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1. Introduction

In biological studies, the luminescence from fluorescent proteins or luminescent enzymes is widely used for monitoring a change of environment at a cell. Biomolecules used for this probing, such as Green Fluorescence Protein (GFP) or luciferase molecules can respond to the existence of specific molecules or ions and subsequently emit a photon. The detection of a specific molecule can then be confirmed by detecting the emitted photons efficiently with a photon detector. A highly efficient detection of the luminescence is normally essential to a high sensitivity to the specific molecules or ions. An improvement of the sensitivity can upgrade the capability of detection in a low concentration of sample solution. Therefore, there are many efforts to improve the efficiency of the collection of emitted photons and of the optical coupling to the photon detector (Yotter, 2004).

A straightforward approach is to directly detect the luminescence from the whole sample solution in a test tube as shown in Fig. 1 (a). However, to realize high efficiency detection, this method needs a single photon detector with a wide photon-sensitive area, which is ideally larger than the photon-emission area in the test tube. The reason is that it is difficult to image incoherent light such as natural light to a smaller area than the emission area. Here, we are introducing an alternative method, where the luminescent biomolecules are immobilized at an optical fiber end and the luminescence is detected by a photon detector which is optically coupled to the other optical fiber end. The sketch of the optical fiber-based systems is shown in Fig. 1 (b). This method has been investigated for application to a fiberoptic biosensor, which is constructed by immobilizing either an enzyme or an antibody. A review is given in (Arnold, 1991), (Blum & Gautier, 1991).

This method has three merits. The first one is to permit a local detection within the sample solution, because the optical fiber end functions as a needle-like probe in the solution. The second one is that the detection scheme does not require that the photon detector is very close to the sample solution. This feature makes it easier to mount the sensing parts in integrated bioengineering, such as μ-TAS. The third merit is that single photon detectors with a small sensitive area can be used, because the photon-emission area, which is almost identical to the cross section of the core part in the optical fiber, is small. In general, single photon detectors have lower dark counts for smaller sensitive area. Low dark counts are very significant, because it essentially gives the upper limit of the sensitivity of photon detection. Recently, single photon detectors using avalanche photodiodes (APDs) have become widely available with good performance, but their sensitive area is small and has a typical size of 0.1 mm. The
Luminescence detection by the optical fiber-based system allows us to fully use the merits of compactness, high quantum efficiency, and low noise of these APD detectors.

We have built a detection system of bioluminescence at an optical fiber end and investigated the sensitivity of Adenosine triphosphate (ATP) detection by using an APD-type photon detector (Iinuma et al., 2009). ATP is a good indicator of biochemical reaction or life activity, since ATP is considered as the universal currency of biological energy for all living things. Therefore, there are a lot of efforts to develop ATP-sensing techniques for compact and efficient ATP detection (Stanley, 1992), (Andreotti & Berthold, 1999), (Gourine et al., 2005). In particular, high-sensitivity detection of ATP can indicate the existence of microorganisms even in low numbers. Thus, a compact and simple detection system with extremely high sensitivity is very desirable.

One powerful method for highly sensitive ATP detection is to use the chemical reaction involved in the bioluminescence, the luciferin-luciferase reaction (Fraga, 2008). In this reaction, after one ATP molecule and one luciferin molecule are bound to one luciferase molecule, the luciferin molecule is oxidized using the energy of ATP. As a consequence, one photon is emitted during the transition from the excited state to the ground state of the oxidized luciferin molecule bound to the luciferase molecule. The emission of one photon indicates the use of the energy of one ATP molecule. In the method using the luciferin-luciferase reaction, the efficient detection of the bioluminescence is essential for high-sensitivity detection of ATP.

Since the oxidation of luciferin is catalyzed by the enzyme luciferase, the immobilization of luciferase molecules on solid probes of various sizes allows highly sensitive and local measurements of ATP. Three types of immobilization have been used: firstly attachment to the cell surface (Nakamura et al., 2006), secondly attachment to small particles, such as nanoparticles (Konno et al., 2006) and glass beads (Lee et al., 1977), thirdly attachment to extended objects with a size in the centimeter range, such as strips (Blum et al., 1984), (Ribeiro et al., 1998) and films (Worsfold & Nabi, 1986). For the ATP-detection on an intermediate scale below 1 millimeter, a fiberoptic probe employing immobilized luciferase (Blum & Gautier, 1991) as well as microchips (Tanii et al., 2001), (Tsuboi et al., 2007), can be utilized. The detection system of bioluminescence at an optical fiber end can achieve local detection of ATP within the sample solution. Realization of high sensitivity potentially provides the local detection of extremely low number of microorganisms. Thus, it is desirable...
to construct a highly efficient detection system of the bioluminescence at an optical fiber end and to evaluate the detection limit with the system.

The rest of this chapter is organized as follows. In sec. 2, we describe a concept for the construction of the optical fiber-based system for efficient detection of a fluorescence at the optical fiber end. In sec. 3, we show how to optimize the parameters of the optical fiber and the coupling optics so as to realize high photon-collection efficiency. In sec. 4, we describe the application of the constructed detection system to ATP sensing. By immobilizing luciferase molecules at the optical fiber end, the bioluminescence by luciferin-luciferase reaction can be detected using the optical fiber-based system. We evaluated the sensitivity of ATP with this system. Sec. 5 summarizes present results and problems.

2. General concept for construction of the optical fiber-based system

For a luminescence detection system using an optical fiber with a core diameter \( \phi_0 \) and a numerical aperture \( NA_0 \), a collection efficiency of the luminescence \( \eta \) at the optical fiber end depends only on \( NA_0 \) as shown in Fig. 2. From a simple calculation based on the solid angle with a maximum opening angle \( \theta_m \), \( \eta(NA_0) \) can be expressed as,

\[
\eta(NA_0) = \frac{1}{2} \left(1 - \cos \theta_m\right) = \frac{1}{2} \left[1 - \sqrt{1 - \left(\frac{NA_0}{n_{w}}\right)^2}\right],
\]  

where \( n_w \) is the refraction index of the substance surrounding the optical fiber end. In immersing the optical fiber end into water, its value should be identical to the value of water, which is about 1.33. Fig. 3 shows the calculated values of the collection efficiency \( \eta(NA_0) \) as a function of \( NA_0 \) using Eq. (1) at \( n_{w} = 1.33 \). One can easily see that \( \eta(NA_0) \) increases with \( NA_0 \).

In the following, let us consider the situation where the other optical fiber end is optically coupled to a photon detector with a circular sensitive area having a diameter \( \phi_3 \) and a numerical aperture \( NA_3 \). The coupling efficiency \( \epsilon \) between the optical fiber end and the photon detector depends on \( \phi_0, NA_0 \) of the optical fiber and \( \phi_3, NA_3 \) of the photon detector

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used. Since the number of emitted photons is proportional to the square of $\phi_0$ and $\eta(NA_0)$ increases with $NA_0$ as expressed by Eq. (1), the number of transmitted photons to the other optical fiber end is proportional to the square of $\phi_0$ and $\eta(NA_0)$. On the other hand, the coupling efficiency $\epsilon$ generally decreases as $\phi_0$ or $NA_0$ increases. Thus, we can define the following formula for a figure of merit (FOM) and optimize $\phi_0$, $NA_0$ and parameters of the coupling optics $x_i$ to maximize this FOM:

$$FOM = \phi_0^2 \cdot \eta(NA_0) \cdot \epsilon(\phi_0, NA_0, x_i; \phi_3, NA_3)$$  \hspace{1cm} (2)

It should be noted that $\epsilon$ can ideally be 100% under the conditions of $\phi_0 \leq \phi_3$ and $NA_0 \leq NA_3$. In many cases, the condition $NA_0 \leq NA_3$ is satisfied when using typical photon detectors. Thus, under the condition of $NA_0 \leq NA_3$, we can classify two cases: case (1) is $\phi_0 \leq \phi_3$ and case (2) is $\phi_0 > \phi_3$. In case (1), $\epsilon$ is constant and can ideally be 100% and the total detection efficiency is limited only by $\eta(NA_0)$. Therefore, optimization of the coupling optics is not necessary. The conditions $\phi_0 = \phi_3$ and $NA_0 = NA_3$ both maximize the FOM and the sensitivity becomes highest. In case (2), however, the optimization of $\phi_0$, $NA_0$, and the parameters $x_i$ for a design of the coupling optics are necessary for given values of $NA_3$ and $\phi_3$, because $\epsilon$ decreases as $\phi_0$ or $NA_0$ increases.

3. Design of coupling optics

3.1 Photon detectors

Photon detectors generally have two significant factors contributing to the sensitivity of detection for weak light: the efficiency and the dark counts of the photodetector. A cooled APD which can detect for single photons is mostly used because of its very low dark counts. The sensitive area must be very small for realizing a large reduction of the dark counts, but the quantum efficiency is several times larger than that of a photomultiplier tube (PMT). Furthermore, the APD has the useful characteristics of compactness, easy operation, and durability in comparison with a PMT. To construct an optical fiber-based system with high...
sensitivity, we chose an APD-type photon counting module (SPCM-AQR-14) manufactured by Perkin Elmer Co. Ltd., which has a quantum efficiency $\eta_{qe}$ of 55% at 550 nm and dark counts of about 100 s$^{-1}$. The APD has a circular sensitive area, where $\phi_3$ is 0.175 mm and $NA_3$ is 0.78, as calculated from the geometrical structure between the sensitive area and the photon detection window. If we use an optical fiber with $\phi_0 > \phi_3$, it is necessary to optimize $\phi_0$, $NA_0$, and to design the coupling optics for maximal sensitivity.

3.2 Design concept and procedure

We can consider the coupling optics between the optical fiber end and the APD as an optical system imaging a light source with $NA_0$ and $\phi_0$ onto the APD with $NA_3$ and $\phi_3$. The basic design of the coupling optics is shown in Fig. 4.

Among the parameters shown in Fig. 4, we firstly determine the parameters of optical components, the focal length $f_1$ and the numerical aperture $NA_1$ of the first lens, the focal length $f_2$ and the numerical aperture $NA_2$ of the second lens, and the distance $d_{12}$ between the first lens and the second lens. The second lens is selected among available lenses to make $NA_2$ as large as possible while staying below $NA_3$. We also select the first lens among available lenses to make $NA_1$ as large as possible while staying below $NA_2$. As the result, $f_1$, $NA_1$, $f_2$, $NA_2$ were determined as shown in Table 1.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>focal length</td>
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</tr>
<tr>
<td>$NA_1$</td>
<td>0.50</td>
</tr>
<tr>
<td>$NA_2$</td>
<td>0.68</td>
</tr>
<tr>
<td>$\phi_0$</td>
<td>8.0 mm</td>
</tr>
<tr>
<td>$\phi_3$</td>
<td>3.1 mm</td>
</tr>
<tr>
<td>$\phi_0$</td>
<td>8.0 mm</td>
</tr>
<tr>
<td>$\phi_3$</td>
<td>3.1 mm</td>
</tr>
</tbody>
</table>

Table 1. Parameters of optical components

Taking into account of geometrical structure of lens mounts and fixing both of the first and second lens, the distance $d_{12}$ should be longer than 3 mm. In this case, we determined $d_{12} = 5$ mm. The remaining parameters in this optical system were $NA_0$, $\phi_0$, the distance $d_1$ between the fiber end and the first lens, and the distance $d_2$ between the second lens and the APD. These parameters can be determined in two steps as follows. In the first step, $NA_0$ and $\phi_0$ are optimized to maximize the FOM expressed as Eq. (2) under the conditions $f_1 = d_1$ and $f_2 = d_2$. In the second step, $d_1$ and $d_2$ are optimized to maximize $\epsilon(d_1, d_2, \phi_0, NA_0)$ for given values of $NA_0$ and $\phi_0$ obtained in the first step.
3.3 Determination of parameters in the optical system

The optimization procedure requires specific values of \( \eta(NA_0) \) and \( e(NA_0, \phi_0) \) for obtaining the FOM. Since the values of \( \eta(NA_0) \) were given from Eq. (1) as shown in Fig. 3, it is necessary to calculate the values of \( e(NA_0, \phi_0) \) at \( d_1 = f_1 \) and \( d_2 = f_2 \). The simple method for obtaining the efficiency is a statistical simulation by ray tracing.

The optical fiber end can be considered as a light source with \( \phi_0 \) and \( NA_0 \). To calculate the efficiency, an event of light emission is randomly generated at an arbitrary position within \( \phi_0 \) and at an arbitrary direction within \( NA_0 \). Subsequently, the final state of light at the APD sensitive area is calculated by the transformation of the initial state based on ray tracing. We repeat a procedure containing the generation of one event in the light emission and the subsequent transformation to the final state at the APD via intermediate states at the first lens and the second lens, and count the number of the events, where the conditions at the first lens \( NA_1, \phi_1 \), at the second lens \( NA_2, \phi_2 \), and at the APD \( NA_3 \) and \( \phi_3 \) are all fulfilled. The efficiency can be obtained as the ratio of number of counts to the total number of event generation.

The propagation of light can be described with matrix formalism in paraxial optics (Yariv, 1997). The light at the initial state \( (r_i, r'_i) \), where \( r_i \) is a distance from an optical axis of optics and \( r'_i \) is a slope of the light direction, can be transferred by the following matrices, for example,

\[
M_{\text{free}} = \begin{bmatrix} 1 & d_0 & 1 \\ 0 & 1 \end{bmatrix}
\]

\[
M_{\text{lens}} = \begin{bmatrix} 1 & 0 & 1 \\ -\frac{1}{f_1} & 1 \end{bmatrix}
\]

where \( M_{\text{free}} \) is the transfer matrix of free-space propagation far from the distance \( d \) and \( M_{\text{lens}} \) is the transfer matrix of thin lens with the focal length \( f \). Any state of light expressed as the vector form \( (r, r') \) can be transferred by any combination of the transfer matrices. Therefore, the transfer matrix \( M_{\text{coupling}} \) describing the optical system shown in Fig. 4 can be expressed as follows,

\[
M_{\text{coupling}} = \begin{bmatrix} 1 & d_2 & 0 & 1 \\ 0 & 1 & 0 & 1 \\ -\frac{1}{f_2} & 1 & 1 & d_1 \end{bmatrix}
\]

\( M_{\text{coupling}} \) can transfer the initial state \( (r_i, r'_i) \) at the optical fiber end to the final state \( (r_f, r'_f) \) at the APD sensitive area.

Thus, the values of \( e \) were obtained by the statistical method with the transfer matrix \( M_{\text{coupling}} \), where random numbers for setting initial states were generated with the software package based on algorithm of Mersenne Twister (Matsumoto & Nishimura, 1998). The calculated results are shown as a function of \( NA_0 \) in Fig. 5.

Fig. 5 shows that \( e \) is decreasing with increasing \( NA_0 \) and \( \phi_0 \). As a consequence, \( e \) becomes 100% at \( \phi_0 = 0.4 \) mm and \( NA_0 \leq 0.25 \). However, the reduction of \( \phi_0 \) and \( NA_0 \) makes both \( \eta \)
Fig. 5. Calculated values of coupling efficiency $\epsilon$ as a function of $N A_0$. The solid circles indicate the values at $\phi_0 = 0.4$ mm, the solid triangles are the values at $\phi_0 = 0.6$ mm, and the solid inverse triangles are the values at $\phi_0 = 0.8$ mm.

Fig. 6. Calculated values of FOM as a function of $N A_0$. The solid circles indicate the values at $\phi_0 = 0.4$ mm, the solid triangles are the values at $\phi_0 = 0.6$ mm, and the solid inverse triangles are the values at $\phi_0 = 0.8$ mm.

and the number of emitted photons smaller. Therefore, the calculation of the FOM is necessary for the optimization of $N A_0$ and $\phi_0$.

The plot of the FOM as a function of $N A_0$ is shown in Fig. 6. One can easily see that the FOM is maximal at $N A_0 = 3.0$. In addition, the value of the FOM for $\phi_0 = 0.6$ mm is almost same as the one for $\phi_0 = 0.8$ mm. This means that the FOM is saturated for larger diameters than $\phi_0 = 0.6$ mm because the size of the transferred image at the APD is larger than the APD sensitive area.
Optical fibers with $NA_0 = 3.0$ are not easily obtainable, whereas optical fibers with $NA_0 = 2.5$ and $NA_0 = 3.7$ are readily available. Fig. 6 shows that the slope above $NA_0 = 3.0$ is flatter than the one below. In the upper part, the misalignment or imperfection of the optical system has less influence on the coupling efficiency. Therefore, we selected $\phi_0 = 0.6 \text{ mm}$ and $NA_0 = 3.7$.

![Graph showing coupling efficiency vs. $d_1$ and $d_2$.]

In a second step, $d_1$ and $d_2$ were optimized to maximize $\epsilon(d_1, d_2)$ for $NA_0 = 0.37$ and $\phi_0 = 0.6 \text{ mm}$. Fig. 7 shows the calculated values of $\epsilon$ as functions of $d_1$ and $d_2$. From the peak of $\epsilon$ in Fig. 7, the results of $d_1 = 11.6 \text{ mm}$ and $d_2 = 2.7 \text{ mm}$ were obtained for $NA_0 = 0.37$ and $\phi_0 = 0.6$. These parameters provide the maximum $\epsilon(d_1, d_2)$ of 33.33 %, $\eta(NA_0)$ of 1.95 % and $\text{FOM} \times 100$ is 0.234 which value is higher than the maximum in Fig. 6.

4. Application to the ATP sensing system

4.1 Luciferin-luciferase reaction

Bioluminescence in living organisms, such as fireflies and some marine bacteria, typically occurs due to the optical transition from the excited state to the ground state of oxidized luciferin molecules produced by the luciferin-luciferase reaction under the catalytic activity of luciferase molecules. This reaction can be expressed by the following sequence of reaction steps:

$$
\begin{align*}
E + LH_2 + S & \rightleftharpoons C + PP_i \\
C + O_2 & \rightarrow E + P^* + AMP + CO_2 \\
P^* & \rightarrow P + \gamma 
\end{align*}
$$

where $E$ indicates luciferase, $LH_2$ luciferin, $PP_i$ pyrophoric acid, $C$ is an enzyme-substrate compound $E \cdot LH_2$-AMP, AMP adenosine monophosphate, $P$ oxidized luciferin, and $\gamma$ a photon (DeLuca, 1976). The emission of one photon at the position of luciferase molecule indicates the use of the energy of one ATP molecule.

The immobilization of luciferase molecules at the optical fiber end enables us to sense the presence of ATP around the fiber end using single photon detection. For this purpose, we used a compound protein containing a silica-binding protein (SBP) molecule and a luciferase
molecule (SBP-luciferase), which were recently synthesized by Taniguchi and co-workers (Taniguchi et al., 2007). This protein makes it possible to immobilize a luciferase molecule on the optical fiber end via a SBP molecule while retaining its activity. The spectrum of the emitted photons shows a central wavelength of 550 nm and a width of about 100 nm (Denburg et al., 1969), (Ugarova & Brovko, 2002). Since the APD module has the large quantum efficiency for the photons from the luciferin-luciferase reaction, the APD photon counting module is suitable for ATP sensing.

4.2 Reaction-diffusion differential equation

Fig. 8 shows the enzyme reaction which describes the sequence of reactions.

\[ s + e \xrightarrow{k^+} c \xrightarrow{h_1} p + e + n, \]

\( s \): ATP (S)  
\( e \): luciferase (E)  
\( c \): oxyluciferin (P)  
\( n \): photon (γ)  
\( h_1 \): reaction constant

Fig. 8. Enzyme reaction describing luciferin-luciferase reaction

Here, \( s, e, c, p, n, γ \) in Fig. 8 represents a concentration of ATP, luciferase, \( E \cdot LH_2\cdotAMP \), oxidized luciferin, and emitted photon, respectively. In addition, \( k^+, k^- \) are kinetic constants for equilibrium and \( h_1 \) is a reaction constant.

In a solution containing nonlocalized homogeneously dispersed luciferase and ATP, the Michaelis-Menten theory can be simply applied to the above reaction. In the presence of enough luciferin molecules in the solution, a rate of emitted photons at steady state \( v_γ \) can be expressed as the Michaelis-Menten formula,

\[ v_γ = \frac{V_0 S}{S + K_M}, \]  

(3)

where \( V_0 \) is a maximum reaction rate which is equivalent to a product of the number of luciferase molecule and \( h_1 \), \( K_M \) is the Michaels constant expressed as \( K_M = \frac{k^- + h_1}{k^+} \) and \( S \) is the ATP concentration.

In the fiber-based system for sensing dispersed ATP molecules, on the other hands, an ATP-flow generated by a gradient of ATP concentration around the luciferase-terminated fiber end carries ATP molecules to the vicinity of immobilized SBP-luciferase molecules. The ATP molecule is bound to the immobilized SBP-luciferase molecule near them and subsequently contributes the luciferin-luciferase reaction at this fiber end. To calculate the rate of emitted photons, therefore, it is necessary to consider not only a reaction rate but also an ATP diffusion rate.

The series of reaction-diffusion equations describing the enzyme reaction shown in Fig. 8 can be expressed as,
\[
\frac{ds}{dt} = D \left( \frac{\partial^2 s}{\partial x^2} + \frac{\partial^2 s}{\partial y^2} \right) + R(s, e, c)
\]
\[
\frac{de}{dt} = -k_+ se + k_- c + h_1 c
\]
\[
\frac{dc}{dt} = k_+ se - k_- c - h_1 c
\]
\[
\frac{dn}{dt} = h_1 c,
\]
(4)

where \(D\) is a diffusion constant of ATP in water and \(R(s, e, c)\) is expressed as follows.

\[
R(s, e, c) \equiv \begin{cases} 
-k_+ se + k_- c & (x, y \in \Gamma_{fiber}) \\
0 & (otherwise)
\end{cases}
\]

Fig. 9. Coordinate system of \(x\) and \(y\) around the fiber end.

Fig. 9 shows the definition of the coordinate system of \(x\) and \(y\). Here, it should be noted that \(s(x, y, t)\) is a function of \(x, y, t\) and \(e(t)\) and \(c(t)\) are functions of only \(t\). \(\Gamma_{fiber}\) is defined as an area where the reaction occurs and equivalent to the core part \(\phi_0\) in the optical fiber. In this coordinate system, it can be represented as the interval of \(x_2 \leq x \leq x_3\) with \(y = y_h\). The both intervals of \(x_1 \leq x \leq x_2\) and \(x_3 \leq x \leq x_4\) with \(y = y_h\) represent the parts of clad in the optical fiber.

By dividing the space around the fiber end into finite spatial steps and also dividing the time into finite time steps, we can numerically solve the series of reaction-diffusion equation Eq. (4) under the boundary conditions presented in Table 2. Here, \(s_0\) in Table 2 is an initial ATP concentration that should be uniform into the whole solution before starting the reaction. The numerical solutions can be given as time evolution of spatial distribution of concentration \(s(x_i, y_i, t_i)\), \(e(t_i)\), and \(c(t_i)\) and the rate of emitted photons can be obtained from \(h_1 c(t_i)\). The peak of the photon-emission rate in time corresponds to the reaction rate given by the Michaelis-Menten formula.

In order to check the possibility that the emission rates are limited by the diffusion rate of ATP, we numerically obtained the peak values of the photon-emission rate at various ATP concentration and compared to the reaction rate calculated by Michaelis-Menten formula. To obtain numerical solutions at each time step, spatial segmented equations derived from Eq.
High-Sensitivity Detection of Bioluminescence at an Optical Fiber End for an ATP Sensor

Table 2. Boundary conditions for solving Eq. (4). $s_0$ is the initial ATP concentration.

<table>
<thead>
<tr>
<th>region</th>
<th>$x = 0$</th>
<th>$0 \leq x &lt; x_1$</th>
<th>$x = x_1$</th>
<th>$x_1 \leq x &lt; x_2$</th>
<th>$x_2 \leq x \leq x_3$</th>
</tr>
</thead>
<tbody>
<tr>
<td>condition</td>
<td>$s = s_0$</td>
<td>$y = 0$</td>
<td>$0 \leq y &lt; y_h$</td>
<td>$y = y_h$</td>
<td>$y = y_h$</td>
</tr>
<tr>
<td>region</td>
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<td>$x_4 &lt; x \leq x_{max}$</td>
<td>$x = x_{max}$</td>
<td>$y = y_{max}$</td>
</tr>
<tr>
<td>condition</td>
<td>$\frac{dx}{dy}$</td>
<td>$y = y_h$</td>
<td>$0 \leq y &lt; y_h$</td>
<td>$y = 0$</td>
<td>$s = s_0$</td>
</tr>
</tbody>
</table>

Fig. 10. Comparison of the results calculated from numerical solutions of reaction-diffusion equations to the results of Michaelis-Menten formula. Closed triangles indicate values of ATP diffusion process using typical values of kinetic constants, closed circles are values given by Michaelis-Menten formula.

(4) were solved in terms of time using the software package of ordinary differential equations DASKR (Brown et al., 1998), where the ATP diffusion constant was $D = 0.5 \times 10^{-5}$ cm$^2$/s (Aflao & DeLuca, 1987). The geometrical parameters were $x_{max} = 1.4$ mm, $y_{max} = 2$ mm, $y_h = 1$ mm, $x_1 = 0.38$ mm, $x_4 = 1.02$ mm, the kinetic constants were $k_+ = 20000$ M$^{-1}$s$^{-1}$, which was estimated from the typical buildup time of 0.3 s (DeLuca & McElory, 1974), and $k_- = 0.515$ s$^{-1}$, which was calculated with the relational form $K_M \cdot k_- = h_1$. As other parameters, we used $h_1 = 0.125$ s$^{-1}$ (Branchini et al., 2001), $K_M = 3.2 \times 10^{-5}$ M, and the surface density of luciferase molecule $c_0 = 9.03 \times 10^{10}$ mm$^{-2}$ (Taniguchi et al., 2007), which were also used for Michaelis-Menten formula. The Michaelis constants $K_M = 3.2 \times 10^{-5}$ M was obtained from data analysis of counts of detected photons, which describes in Sec. 4.

Fig. 10 shows the results of two kinds of calculations. The closed triangles indicate the values obtained from numerical solutions of Eq. (4), whereas the closed circles are the values deduced from Michaelis-Menten formula. The results of ATP diffusion process have good agreements with ones given by the Michaelis-Menten theory. Therefore, we can conclude that the ATP diffusion is not a rate-limiting process for the present rate of the chemical reaction.
4.3 Measurement of the sensitivity

4.3.1 Immobilization of luciferase

Before immobilizing the luciferase molecules, we cut optical fiber and cleaned the cut surface with ethanol and Tris buffer (0.25 mM Tris-HCl with 0.15 M NaCl). After cleaning, the surface was immersed in a solution of SBP-luciferase and was left at a temperature of 3°C to 6°C for a period of about two hours.

4.3.2 Sample solutions

The samples were a 1:4:4:31 mixture of 20 mM D-luciferin solution, Tris buffer solution (250 mM Tris-HCl mixed with 50 mM MgCl₂), ATP solution, and distilled water. Several solutions of ATP with different ATP concentrations were made by diluting the ATP standard in ATP Bioluminescence Assay Kit CLS II manufactured by Roche Co. Ltd. Thus, a series of sample solutions with different ATP concentrations were prepared in advance. An additional sample without ATP was also produced by mixing distilled water instead of the ATP solution. This sample was measured in order to obtain a background before the ATP measurements.

4.3.3 Data taking system

TTL pulses outputted from the APD photon counting module were counted by a PC card installed in a personal computer (PC). The number of pulses occurring during 10 s were recorded every 10 s by the PC.

4.3.4 Results

The time dependence of photon counts per 10-s interval were measured after the luciferase-terminated end were immersed in the sample solutions with various ATP concentration. The results for 100 µl solution from $1.65 \times 10^{-4}$ M to $1.65 \times 10^{-9}$ M are shown in Fig. 11.

The photon counts increase and reach a maximum at about 150 s after immersion. Then, they decrease very slowly to the background level. Therefore, for obtaining high sensitivity, it is practical to continue the measurement for about 300 s after the luciferase-terminated end is immersed in the solution. A background level of approximately 120 s⁻¹ was determined as an average of counts for background data. It was found to correspond well to the dark counts of the photon counting module.

<table>
<thead>
<tr>
<th>ATP concentration</th>
<th>integrated counts</th>
<th>BG-subtracted counts</th>
<th>statistical errors</th>
</tr>
</thead>
<tbody>
<tr>
<td>no ATP (background)</td>
<td>34490</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$1.65 \times 10^{-11}$ M</td>
<td>34664</td>
<td>174</td>
<td>262.97</td>
</tr>
<tr>
<td>$1.65 \times 10^{-10}$ M</td>
<td>35346</td>
<td>856</td>
<td>264.27</td>
</tr>
<tr>
<td>$1.65 \times 10^{-9}$ M</td>
<td>46770</td>
<td>12280</td>
<td>283.06</td>
</tr>
<tr>
<td>$1.65 \times 10^{-8}$ M</td>
<td>139300</td>
<td>104810</td>
<td>416.88</td>
</tr>
</tbody>
</table>

Table 3. Integrated counts for ATP sample solutions

Table 3 presents integrated counts of detected photons during 300 s from the origin of time. Statistical errors are estimated as one standard deviation assuming Poisson distribution. From Table 3, the sensitivity in this optical fiber-based system is limited to $1.65 \times 10^{-10}$ M, which
corresponds to a number of ATP molecules of about $10^{-14}$ mol in the 100 $\mu\text{l}$ solution. We also ascertained that our system is sensitive to the $10^{-10}$ M level even with a 10$\mu\text{l}$ solution, in which the absolute ATP concentration is $10^{-15}$ mol.

In order to check the ATP concentration dependence of the photon counting rate at maximum, the average of counts in six 10-s intervals around the time at which the counting rate became maximal was calculated for each ATP concentration. The results are indicated by solid circles in Fig. 12. This figure shows that the lower limit of the sensitivity is $1.65 \times 10^{-9}$ M taking into account of statistical errors. If the statistical errors reduce to half of the present ones, which means that statistics at each point becomes four times as large as 60-s interval, the detection of $1.65 \times 10^{-10}$ M also becomes feasible. This is consistent with the results presented in Table 3.

The analysis of fitting data points in Fig. 12 provided the Michaelis constant of $3.2 \times 10^{-5}$ M.

4.3.5 Discussion

In Fig. 12, the open squares show predictions of the counting rate estimated from the results of numerical solutions shown in Fig. 10 with the total detection efficiency in our detection system, which can be expressed as $\epsilon_{\text{total}} = \eta \cdot \epsilon \cdot \eta_{\text{fgr}}$. Its value was calculated to be 0.00354 for photons with the wavelength 550 nm. Comparing the experimental data with the predictions, there are disagreements amounting to roughly one order of magnitude.

The results suggest the ATP diffusion does not limit the detection limit. Therefore, the possible reasons for the discrepancies may be the following.

(1) The rate of the chemical reaction per luciferase molecule itself becomes small.
(2) The number of immobilized active molecules is lower than expected.

We have also confirmed that the buildup time in Fig. 11 is about 30 times longer than that in a solution containing only nonlocalized homogeneously dispersed SBP-luciferase.
ATP concentration

Counting rate

$[\text{M}]$ $[\text{counts/s}]$

$0.1 \times 10^0$ $1 \times 10^0$

$3 \times 10^2$ $5 \times 10^2$

$7 \times 10^3$ $9 \times 10^3$

$1 \times 10^5$ $3 \times 10^5$

$5 \times 10^6$ $7 \times 10^6$

$9 \times 10^7$ $1 \times 10^8$

$3 \times 10^9$ $5 \times 10^9$

$7 \times 10^{10}$ $9 \times 10^{10}$

$1 \times 10^{12}$

$1 \times 10^{14}$

Fig. 12. Measured photon counting rate as a function of ATP concentration. Solid line is a curve obtained by fitting data with the Michaelis-Menten formula. Open squares are values estimated from the results of numerical solutions shown in Fig. 10 with total detection efficiency $\epsilon_{\text{total}} = 0.00354$.

Fig. 12. Measured photon counting rate as a function of ATP concentration. Solid line is a curve obtained by fitting data with the Michaelis-Menten formula. Open squares are values estimated from the results of numerical solutions shown in Fig. 10 with total detection efficiency $\epsilon_{\text{total}} = 0.00354$.

molecules. There are also other experimental results claiming the decrease in activity of immobilized luciferase molecules (Nishiyama et al., 2008), (Tanaka et al., 2011). The reason of this discrepancy is still an open question. These results suggest that it will be necessary to study the reaction at the end of the optical fiber in more detail in order to improve the sensitivity.

5. Summary

We introduced a method of high-sensitivity detection of bioluminescence at an optical fiber end for an ATP sensor as an efficient alternative to direct detection of bioluminescence from a sample solution. For efficiently detection of the bioluminescence, we have constructed an optical fiber-based system, where the luciferase molecules are immobilized on the optical fiber end and the other end is optically coupled to an APD-type photon counting module. We have evaluated the sensitivity of the ATP detection of this system and found that the optimization of the optical coupling system and the use of the SBP-luciferase molecules provide the detection limit of $10^{-10} \text{M}$, which allows us to detect the absolute ATP concentration of $10^{-15} \text{mol}$ with a $10 \mu\ell$ solution. To improve the sensitivity, it is necessary to study the details of the reaction at luciferase-terminated end of the optical fiber.

6. Acknowledgements

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7. References


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This book presents a comprehensive account of recent advances and researches in fiber optic sensor technology. It consists of 21 chapters encompassing the recent progress in the subject, basic principles of various sensor types, their applications in structural health monitoring and the measurement of various physical, chemical and biological parameters. It also highlights the development of fiber optic sensors, their applications by providing various new methods for sensing and systems, and describing recent developments in fiber Bragg grating, tapered optical fiber, polymer optical fiber, long period fiber grating, reflectometry and interferometry based sensors. Edited by three scientists with a wide knowledge of the field and the community, the book brings together leading academics and practitioners in a comprehensive and incisive treatment of the subject. This is an essential reference for researchers working and teaching in optical fiber sensor technology, and for industrial users who need to be aware of current developments and new areas in optical fiber sensor devices.

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