fungi identified from a large and diverse ecological group are mostly members of the Ascomycota phylum, with a limited occurrence of basidiomycetes [53, 54]. Since 1955, when Cribb first described endophytic fungi isolated from mangrove roots, several studies on the fungi residing in mangroves along the coastlines of the Indian, Pacific, and Atlantic Oceans have been conducted [55]. Hyde [56] listed approximately 120 fungal species that colonize 29 mangrove plants globally, including 87 ascomycetes, 31 mitosporic fungi, and 2 basidiomycetes. Schmit and Shearer [57, 58] reported 625 mangrove-associated fungi, including 279 ascomycetes, 277 mitosporic fungi, 29 basidiomycetes, 3 chytridiomycetes, 2 myxomycetes, 14 oomycetes, 9 thraustochytrids, and 12 zygomycetes. According to the frequency of their appearance, Alternaria, Aspergillus, Cladosporium, Colletotrichum, Fusarium, Paecilomyces, Penicillium, Pestalotiopsis, Phoma, Phomopsis, Phyllosticta, and Trichoderma have been recognized as the predominant culturable mangrove endophytic fungi [59].

<table>
<thead>
<tr>
<th>Plants species</th>
<th>Distribution</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>R. stylosa</em></td>
<td>China (Hainan, Guangdong, Guangxi); Philippines; New Caledonia; Fiji (Viti Levu); Australia; Japan (Ryukyu Archipelago)</td>
<td>Hainan plant flora [12]; Xing [14]; Villamayor [15]; Dangan [16]; Morton [17]; Arli [18]; Chen [11]; Tyagi [19]; Kohlmeyer [20]</td>
</tr>
<tr>
<td><em>R. apiculata</em></td>
<td>China (Hainan, Guangdong, Guangxi); India; Indonesia; Philippines; Vietnam; Thailand; Singapore; Malaysia</td>
<td>Hainan plant flora [12]; Xing [14]; Selvaraj [21]; Villamayor [15]; Dangan [16]; Rossiana [22]; Clough [23]; Piapukiew [24]; Klaiklay, [25]; Rukachaisirikul [26]; Tan [27]</td>
</tr>
<tr>
<td><em>R. mucronata</em></td>
<td>China (Taiwan); Vietnam; South Africa; Philippines; Indonesia; India; Thailand; Japan; Singapore; Pakistan</td>
<td>Hainan plant flora [12]; Trinh [28]; Osorio [29]; Villamayor [15]; Dangan [16]; Tarman [30]; Suryanarayanan [31]; Rani [32]; Kandasamy [33]; Rukachaisirikul [26]; Tan [27]; Tariq [34]</td>
</tr>
<tr>
<td><em>R. mangle</em></td>
<td>Brazil; Venezuela; Dominican Republic; Guad de Ropp; Mexico; America (Florida, Hawaii); Senegal; Gabon; French Guiana; Australia</td>
<td>Boehm [35]; Ferreira [36]; Barreto [37]; Ball [38]; Afzal [39]; Wanderley [40]; Dourado [41]; Godoy [42]; Kohlmeyer [20]</td>
</tr>
<tr>
<td><em>R. harrisonii</em></td>
<td>Nigeria (Port Harcourt); Ecuador; America; West Africa; Equatorial Guinea; Senegal; Gabon</td>
<td>Hemphill [43]; Twilley [44]; Breteler [45]; Cerónsouza [46]; Cornejo [47]; Afzal [39]</td>
</tr>
<tr>
<td><em>R. racemosa</em></td>
<td>Nigeria; Ecuador; French Guiana; Gambia; Senegal; Gabon; Togo; America (Hawaii); Mexico</td>
<td>Ukoima [48]; Xavier [49]; Afzal [39]; Osorio [29]</td>
</tr>
<tr>
<td><em>R. annamalayana</em></td>
<td>India</td>
<td>Elavarasi [50]</td>
</tr>
<tr>
<td><em>R. samoensis</em></td>
<td>Fiji (Viti Levu); America; Southwest Pacific Islands (Caledonia, Hebrides); Samoa; Marshall Islands</td>
<td>Tyagi [19]; Duke [51]</td>
</tr>
</tbody>
</table>

Table 1. The distribution of *Rhizophora* in the world.
As a relatively underappreciated reservoir of bioresources, endophytic fungi from mangroves have been considered potential pharmaceutical and agricultural resources. Recent studies have investigated the biodiversity and distribution of mangrove endophytic fungi in the South China Sea. The taxonomic identities and diversity of endophytic fungal communities isolated from five species of the genus *Sonneratia* (*S. caseolaris*, *S. hainanensis*, *S. ovata*, *S. paracaseolaris*, and *S. apetala*) and four species of Rhizophoraceae (*Ceriops tagal*, *R. apiculata*, *R. stylosa*, and *Bruguiera sexangula* var. *rhynchopetala*) have been addressed [14].

Identification of biologically interesting metabolites from these endophytic fungi is an important initial step in understanding the role of endophytes to host mangrove plants. According
<table>
<thead>
<tr>
<th>Plants species</th>
<th>Isolated endophytic fungi</th>
<th>Sampling location</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>R. stylosa</em></td>
<td>Aureobasidium, Aspergillus, Cladosporium, Diaporthe, Fusarium, Guignardia, Pestalotiopsis</td>
<td>China</td>
<td>Xing [14]</td>
</tr>
<tr>
<td></td>
<td>Acremonium, Alternaria, Aspergillus, Bionectria, Colletotrichum, Epicoccum, Nigrospora, Penicillium, Pestalotiopsis, Phoma, Phomopsis, Phialophora, Talaromyces, Trichoderma</td>
<td>Hyde [60]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Chaetomium, Corynespora, Fusarium, Geniculosporium, Glomerella, Guignardia, Melanzonium, Sphaecoma, Pestalotiopsis, Phoma</td>
<td>Liu [59]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Penicillium</td>
<td>Hainan</td>
<td>Gao [62], Zhang [63], Sun [64]</td>
</tr>
<tr>
<td><em>R. apiculata</em></td>
<td>Aspergillus, Aureobasidium, Cladosporium, Diaporthe, Fusarium, Massarina, Penicillium, Pestalotiopsis, Phomopsis</td>
<td>Hong Kong</td>
<td>Xing [14]</td>
</tr>
<tr>
<td></td>
<td>Acremonium, Flavodon, Phomopsis, Pestalotiopsis</td>
<td>Thailand</td>
<td>Klaiklay [25], Klaiklay [65, 66], Buatong [67], Rukachaisirikul [26]</td>
</tr>
<tr>
<td></td>
<td>Acremonium, Alternaria, Aureobasidium, Cladosporium, Curcularia, Drechslera, Fusarium, Nodulisporium, Pestalotiopsis, Phialophora, Phoma, Phomopsis, Phyllosticta, Pithomyces, Glomerella, Sporothrix, Sporormiella, Xylariaceae</td>
<td>India</td>
<td>Kumaresan [68]</td>
</tr>
<tr>
<td></td>
<td>Acremonium, Alternaria, Cladosporium, Chaetomium, Penicillium, Pestalotiopsis, Phialophora, Phoma, Phyllosticta, PseudEurotium, Sporormiella, Titulavia</td>
<td>Suryanarayanan [31]</td>
<td></td>
</tr>
<tr>
<td><em>R. mucronata</em></td>
<td>Pestalotiopsis</td>
<td>Dong Zhai Gang</td>
<td>Xu [69]</td>
</tr>
<tr>
<td></td>
<td>Aspergillus</td>
<td>Indonesia</td>
<td>Tarman [30]</td>
</tr>
<tr>
<td></td>
<td>Phomopsis</td>
<td>Shiono</td>
<td>[70]</td>
</tr>
<tr>
<td></td>
<td>Diaporthe, Neofusicoccum</td>
<td>South Africa</td>
<td>Osorio [29]</td>
</tr>
<tr>
<td></td>
<td>Acremonium, Alternaria, Aspergillus, Betyptrichum, Cladosporium, Chaetomium, Glomerella, Nigrospora, Pestalotiopsis, Phialophora, Phomopsis, Phyllosticta, Sporormiella, Trichoderma</td>
<td>India</td>
<td>Suryanarayanan [31]</td>
</tr>
<tr>
<td></td>
<td>Ascotricha, Aspergillus, Cirrenulia, Cladosporium, Dicyma, Fusariella, Pseudomycetes, Penicillium, Phoma, Phomopsis, Trichodadium, Zalerion, Zygosporium</td>
<td>Ananda [71]</td>
<td></td>
</tr>
<tr>
<td><em>R. mangle</em></td>
<td>Glomerella, Guignardia, Nodulisporium, Phyllosticta</td>
<td>Brazil</td>
<td>Wanderley [40]</td>
</tr>
<tr>
<td></td>
<td>Leucostoma</td>
<td>Beu [72]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Betgrosphaeria, Colletotrichum, Coprinellus, Cytopora, Diaporthe, Endothia, Epicoccum, Fusarium, Gibberella, Glomerella, Guignardia, Hypocrea, Leptosphaeria, Neofusicoccum, Penicillium, Phomopsis, Pichia, Trichoderma, Xylaria, Valsa</td>
<td>Sebastians [73]</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Cytospora</td>
<td>Wier [74]</td>
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</tr>
</tbody>
</table>
to the previous studies, the identification and phylogenetic diversity of mangrove endophytic fungi was largely associated with mangroves located in China, Thailand, Indonesia, Brazil, and India. In total, 26 genera of mangrove endophytic fungi were isolated from *R. stylosa*; 27 genera were isolated from *R. apiculata*; 26 genera were obtained from *R. mucronata*; 23 genera were isolated from *R. mangle*; 1 genus was isolated from *R. harrisonii* and *R. annamalayana* (namely *Pestalotiopsis* and *Fusarium*); and 4 genera of endophytic fungi were isolated from *R. racemosa*. Until now, no studies have been conducted on *R. samoensis*. In comparison with the previous reports, the frequently occurring fungi entophytes in *Rhizophora*, including 41 families and 64 genera belonging to 23 taxonomic orders of Ascomycota have been reported. The fungi of Basidiomycota are rarely found in *Rhizophora*. The dominant endophytic fungi of the *Rhizophora* genus are mainly distributed in *Aspergillus*, *Cladosporium*, *Chaetomium*, *Fusarium*, *Lasiodiplodia*, *Penicillium*, *Pestalotiopsis*, *Phomopsis*, *Phoma*, *Phyllosticta*, and *Trichoderma* (Table 2).

### Table 2. The endophytic fungi isolated from *Rhizophora*.

<table>
<thead>
<tr>
<th>Plants species</th>
<th>Isolated endophytic fungi</th>
<th>Sampling location</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>R. harrisonii</em></td>
<td>Pestalotiopsis</td>
<td>Nigeria</td>
<td>Hemphill [43]</td>
</tr>
<tr>
<td><em>R. racemosa</em></td>
<td>Aspergillus, Lasiodiplodia, Paecilomyces, Penicillium</td>
<td>Nigeria</td>
<td>Ukoima [48]</td>
</tr>
<tr>
<td><em>R. annamalayana</em></td>
<td>Fusarium</td>
<td>Vellar estuary</td>
<td>Elavarasi [50]</td>
</tr>
<tr>
<td><em>R. samoensis</em></td>
<td></td>
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</tr>
</tbody>
</table>

3. The secondary metabolites of endophytic fungi of *Rhizophora*

There is a wide range of endophytic fungi in mangroves, and their growing environment is unique. Thus, in the formation of special fungal communities, they will certainly metabolize compounds with rich structures, unlike that of terrestrial fungi. Many of these metabolites provide a rich model structure for the screening of new drugs, which have become increasingly valuable in drug-lead research [5]. A total of 195 metabolites were discovered from *Rhizophora*-derived endophytic fungi reported so far are included. The secondary metabolites of endophytic fungi of mangrove are classified as alkaloids, terpenes, coumarins, chromones, quinones, anthraquinones, peptides, phenolic acids, lactones, and other compounds.

#### 3.1. Alkaloids

*Fusarium equisetin* AGR12 from *R. stylosa* produced two cyclic acetyl phytotoxin derivatives, equisetin (1) and *epi-equisetin* (2) [75, 76]. Both equisetin (1) and *epi-equisetin* (2) exhibit modest antibacterial activity, and equisetin (1) had selective antimicrobial activity against some Gram-positive bacteria [77]. The metabolite equisetin was first purified from maize grit medium cultures of *F. equiseti* strain NRRL 5337, and equisetin can inhibit the ATPase activity of mitochondria in rat hepatocytes induced by 2,4-dinitrophenol (DNP) in a concentration-dependent manner.
At a concentration of 8 nmol equisetin/mg protein, the inhibition rate can reach 50% [78]. New cerebroside lipids, chrysogesides A–E (3–8), and new pyridone ketones, chrysogedones A and B (9, 10), were isolated from the fermentation extract of *Penicillium chrysogenum* PXP-55, isolated from *R. stylosa*. Compound (6) exhibited inhibitory activity against *Enterobacter aerogenes* with MIC value of 1.72 μM [61]. The fungus species *Pestalotiopsis* JCM2A4, isolated from the Chinese mangrove plant *Rhizophora mucronata*, is one of the most abundant resources for screening natural products with different biological activities [79]. New N-substituted amide derivatives, pestalotiopamides A–E (12–16), and a new succinimide, pestalotiopsoid A (11), were isolated from the fermented crude extracts of *Pestalotiopsis* sp. JCM2A4, which was collected from *R. mucronata* [69, 80, 81]. A culture of the fungus *Aspergillus nidulans* MA-143, isolated from *R. stylosa* leaves, yielded six new compounds, and all the compounds contained the structural unit 4-phenyl-3,4-dihydroquinolin-2(1H)-one, aniduquinolones A–C (17–19), 6-deoxyaflaquinolone E (20), isoaflaquinolone E (21), 14-hydroxyaflaquinolone F (22), and aflaquinolone A (23). The bioactivity results showed that compounds 17–23 had no inhibitory activity against human hepatocellular carcinoma BEL-7402, breast cancer cell MDA-MB-231, leukemia myeloid cell HL-60, or chronic myeloid leukemia cell K562. Additionally, these compounds had no antibacterial activity against *Staphylococcus aureus* or *Escherichia coli*. Compounds 17, 19, and 23 exhibited lethal activity against *Artemia salina*, with LD₅₀ values of 7.1, 4.5, and 5.5 μM, respectively [82]. About 6 new indole diterpenoid alkaloid derivatives (24–29) and 5 known similar metabolites, including 21-isopentenyloxilpine (30), pxiline (31), chydroxypxiline (32), emindole (33), and paspaline (34), were identified from a culture of *Penicillium camemberti* OUCMDZ-1492, isolated from the *R. apiculata*. Among them, compounds 24, 26–28, and 30–33 all showed strong H1N1 influenza virus inhibitory activity, with IC₅₀ values ranging from 6 to 80 μM [83]. A new paspaline (34) and three known analogs, penijanthine A (35), paspalinine (36), and pentrem (37), were isolated from *Alternaria tenuissima* EN-192 from *R. stylosa* stems. Compounds 34–37 had slight antimicrobial activity against *Staphylococcus aureus*, *Escherichia coli*, *Bacillus subtilis*, and *Vibrio anguillarum* [64]. The cultivable *Phomopsis* sp. PSU-MA214 from *R. apiculata* leaves can produce phenylethanol compounds, including phomonitroester (38). Compound 38 was initially isolated from *Phomopsis* sp. PSU-D15, which was from another plant of *Garcinia dulcis* [84]. The bioassay test showed that compound 38 had a weak inhibitory effect on breast cancer cells MCF-7 and KB85. The four new quinazolone alkaloid derivatives, aniquinazolines A–D (39–42) which were isolated from *Aspergillus nidulans* MA-143 in *R. stylosa*, showed strong lethal activity in shrimp, with LD₅₀ values of 1.27, 2.11, 4.95, and 3.42 μM, respectively. Meanwhile, they had no inhibitory activity against hepatoma cell BEL-7402, breast cancer cell MDA-MB-231, leukemia myeloid cell HL-60, and chronic myeloid leukemia cell K562. Moreover, no antibacterial activity against *Staphylococcus aureus* and *Escherichia coli* was observed [82]. Two new indole alkaloids, penoxamide A (43) and 18-hydroxydecaturin B (44), and a known compound decaturin B (45) were isolated from the fermented rice extract of *R. stylosa* endophytic fungi *Penicillium oxalidum* EN-201 [85]. Mucor irregularis QEN-189 was isolated from *R. stylosa*, from which 6 indole diterpenoid alkaloid derivatives and 14 analogs were separated, namely rhizovarins A–F (46–50, 53), secopentrem D (51), PC-M4 (52), penijanthine A (54), penitrem A–F (55–60), pxiline (61), 27-O-acetylpxiline (62), 13-deoxy-27-O-acetylpxiline (63), 10-deoxy-13-deoxypxiline
As for antitumor activity, compounds 46, 47, 50, 55, 60, and 65 had inhibitory activity against lung cancer cell A549, and the IC₅₀ values were 11.5, 6.3, 8.4, 8.0, 8.2, and 4.6 μM, respectively. They also had inhibitory activity against leukemia cells of HL-60, with IC₅₀ values of 9.6, 5.0, 7.0, 4.7, 3.3, and 2.6 μM, respectively [62]. The Hypocrea virens of R. apiculata is capable of producing isoquinoline alkaloids, 2-methylimidazo[1,5-b]isoquinoline-1,3,5(2H)-trione (66) [86] (Figure 2).

3.2. Terpenoids

A new sesquiterpene, diaporol A (67), with a tricyclic lactone structure; eight new sesquiterpenes, diaporols B–I (68–75); drimane; 3β-hydroxyconfertifolin (76); and diplodiatoxin (77) were isolated from Diaporthe sp. of R. stylosa. The bioactivity test showed that compounds 67–77 had no cytotoxicity on human gastric cancer cell SGC-7901, breast cancer cell MCF-7, lung cancer cell A549, and hepatocellular carcinoma cell line QGY-7701 at a concentration of 20 μM [63]. Flavodon flavus PSU-MA201 was isolated from R. apiculata, from which a known perhydroazulene compound, tremulenolide A (78), was separated, and the bioassay test showed that compound 78 exhibited modest antibacterial activity against Staphylococcus aureus ATCC25923 and Cryptococcus neoformans ATCC90113 with MIC values of 128 μg/ml [65, 66]. A known altitoxin B (79) with drimane was isolated from Pestalotiopsis sp. of R. macronata [87]. Two known mycotoxins, 8-deoxytrichothecin (80) and trichodermol (81), were isolated from the Acromonium sp. PSU-MA70 of R. apiculata [26]. As a plant-derived anticancer drug with a unique mechanism, taxol (82) was isolated from Taxus brevifolia bark and wood for the first time by American chemists Wani and Wall in 1963 [88, 89]. Subsequently, it has been found that endophytic fungi Taxomyces [90], Pestalotiopsis [91], Alternaria [92], and Fusarium [93] could also produce taxol and its analogs. Taxol (82) was also isolated from endophytic fungus Fusarium oxysporum in R. annamalayana [50]. Two new compounds, pestalotiopeps A and B (83, 84), were separated from the Pestalotiopsis sp. JCM2A4 from leaves of R. macronata, and the bioactivity assay revealed that compound 83 was slightly

Figure 2. The structures of alkaloids in Rhizophora-derived endophytic fungi.
resistant to Staphylococcus aureus, Escherichia coli, Enterococcus faecalis, Streptococcus pyogenes, Pseudomonas aeruginosa, and Klebsiella pneumoniae, with the MIC values ranging from 125 to 250 μM [87] (Figure 3).

3.3. Coumarins

A strain of Pestalotiopsis sp. was isolated from the leaves of R. mucronata, which is an important resource of coumarin compounds. Pestalasins A–E (85–89) and one known compound, 3-dydroxymethyl-6,8-dimethoxycoumarin (90), were separated from fermentation extracts, and this was the first time that coumarin had been found in the mangrove microbes [69]. A more in-depth study of the chemical constituents of Pestalotiopsis sp. led to the discovery of a new isocoumarin derivative, pestalotiopisorin A (91) [80]. Seven new structural analogs, acremonones B–H (92–98), were isolated from Acremonium sp. PSU-MA70, which was from R. apiculata [26]. Pestalotiopsis clavispora was isolated from the leaves of R. harrisonii, and four new polyketide derivatives were separated from endophytic fungi, including pestapyrones A–C (99–101), (R)-periplanetin D (103), and similarpyrone B (102) [43] (Figure 4).

3.4. Chromones

Three rare chlorinated chromone derivatives, pestalochromones A–C (104–106), were isolated from Pestalotiopsis sp. PSU-MA69 in R. apiculata [25]. Further studies on the chemical composition of Pestalotiopsis sp. from R. mucronata led to the discovery of a series of rare lipophilic chromone derivatives, pestalotiopsones A–F (107–112), and the known compound, 5-carbomethoxymethyl-heptyl-7-hydroxychromone (113). The bioactivity test showed that compound 111 had weak cytotoxic activity against mouse lymphoma cell L5178Y, with an EC$_{50}$ value of 29.4 μM [69]. Four new chromone derivatives, phomopsichins A–D (114–117), along with a known compound, phomoxanthone A (118), were isolated from the fermentation products of Phomopsis sp. 33 from R. stylosa. The bioassay results showed that compounds

Figure 3. The structures of terpenoids in Rhizophora-derived endophytic fungi.

Figure 4. The structures of coumarins in Rhizophora-derived endophytic fungi.
114–118 had weak inhibitory effects on acetylcholinesterase (AchE), α-glucanase, DPPH radical and hydroxyl radical, as well as weak inhibitory activity against 18 kinds of plant pathogenic bacteria [94]. A new polyketone derivative, pestalpolyol I (119), was isolated from Pestalotiopsis clavispora in *R. harrisonii*. The bioactivity test showed that compound 119 had strong inhibitory activity against tumor cells LS17Y, with an IC_{50} value of 4.1 μM. Compound 119 also showed inhibitory activity against leukemia myeloid cells HL-60, hepatoma cells SMMC-7721, lung cancer cells A-549, breast cancer cells MCF-7, and human colon cancer cells SW480, with IC_{50} values of 10.4, 11.3, 2.3, 13.7 and 12.4 μM, respectively [43] (Figure 5).

3.5. Anthraquinones

One new tetrahydroanthraquinone derivative, (2R, 3S)-7-ethyl-1,2,3,4-tetrahydro-2,3,8-trihydroxy-6-methoxy-3-methyl-9,10-anthracenenedione (120) and five known anthraquinones derivatives (121–125) were isolated from the endophytic fungi *Phomopsis* sp. PSU-MA214 from *R. apiculata* leaves. Compound 120 had the structure of ethyl tetrahydroanthraquinone, which was weakly cytotoxic to human breast cancer cell MCF-7 and had antibacterial activity against *Staphylococcus aureus* ATCC25923 and meticillin-resistant *S. aureus* SK1 [25]. Three known tricyclic alternarene derivatives (126–128) were isolated from the endophytic fungus *Alternaria tenuissima* EN-192 from *R. stylosa* branches, and the antimicrobial activity, tested by filter paper diffusion method, showed that compound 126 had moderate antibacterial activity against *Vibrio anguillarum* [64]. One new xanthone, pestaloxanthone (129), was isolated with two known analogs, isosulochrin dehydrate (130) and chloroisosulochrin dehydrate (131), from endophytic fungi Pestalotiopsis sp. PUS-MA69 from *R. apiculata* branches [25]. A known tetrahydrogenated xanthone dimer, phomoxanthone A (132), and a new compound with similar structure, 12-O-deacetyl-phomoxanthone A (133), were isolated from a rice fermentation extract of the fungus *Phomopsis* sp. IM 41-1 from *R. mucronata*. Two compounds (132, 133) had weak antibacterial activity against *Botrytis cinerea*, *Sclerotinia aureus*, *Diaporthe medusaea*, and *Staphylococcus aureus*, while acetylation of the compound had no significant effect on the antimicrobial activity [70]. A known compound, pestaxanthone (134), was isolated from *Pestalotiopsis clavispora* from the leaves of the genus *R. harrisonii* [43] (Figure 6).

3.6. Peptides

Four known compounds, two ring-phthalocyanines, guangomides A and B (135, 136), and two diketopiperazine derivatives, Sch 54794 and Sch 54796 (137, 138), were isolated from the

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**Figure 5.** The structures of chromones in *Rhizophora*-derived endophytic fungi.
Acremonium sp. PSU-MA70 from R. apiculata [26]. Activity tests showed that compounds 135 and 136 had weak antibacterial activity against Staphylococcus epidermidis and Enterococcus durans [95] (Figure 7).

3.7. Phenolics

In this category, four new diphenyl ether compounds, pestalotethers A–D (141, 143–145), and three known compounds, pestheic acid (142), chloroisosulochrin (139), and isosulochrin (140), were isolated from Pestalotiopsis sp. PSU-MA69 of R. apiculata [25]. A new compound, norpe-staphthalide A (146), and three known compounds, (R, S)-5,7-dihydroxy-3-(1-hydroxyethyl) phthalide (148) and pestaphthalides A and B (147, 149), were isolated from Pestalotiopsis clavispora in the leaves of R. harrisonii. These compounds had no inhibitory effect on leukemia myeloid cells HL-60, hepatoma cells SMMC-7721, lung cancer cells A-549, breast cancer cells MCF-7, and human colon cancer cells SW480 [43] (Figure 8).

3.8. Lactones

Five new compounds, including cytosporones J–N (152–156), together with known metabolites, dothiorelones A (150) and cytosporones C (151), were isolated from the Pestalotiopsis sp. from R. mucronata. Biological tests showed that compound 150 was cytotoxic to human oral epidermoid carcinoma KB cells, lymphoma cells Raji, and human osteosarcoma cells Mg-63. Compounds 151–156 had no significant antitumor activity [69]. In the further study of Pestalotiopsis sp. of R. mucronata, eight new pyrone compounds, pestalotiopyrones A–H

Figure 6. The structures of anthraquinones in Rhizophora-derived endophytic fungi.

Figure 7. The structures of peptides in Rhizophora-derived endophytic fungi.

Figure 8. The structures of phenolics in Rhizophora-derived endophytic fungi.
(157–164); two new compounds, pestalotiollides A and B (166, 167); and one known compound, nigrosporapyrone D (165), were found in large amounts of fermentation products in the rice culture medium [80]. Three new α-pyrene pestalotiopyrones A–C (168–170); two new seiricuprolide macrolides, pestalotiopyrones A (171) and B (173); and two known compounds, seiricuprolide (174) and 2′-hydroxy-3′,4′-didehydroenepicolide (172), were isolated from two endophytic fungi Pestalotiopsis sp. PSU-MA92 and Pestalotiopsis sp. PSU-MA119 of R. apiculata and R. mucronata [96]. Among these, compounds 168–170 were repetitive names of pestalotiopyrones A–C [80]. Thus far, the carbon skeleton of phenyleol lactones has been rarely found among natural products [97]. One new butenolactone, pestalolide (175), and one known phytotoxin, seiridin (176), were isolated from two endophytic fungi Pestalotiopsis sp. PSU-MA69, which was from R. apiculata. The bioactivity analysis showed that compound 175 had weak antimicrobial activity against Candida albicans and Cryptococcus neoformans, with MIC values of 653.06 μM [25]. A new phthalic acid derivative, acrenomone (177), and one new depsidone, acrenomone A (179), together with two known substances, (+)-brefelin A (180) and 5,7-dimethoxy-3,4-dimethyl-3-hydroxyxphthalide (178), were separated from the Acremonium sp. PSU-MA70, which was isolated from R. apiculata [26]. Brefelin A (BFA) is a fungal metabolite that was originally used as an antiviral agent and is now primarily used to study protein transport. It can specifically and reversibly inhibit the Golgi membrane protein protease, prohibiting the linkage of guanine nucleotides to ADP ribosylation factor and, therefore, preventing the transport of proteins from the endoplasmic reticulum (ER) to the Golgi. BFA is also used to inhibit the secretion of cytokine and other proteins as well as enhance the immunostaining of secretory proteins. BFA can activate the neural sheath phosphoric acid cycle, inducing the apoptosis of some tumor cells [98], and it has a weak antibacterial activity against Candida albicans NCPF3153 [26]. Three known substances, macrolides pestalotiollides A and B (181, 182) and 2-epi-herbarumin II (183), were isolated from the fermentation extract of Pestalotiopsis clavispora from R. harrisonii. Bioactivity tests showed that compounds 181–183 had no antitumor effect on leukemia myeloid cells HL-60, hepatoma cell SMMC-7721, lung cancer cell line A-549, breast cancer cell MCF-7, or human colon cancer cell SW480 [43]. In order to effectively control the biosynthesis of Leucostoma persoonii from R. mangle and stimulate the production of cytosporone compounds, a known antibacterial trihydroxy lactone compound, cytosporone E (184), was induced by epigenetic modification [72]. Compound 184 showed a strong anti-infective activity against Plasmodium falciparum with an IC_{50} value of 13 μM. Additionally, compound 184 showed strong inhibitory activity against human lung cancer cell A549, with an IC_{50} value of 437 μM, and a strong inhibitory effect on methicillin-resistant S. aureus, with a MIC value of 72 μM [97] (Figure 9).

3.9. Others

A new difuranylmethane-derived furan fatty acid, flavodonfuran (185), was isolated from the endophytic fungus Flavodon flavus PSU-MA201 from R. apiculata [65, 66]. Xu isolated a new enolic acid compound, pestalotiopin A (187), and two known compounds, 2-anhydromevalonic acid (186) and p-hydroxybenzaldehyde (188), from the Pestalotiopsis sp. of R. mucronata [80]. Rukachaisirikul and coworkers isolated two known compounds, 4-methyl-1-phenyl-2,3-hexanediol (189) and (2R,3R)-4-methyl-1-phenyl-2,3-pentanediol (190), from the Acremonium sp. PSU-MA70 of R. apiculata [26]. One known phenylethanol propionate (191) and a known butanamide compound, butanamide (192), were isolated from the endophytic fungus Phomopsis
sp. PSU-MA214 from R. apiculata [25]. (S)-penipratynolene (193), DNA-damaging active anofinic acid (194), and p-hydroxybenzoic acid methyl ester (195) were isolated from Pestalotiopsis sp. PSU-MA69 of R. apiculata [25] (Figure 10).

4. Conclusion

In this review, we summarize the distribution of frequently occurring fungal endophytes in Rhizophora: 26 genera of mangrove endophytic fungi were isolated from R. stylosa; 27 genera were isolated from R. apiculata; 26 genera were obtained from R. mucronata; 23 genera were isolated from R. mangle; 1 genus was isolated from R. harrisonii and R. annamalayana (namely Pestalotiopsis and Fusarium); and 4 genera of endophytic fungi were isolated from R. racemosa. Until now, no studies have been conducted on R. samoensis. In total, the frequently occurring fungi entophytes in Rhizophora, including 41 families and 64 genera belonging to 23 taxonomic orders of Ascomycota have been reported. Although the biological potential of endophytic fungi from the abovementioned Rhizophora species has not been thoroughly investigated, the core group of fungi can be recognized from different geographic locations. The distribution and molecular phylogeny of the fungi are discussed as well as new findings regarding the chemistry and bioactivity of natural products found in Rhizophora endophytic fungi. The Pestalotiopsis, Penicillium, and Mucor genera of endophytic fungi were identified as the most promising fungal groups in terms of chemical diversity. In particular, the Pestalotiopsis genus constituted 42.56% of the compounds reported, as shown in Figure 11. R. apiculata (34.36%) was observed to be the most investigated host plant, followed by R. stylosa (33.85%) and R. mucronata (23.59%). The chemical identification of metabolites of R. racemosa endophytic fungi has not yet been reported (Figure 11).
Some secondary metabolites with unusual structures were identified in *Rhizophora endophytic* fungi. Novel hybrid sesquiterpene-cyclopaldic acid metabolites with unusual carbon skeletons, pestalotiopes A and B (83, 84), were obtained from the endophytic fungus *Pestalotiopsis* sp. JCM2A4 isolated from the leaves of the Chinese mangrove, *R. mucronata*. Bioassays revealed that antitumor, antimicrobial, and anti-H1N1 activities are the most notable bioactivities of the secondary metabolites from *Rhizophora endophytic* fungi. Some compounds had significant bioactivities, as exemplified by pestal polyol 1 (119), a novel polyketone derivative isolated from *P. clavispora*. Compound 119 has a strong inhibitory effect on mouse lymphoma cell line L5178Y with an IC$_{50}$ value of 4.10 $\mu$M. The indole diterpene alkaloids, rhizovrin A, B, and F (46, 47, 50), isolated from endophytic fungi *Mucor irregularis* QEN-189, have strong inhibitory effects on lung cancer cells A549, with IC$_{50}$ values of 11.5, 6.3, and 9.2 $\mu$M, respectively, as well as inhibitory effects on leukemia myeloid cells HL-60, with IC$_{50}$ values of 9.6, 5.0, and 7.0 $\mu$M, respectively. These findings suggest that *Rhizophora* endophytic fungi offering numerous useful products with medicinal and pathogenic significance have yet to be established.

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